

Medical Policy



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Title: Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes

Professional

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Populations	Interventions	Comparators	Outcomes
Individuals: <ul style="list-style-type: none"> Who are suspected of attenuated FAP, MAP, and Lynch syndrome, or are at-risk relatives of patients with FAP 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>APC</i> 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity

Populations	Interventions	Comparators	Outcomes
Individuals: <ul style="list-style-type: none"> Who are suspected of attenuated FAP, MAP, and Lynch syndrome 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>MUTYH</i> after a negative <i>APC</i> test result 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> Who are suspected of attenuated FAP, MAP, and Lynch syndrome; CRC; or endometrial cancer and first-degree relative with Lynch 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for MMR genes 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> Who are at-risk relatives of patients with Lynch or family history meeting appropriate criteria, but do not have CRC 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for MMR genes 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> Who warrant Lynch testing, screen negative on MMR testing, but positive for MSI and lack MSH2 protein expression 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>EPCAM</i> variants 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> With CRC in whom MLH1 protein is not expressed on immunohistochemical analysis 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>BRAFV600E</i> or <i>MLH1</i> promoter methylation 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> Who are suspected of juvenile polyposis syndrome or are at-risk relatives of patients suspected of or diagnosed with JPS 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>SMAD4</i> and <i>BMPR1A</i> genes 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity
Individuals: <ul style="list-style-type: none"> Who are suspected of Peutz-Jeghers syndrome or are at-risk relatives of patients suspected of or diagnosed with PJS 	Interventions of interest are: <ul style="list-style-type: none"> Genetic testing for <i>STK11</i> gene 	Comparators of interest are: <ul style="list-style-type: none"> No genetic testing 	Relevant outcomes include: <ul style="list-style-type: none"> Overall survival Disease-specific survival Test accuracy Test validity

CRC: colorectal cancer; FAP: familial adenomatous polyposis; MAP: *MUTYH*-associated polyposis; MMR: mismatch repair; MSI: microsatellite instability.

DESCRIPTION

Genetic testing is available for both those with and those at risk for various types of hereditary cancer. This policy evaluates genetic testing for hereditary colorectal cancer and polyposis syndromes, including familial adenomatous polyposis (FAP), Lynch syndrome (formerly known as hereditary nonpolyposis colorectal cancer), *MUTYH*-associated polyposis (MAP), Lynch syndrome-related endometrial cancer, and Peutz-Jeghers syndrome.

OBJECTIVE

The objective of this policy is to assess whether the use of genetic testing improves the net health outcome in patients with Lynch syndrome and other inherited colon cancer syndromes.

BACKGROUND

Hereditary Colorectal Cancers

There are currently 2 well-defined types of hereditary colorectal cancer, familial adenomatous polyposis and Lynch syndrome (formerly hereditary nonpolyposis colorectal cancer [CRC]). Lynch syndrome has been implicated in some endometrial cancers as well.

FAP and Associated Variants

FAP typically develops by age 16 years and can be identified by the appearance of hundreds to thousands of characteristic, precancerous colon polyps. If left untreated, all affected individuals will go on to develop colorectal cancer. The mean age of colon cancer diagnosis in untreated individuals is 39 years. FAP accounts for about 1% of colorectal cancer and may also be associated with osteomas of the jaw, skull, and limbs; sebaceous cysts; and pigmented spots on the retina referred to as congenital hypertrophy of the retinal pigment epithelium (CHRPE). FAP associated with these collective extraintestinal manifestations is sometimes referred to as Gardner syndrome. FAP may also be associated with central nervous system (CNS) tumors, referred to as Turcot syndrome.

Germline variants in the adenomatous polyposis coli (*APC*) gene, located on chromosome 5, are responsible for FAP and are inherited in an autosomal dominant manner. Variants in the *APC* gene result in altered protein length in about 80% to 85% of cases of FAP. A specific *APC* gene variant (I1307K) has been found in subjects of Ashkenazi Jewish descent that may explain a portion of the familial colorectal cancer occurring in this population.

A subset of FAP patients may have attenuated FAP (AFAP), typically characterized by fewer than 100 cumulative colorectal adenomas occurring later in life than in classical FAP, colorectal cancer occurring at an average age of 50-55 years, but a high lifetime risk of colorectal cancer of about 70% by age 80. The risk of extra-intestinal cancer is lower compared to classical FAP but still high at an estimated cumulative lifetime risk of 38% compared to the general population.¹ Only 30% or fewer of AFAP patients have *APC* variants; some of these patients instead have variants in the *MUTYH* (formerly *MYH*) gene and are then diagnosed with *MUTYH*-associated polyposis (MAP). MAP occurs with a frequency approximately equal to FAP, with some variability among prevalence estimates for both. While clinical features of MAP are similar to FAP or AFAP, a strong multigenerational family history of polyposis is absent. Biallelic *MUTYH* variants are associated with a cumulative colorectal cancer risk of about 80% by age 70, whereas monoallelic *MUTYH* variant-associated risk of colorectal cancer appears to be relatively minimal, although still under debate.² Thus, inheritance for high-risk colorectal cancer predisposition is autosomal recessive in contrast to FAP. When relatively few (ie, between 10 and 99) adenomas are present and family history is unavailable, the differential diagnosis may include both MAP and Lynch syndrome; genetic testing in this situation could include *APC*, *MUTYH* if *APC* is negative for variants, and screening for variants associated with Lynch syndrome.

It is important to distinguish among classical FAP, attenuated FAP, and MAP (mono- or biallelic) by genetic analysis because recommendations for patient surveillance and cancer prevention vary according to the syndrome.³

Testing

Genetic testing for APC variants may be considered in the following situations:

- Patients at high risk such as those with a family member who tested positive for FAP and have a known *APC* variant.
- Patients undergoing differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome. These patients do not meet the clinical diagnostic criteria for classical FAP and have few adenomatous colonic polyps.
- To confirm FAP in patients with colon cancer with a clinical picture or family history consistent with classical FAP.

Lynch Syndrome

Lynch syndrome is an inherited disorder that results in a higher predisposition to CRC and other malignancies including endometrial and gastric cancer. Lynch syndrome is estimated to account for 3% to 5% of all CRC. People with Lynch syndrome have a 70% to 80% lifetime risk of developing any type of cancer.^{4,5} However the risk varies by genotype. It occurs as a result of germline variant in the mismatch repair (MMR) genes that include *MLH1*, *MSH2*, *MSH6*, and *PMS2*. In approximately 80% of cases, the variants are located in the *MLH1* and *MSH2* genes, while 10% to 12% of variants are located in the *MSH6* gene and 2% to 3% in the *PMS2* gene. Also, variants in 3 additional genes (*MLH3*, *PMS1*, *EXO1*) have also been implicated with Lynch Syndrome. Notably, in individuals meeting the various clinical criteria for Lynch syndrome, 50% individuals have a variant in the *MLH1*, *MSH2*, *MSH6*, and *PMS2* genes. The lifetime risk of CRC is nearly 80% in individuals carrying a variant in one of these genes.

Testing

Testing approach to identify patients with Lynch syndrome is summarized next. Preliminary screening of tumor tissue does not identify MMR gene variants but is used to guide subsequent diagnostic testing via DNA analysis for specific variants. Genetic testing or DNA analysis (gene sequencing, deletion and duplication testing) for the MMR genes involves assessment for *MLH1*, *MSH2*, *MSH6*, and *PMS2* variants. The following are 3 testing strategies.

1. Microsatellite instability (MSI) testing (phenotype): Individuals with high MSI either proceed to genetic testing for *MLH1*, *MSH2*, *MSH6*, and *PMS2* or to immunohistochemical (IHC) testing.
2. IHC testing (phenotype): Individuals with negative staining would proceed to genetic testing for *MLH1*, *MSH2*, *MSH6*, and *PMS2*.
3. Modification strategy: Tumor tissue of patients with negative staining for *MLH1* on IHC is tested for the *BRAFV600E* variant to determine methylation status. If the *BRAF* variant is not detected, the individual receives *MLH1* DNA analysis.

The phenotype tests used to identify individuals with who may be at a high-risk of Lynch syndrome are explained next. The first screening test measures MSI. As a result of variance in the MMR gene family, the MMR protein is either absent or deficient, resulting in an inability to correct DNA replication errors causing MSI. Approximately 80% to 90% of Lynch syndrome CRC tumors have MSI. The National Cancer Institute has recommended screening for 5 markers detect MSI (Bethesda markers). MSI detection in 2 of these markers is considered a positive result or "high probability of MSI".⁶

The second phenotype screening test is IHC, which involves staining of tumor tissue for the presence of 4 MMR proteins (MLH1, MSH2, MSH6, PMS2). The absence of one or more protein is considered abnormal.

BRAF testing is an optional screening method that may be used in conjunction with IHC testing for *MLH1* to improve efficiency. A methylation analysis of the *MLH1* gene can largely substitute for *BRAF* testing, or be used in combination to improve efficiency slightly.

Both MSI and IHC have a 5% to 10% false-negative rate. MSI testing performance depends on the specific MMR variant. MSI screening has a sensitivity of about 89% for *MLH1* and *MSH2* and 77% for *MSH6* and a specificity of about 90% for each. The specificity of MSI testing is low because approximately 10% of sporadic CRCs are MSI-positive due to somatic hypermethylation of the *MLH1* promoter. Additionally, some tumors positive for *MSH6* variants are associated with the MSI-low phenotype rather than MSI-high; thus MSI-low should not be a criterion against proceeding to MMR variant testing.^{7,8} IHC screening has sensitivity for *MLH1*, *MSH2*, and *MSH6* of about 83% and a specificity of about 90% for each.

Screening of tumor tissue from patients enables genetic testing for a definitive diagnosis of Lynch syndrome and leads to counseling, cancer surveillance (eg, through frequent colonoscopic or endometrial screening examinations), and prophylaxis (eg, risk-reducing colorectal or gynecologic surgeries) for CRC patients, as well as for their family members.

Genetic testing for an MMR gene variant is often limited to *MLH1* and *MSH2* and, if negative, then *MSH6* and *PMS2*. The *BRAF* gene is often mutated in CRC when a particular *BRAF* variant (V600E, a change from valine to glutamic acid at amino acid position 600 in the BRAF protein) is present; to date, no *MLH1* gene variants have been reported.⁹ Therefore, patients negative for MLH1 protein expression by IHC, and therefore potentially positive for an *MLH1* variant, could first be screened for a *BRAF* variant. *BRAF*-positive samples need not be further tested by *MLH1* sequencing. *MLH1* gene methylation largely correlates with the presence of *BRAF*V600E and in combination with *BRAF* testing can accurately separate Lynch from sporadic CRC in IHC *MLH1*-negative cases.¹⁰

Recently, novel deletions have been reported to affect the expression of the *MSH2* gene in the absence of an *MSH2* gene variant, and thereby cause Lynch syndrome. In these cases, deletions in *EPCAM*, the gene for the epithelial cell adhesion molecule, are responsible. *EPCAM* testing has been added to many Lynch syndrome profiles and is conducted only when tumor tissue screening results are MSI-high and/or IHC shows a lack of *MSH2* expression, but no *MSH2* variant is found by sequencing. *EPCAM* is found just upstream, in a transcriptional sense, of *MSH2*. Deletions of *EPCAM* that encompass the last 2 exons of the *EPCAM* gene, including the polyadenylation signal that normally ends transcription of DNA into messenger RNA, results in transcriptional “read-through” and subsequent hypermethylation of the nearby and downstream *MSH2* promoter. This hypermethylation prevents normal MSH2 protein expression and leads to Lynch syndrome in a fashion similar to Lynch cases in which an *MSH2* variant prevents *MSH2* gene expression. Several studies have characterized such *EPCAM* deletions, established their correlation with the presence of *EPCAM-MSH2* fusion messenger RNAs (apparently nonfunctional) and with the presence of *MSH2* promoter hypermethylation, and, most importantly, have shown the cosegregation of these *EPCAM* variants with Lynch-like disease in families.¹¹⁻¹⁶

Distinct from patients with *EPCAM* deletions, rare cases of Lynch syndrome have been reported without detectable germline MMR variants, although IHC testing demonstrated a loss of expression of one of the MMR proteins. In at least some of these cases, research has identified germline “epivariants,” ie, methylation of promoter regions that control the expression of the MMR genes.^{11,17,18} Such methylation may be isolated or be in conjunction with a linked genetic alteration near the affected MMR gene. The germline epivariants may arise de novo or may be heritable in Mendelian or non-Mendelian fashion. This is distinct from some cases of MSI-high sporadic CRC wherein the tumor tissue may show *MLH1* promoter methylation and IHC nonexpression, but the same is not true of germline cells. Clinical testing for Lynch syndrome-related germline epivariants is not routine but may help in exceptional cases.

Female patients with Lynch syndrome have a predisposition to endometrial cancer. Lynch syndrome is estimated to account for 2% of all endometrial cancers in women and 10% of endometrial cancers in women younger than 50 years of age. Female carriers of the germline variants *MLH1*, *MSH2*, *MSH6*, and *PMS2* have an estimated 40% to 62% lifetime risk of developing endometrial cancer, as well as a 4% to 12% lifetime risk of ovarian cancer.

Population Selection

Various attempts have been made to identify which patients with colon cancer should undergo testing for MMR variants, based primarily on family history and related characteristics using criteria such as the Amsterdam II criteria¹⁹ (low sensitivity but high specificity), Revised Bethesda guidelines²⁰ (better sensitivity but poorer specificity), and risk prediction models (eg, MMRpro; PREMM5; MMRpredict).²¹ While family history is an important risk factor and should not be discounted in counseling families, it has poor sensitivity and specificity for identifying Lynch syndrome. Based on this and other evidence, the Evaluation of Genomic Applications in Practice and Prevention Working Group recommended testing all newly diagnosed CRC patients for Lynch syndrome, using a screening strategy based on MSI or IHC (with or without *BRAF*) followed by sequencing in screen-positive patients. This recommendation includes genetic testing for the following types of patients:

- Family members of Lynch syndrome patients with a known MMR variant; family members would be tested only for the family variant; those testing positive would benefit from early and increased surveillance to prevent future CRC.
- Patients with a differential diagnosis of Lynch syndrome vs attenuated FAP vs MAP.
- For Lynch syndrome patients, genetic testing of the proband with CRC likely benefits the proband where Lynch syndrome is identified, and appropriate surveillance for associated malignancies can be initiated and maintained and benefits family members by identifying the family variant.

Juvenile Polyposis Syndrome

Juvenile polyposis syndrome (JPS) is an autosomal dominant genetic disorder characterized by the presence of multiple hamartomatous (benign) polyps in the digestive tract. It is rare, with an estimated incidence of 1 in 100,000 to 160,000. Generalized juvenile polyposis refers to polyps in the upper and lower gastrointestinal tract, and juvenile polyposis coli refers to polyps of the colon and rectum. Those with JPS are at a higher risk for colorectal and gastric cancer.²² Approximately 60% of patients with JPS have a germline variant in the *BMPR1A* gene or the *SMAD4* gene.^{23,24} Approximately 25% of patients have de novo variants.^{25,26} In most cases, polyps appear in the first decade of life and most patients are symptomatic by age 20 years.²⁷ Rectal bleeding is the

most common presenting symptom, occurring in more than half of patients. Other presenting symptoms include prolapsing polyp, melena, pain, iron deficiency anemia, and diarrhea.^{22,26,27} As noted, individuals with JPS are at increased risk for colorectal and gastric cancer. By 35 years of age, the cumulative risk of CRC is 17% to 22%, which increases to 68% by age 60 years.^{28,29} The estimated lifetime risk of gastric cancer is 20% to 30%, with a mean age at diagnosis of 58 years.^{22,26,28} JPS may also be associated with hereditary hemorrhagic telangiectasia.³⁰ The most common clinical manifestations of hereditary hemorrhagic telangiectasia are telangiectasias of the skin and buccal mucosa, epistaxis, and iron deficiency anemia from bleeding.

Diagnosis

A clinical diagnosis of JPS is made on the basis of the presence of any one of the following: at least 3 to 5 juvenile polyps in the colon or multiple juvenile polyps in other parts of the gastrointestinal tract or any number of juvenile polyps in a person with a known family history of juvenile polyps.³¹ It is recommended that individuals who meet clinical criteria for JPS undergo genetic testing for a germline variant in the *BMPRI1A* and *SMAD4* genes for a confirmatory diagnosis of JPS and to counsel at-risk family members.

Peutz-Jeghers Syndrome

Peutz-Jeghers syndrome (PJS) is also an autosomal dominant genetic disorder, similar to JPS, and characterized by the presence of multiple hamartomatous (benign) polyps in the digestive tract, mucocutaneous pigmentation, and an increased risk of gastrointestinal and nongastrointestinal cancers. It is rare, with an estimated incidence of 1 in 8000 to 200,000. In most cases, a germline variant in the *STK11 (LKB1)* gene is responsible for PJS, which has a high penetrance of over 90% by the age of 30 years.³²⁻³⁴ However, 10% to 20% of individuals with PJS have no family history and are presumed to have PJS due to de novo variants.³⁵ A variant in *STK11* is detected in only 50% to 80% of families with PJS, suggesting that there is a second PJS gene locus.

The reported lifetime risk for any cancer is between 37% and 93% among those diagnosed with PJS with an average age of cancer diagnosis at 42 years. The most common sites for malignancy are colon and rectum, followed by breast, stomach, small bowel, and pancreas.³⁶ The estimated lifetime risk of gastrointestinal cancer ranges from 38% to 66%.³⁶ Lifetime cancer risk stratified by organ site is colon and rectum (39%), stomach (29%), small bowel (13%), and pancreas (11%-36%).

Diagnosis

A clinical diagnosis of PJS is made if an individual meets two or more of the following criteria: presence two or more histologically confirmed PJ polyps of the small intestine or characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, fingers, or family history of PJS.³¹ Individuals who meet clinical criteria for PJS should undergo genetic testing for a germline variant in the *STK11* gene for a confirmatory diagnosis of PJS and counseling at-risk family members. In addition, if there is a known *SMAD4* variant in the family, genetic testing should be performed within the first 6 months of life due to hereditary hemorrhagic telangiectasia risk.³¹

REGULATORY STATUS

Clinical laboratories may develop and validate tests in-house and market them as a laboratory service; laboratory-developed tests (LDTs) must meet the general regulatory standards of the

Clinical Laboratory Improvement Act (CLIA). Genetic tests reviewed in this evidence review are available under the auspices of CLIA. Laboratories that offer LDTs must be licensed by CLIA for high-complexity testing. To date, the U.S. Food and Drug Administration has chosen not to require any regulatory review of this test.

POLICY

A. MMR Gene Testing

1. Genetic testing for MMR genes (*MLH1*, *MSH2*, *MSH6*, *PMS2*) may be considered **medically necessary** in the following patients:
 - a) Patients with colorectal cancer (CRC), for the diagnosis of Lynch syndrome (see Policy Guidelines).
 - b) Patients with endometrial cancer and a first-degree relative diagnosed with a Lynch-associated cancer (see Policy Guidelines), for the diagnosis of Lynch syndrome.
 - c) At-risk relatives (see Policy Guidelines) of patients with Lynch syndrome with a known MMR gene variant.
 - d) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome. Whether testing begins with *APC* variants or screening for MMR genes depends on clinical presentation.
 - e) Patients without CRC but with a family history meeting the Amsterdam criteria, when no affected family members have been tested for MMR variants.

B. APC Testing

1. Genetic testing for adenosis polyposis coli (*APC*) may be considered **medically necessary** in the following patients:
 - a) At-risk relatives (see Policy Guidelines) of patients with familial adenomatous polyposis (FAP) and/or a known *APC* variant.
 - b) Patients with a differential diagnosis of attenuated FAP vs *MUTYH*-associated polyposis (MAP) vs Lynch syndrome. Whether testing begins with *APC* variants or screening for mismatch repair (MMR) variants depends on clinical presentation.
2. Genetic testing for *APC* gene variants is **not medically necessary** for colorectal cancer patients with classical FAP for confirmation of the FAP diagnosis.

C. *MUTYH* Testing

1. Testing for *MUTYH* gene variants may be considered **medically necessary** in the following patients:
 - a) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome and a negative result for *APC* gene variants. A family history of no parents or children with FAP is consistent with MAP (autosomal recessive).

D. *EPCAM* Testing

1. Genetic testing for *EPCAM* gene variants may be considered **medically necessary** when any one of the following 3 major criteria (solid bullets) is met:
 - a) Patients with CRC, for the diagnosis of Lynch syndrome (see Policy Guidelines section) when:
 - i. Tumor tissue shows lack of MSH2 protein expression by immunohistochemistry and patient is negative for a *MSH2* germline variant; OR
 - ii. Tumor tissue shows a high level of microsatellite instability and patient is negative for a germline variant in *MLH1*, *MSH2*, *MSH6*, and *PMS2*; OR
 - b) At-risk relatives (see Policy Guidelines section) of patients with Lynch syndrome with a known *EPCAM* variant; OR
 - c) Patients without CRC but with a family history meeting the Amsterdam criteria, when no affected family members have been tested for MMR variants.

E. *BRAFV600E* OR *MLH1* PROMOTER METHYLATION

1. Genetic testing for *BRAFV600E* or *MLH1* promoter methylation may be considered **medically necessary** to exclude a diagnosis of Lynch syndrome when the MLH1 protein is not expressed in a CRC tumor on immunohistochemical analysis.

F. *SMAD4* AND *BMPRI1A* TESTING

1. Genetic testing for *SMAD4* and *BMPRI1A* gene variants may be considered **medically necessary** when any one of the following major criteria (solid bullets) is met:
 - a) Patients with a clinical diagnosis of juvenile polyposis syndrome based on the presence of any one of the following:

- i. at least 3 to 5 juvenile polyps in the colon
- ii. multiple juvenile polyps in other parts of the gastrointestinal tract
- iii. any number of juvenile polyps in a person with a known family history of juvenile polyps.

- b) At-risk relative of a patient suspected of or diagnosed with juvenile polyposis syndrome.

G. *STK11* Testing

1. Genetic testing for *STK11* gene variants may be considered **medically necessary** when any one of the following major criteria (solid bullets) is met:

- a) Patients with a clinical diagnosis of Peutz-Jeghers syndrome based on the presence of any 2 of the following:
 - i. presence of 2 or more histologically confirmed Peutz-Jeghers polyps of the small intestine
 - ii. characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, or fingers
 - iii. family history of Peutz-Jeghers syndrome

- b) At-risk relative of a patient suspected of or diagnosed with Peutz-Jeghers syndrome.

- H. Genetic testing for all other gene variants for Lynch syndrome or CRC is considered **experimental / investigational**.

Policy Guidelines

1. Testing At-Risk Relatives

Due to the high lifetime risk of cancer of the majority of the genetic syndromes discussed in this policy, "at-risk relatives" primarily refers to first-degree relatives. However, some judgment must be allowed, for example, in the case of a small family pedigree, when extended family members may need to be included in the testing strategy.

2. Targeted Familial Variant Testing

- a. It is recommended that, when possible, initial genetic testing for FAP or Lynch syndrome be performed in an affected family member so that testing in unaffected family members can focus on the variant found in the affected family member.
- b. In many cases, genetic testing for *MUTYH* gene variants should first target the specific variants *Y165C* and *G382D*, which account more than 80% of variants in Caucasian populations, and subsequently proceed to sequencing only as necessary. In other ethnic populations, however, proceeding directly to sequencing is appropriate.

3. Evaluation for Lynch Syndrome

- a. For patients with colorectal cancer being evaluated for Lynch syndrome, either the microsatellite instability (MSI) test, or the immunohistochemistry (IHC) test with or without BRAF gene variant testing, should be used as an initial evaluation of tumor tissue prior to MMR gene analysis. Both tests are not necessary. Consideration of proceeding to MMR gene sequencing would depend on results of MSI or IHC testing. IHC testing in particular may help direct which MMR gene likely contains a variant, if any, and may also provide some additional information if MMR genetic testing is inconclusive.
- b. When indicated, genetic sequencing for MMR gene variants should begin with MLH1 and MSH2 genes unless otherwise directed by the results of IHC testing. Standard sequencing methods will not detect large deletions or duplications; when MMR gene variants are expected based on IHC or MSI studies but none are found by standard sequencing, additional testing for large deletions or duplications is appropriate.
- c. Several Clinical Laboratory Improvement Amendments (CLIA)-licensed clinical laboratories offer MMR gene variant testing for Lynch syndrome. For example, the GeneTests website (available online at: http://www.ncbi.nlm.nih.gov/sites/GeneTests/lab/clinical_disease_id/2622?db=genetests) lists 32 U.S.-located laboratories that offer this service. In at least 1 laboratory, Lynch syndrome variant testing is packaged under 1 copyrighted name. The COLARIS® test from Myriad Genetic Laboratories includes sequence analysis of *MLH1*, *MSH2*, *MSH6*, and *PMS2*; large rearrangement analysis for *MLH1*, *MSH2*, *PMS2*, and *MSH6* large deletions/duplications; and analysis for large deletions in the *EPCAM* gene near *MSH2*. Note that there may be 2 versions of this test, the COLARIS (excludes *PMS2* testing) and COLARIS Update (includes *PMS2* testing). Individualized testing (eg, targeted testing for a family variant) can also be requested. The COLARIS^{PLUS} test includes full sequence analysis of *MLH1*, *MSH2*, *MSH6*, *PMS2*, and *MYH* genes and rearrangement analysis of *MLH1*, *MSH2*, *MSH6*, *MYH*, and *EPCAM* by microarray comparative genomic hybridization analysis, and multiplex ligation-dependent probe amplification analysis for *PMS2*.
- d. Similarly, GeneTests lists 15 U.S.-based CLIA-licensed clinical laboratories that provide APC variant testing and 14 that provide MUTYH variant testing. The COLARIS® AP test from Myriad Genetic Laboratories includes DNA sequencing analysis of the APC and MUTYH genes, as well as analysis of large rearrangements in the APC gene that are not detected by DNA sequencing.
- e. The Amsterdam II Clinical Criteria (all criteria must be fulfilled) are the most stringent criteria for defining families at high risk for Lynch syndrome (Vasen et al, 1999):
 - 1) 3 or more relatives with an associated cancer (colorectal cancer, or cancer of the endometrium, small intestine, ureter or renal pelvis);
 - 2) 1 should be a first-degree relative of the other 2;
 - 3) 2 or more successive generations affected;

- 4) 1 or more relatives diagnosed before the age of 50 years;
 - 5) Familial adenomatous polyposis (FAP) should be excluded in cases of colorectal carcinoma;
 - 6) Tumors should be verified by pathologic examination.
 - 7) Modifications:
 - 8) EITHER: very small families, which cannot be further expanded, can be considered to have HNPCC with only 2 colorectal cancers in first-degree relatives if at least 2 generations have the cancer and at least 1 case of colorectal cancer was diagnosed by the age of 55 years; OR
 - 9) in families with 2 first-degree relatives affected by colorectal cancer, the presence of a third relative with an unusual early-onset neoplasm or endometrial cancer is sufficient.
4. The revised Bethesda Guidelines (fulfillment of any criterion meets guidelines) are less stringent than the Amsterdam criteria and are intended to increase the sensitivity of identifying at-risk families (Umar et al [2004]). The Bethesda guidelines are also considered more useful in identifying which patients with colorectal cancer should have their tumors tested for microsatellite instability and/or immunohistochemistry:
- a. Colorectal carcinoma (CRC) diagnosed in a patient who is less than 50 years old;
 - b. Presence of synchronous (at the same time) or metachronous (at another time, ie, a recurrence of) CRC or other Lynch syndrome–associated tumors, regardless of age;
 - c. CRC with high microsatellite instability histology diagnosed in a patient less than 60 years old;
 - d. CRC diagnosed in 1 or more first-degree relatives with a Lynch syndrome-associated tumor, with one of the cancers being diagnosed before 50 years of age;
 - e. CRC diagnosed in 2 or more first- or second-degree relatives with HNPCC-related tumors,^a regardless of age.
- ^a HNPCC-related tumors include colorectal, endometrial, stomach, ovarian, pancreas, ureter and renal pelvis, biliary tract, brain (usually glioblastoma as seen in Turcot syndrome), sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel.
5. Genetic Counseling
Experts recommend formal genetic counseling for patients who are at risk for inherited disorders and who wish to undergo genetic testing. Interpreting the results of genetic tests and understanding risk factors can be difficult for some patients; genetic counseling helps individuals understand the impact of genetic testing, including the possible effects the test results could have on the individual or their family members. It should be noted that genetic counseling may alter the utilization of genetic testing substantially and may reduce inappropriate testing; further, genetic counseling should be performed by an individual with experience and expertise in genetic medicine and genetic testing methods.

6. Genetics Nomenclature Update

- a. The Human Genome Variation Society nomenclature is used to report information on variants found in DNA and serves as an international standard in DNA diagnostics. It is being implemented for genetic testing medical evidence review updates starting in 2017 (see Table PG1). The Society's nomenclature is recommended by the Human Variome Project, the HUman Genome Organization, and by the Human Genome Variation Society itself.
- b. The American College of Medical Genetics and Genomics and the Association for Molecular Pathology standards and guidelines for interpretation of sequence variants represent expert opinion from both organizations, in addition to the College of American Pathologists. These recommendations primarily apply to genetic tests used in clinical laboratories, including genotyping, single genes, panels, exomes, and genomes. Table PG2 shows the recommended standard terminology—"pathogenic," "likely pathogenic," "uncertain significance," "likely benign," and "benign"—to describe variants identified that cause Mendelian disorders.

Table PG1. Nomenclature to Report on Variants Found in DNA

Previous	Updated	Definition
Mutation	Disease-associated variant	Disease-associated change in the DNA sequence
	Variant	Change in the DNA sequence
	Familial variant	Disease-associated variant identified in a proband for use in subsequent targeted genetic testing in first-degree relatives

Table PG2. ACMG-AMP Standards and Guidelines for Variant Classification

Variant Classification	Definition
Pathogenic	Disease-causing change in the DNA sequence
Likely pathogenic	Likely disease-causing change in the DNA sequence
Variant of uncertain significance	Change in DNA sequence with uncertain effects on disease
Likely benign	Likely benign change in the DNA sequence
Benign	Benign change in the DNA sequence

ACMG: American College of Medical Genetics and Genomics; AMP: Association for Molecular Pathology.

RATIONALE

The most recent literature review was performed through July 9, 2018. The following is a summary of the key findings to date.

Evidence reviews assess whether a medical test is clinically useful. A useful test provides information to make a clinical management decision that improves the net health outcome. That is, the balance of benefits and harms is better when the test is used to manage the condition than when another test or no test is used to manage the condition.

The first step in assessing a medical test is to formulate the clinical context and purpose of the test. The test must be technically reliable, clinically valid, and clinically useful for that purpose. Evidence reviews assess the evidence on whether a test is clinically valid and clinically useful.

Technical reliability is outside the scope of these reviews, and credible information on technical reliability is available from other sources.

Genetic Testing for Familial Adenomatous Polyposis and *MUTYH*-Associated Polyposis Clinical Context and Test Purpose

The purpose of genetic testing for familial adenomatous polyposis (FAP) and *MUTYH*-associated polyposis (MAP) is to

- Identify at-risk relatives of patients with FAP and/or a known adenomatous polyposis coli (*APC*) gene variant
- Make a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome.

The questions addressed in this evidence review are: (1) Is there evidence that genetic testing for FAP has clinical validity?; and (2) Does genetic testing for attenuated FAP change patient management in a way that improves outcomes as a result of genetic testing?

The following PICOTS were used to select literature to inform this review.

Patients

The relevant populations of interest are at-risk relatives of patients with FAP and/or a known *APC* variant or those who require a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome.

Interventions

The relevant intervention is genetic testing for *APC* or *MUTYH*. Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing FAP and MAP: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be the early detection of colorectal cancer (CRC) and appropriate and timely interventional strategies (eg, endoscopic resection, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse events from that treatment or undertreatment.

Timing

Genetic testing for FAP may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual of having or developing FAP.

Setting

Ordering and interpreting genetic testing may be complex and is best done by experienced specialists such as gastroenterologists. Most patients are likely to be tested in an outpatient

setting. Referral for genetic counseling is important for the explanation of genetic disease, heritability, genetic risk, test performance, and possible outcomes.

Study Selection Criteria

For the evaluation of clinical validity of the genetic test, studies that meet the following eligibility criterion were considered:

- Reported on the analytic sensitivity and specificity and/or diagnostic yield of the test.

Technically Reliable

Assessment of technical reliability focuses on specific tests and operators and requires review of unpublished and often proprietary information. Review of specific tests, operators, and unpublished data are outside the scope of this evidence review and alternative sources exist. This evidence review focuses on the clinical validity and clinical utility.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

The evidence review for FAP genetic testing was informed by a TEC Assessment (1998).³⁷ The additional information on attenuated FAP and on MAP diagnostic criteria and genetic testing is based on information from *GeneReviews*³⁸ and from several publications³⁹⁻⁴² that build on prior, cited research.

The analytic sensitivity and specificity for *APC* and *MUTYH* are both 99%. Clinical sensitivity for classic FAP is about 95%; about 90% of pathogenic variants are detected by sequencing^{38,43} while 8% to 12% of pathogenic variants are detected by deletion and duplication testing.^{44,45}

Among Northern European whites, 85% of pathogenic *MUTYH* variants are detected by the 2 variant test (Y165C, G382D)^{46,47} and 98% of pathogenic *MUTYH* variants are detected by full gene sequencing.^{48,49}

A comprehensive review of the *APC* pathogenic variant and its association with classical FAP and attenuated FAP and MAP is beyond the scope of this evidence review. *GeneReviews*³⁸ reported that the likelihood of detecting an *APC* pathogenic variant is highly dependent on the severity of colonic polyposis^{44,50-52} and on the family history.⁵³ Detection rates are higher in classic polyposis (88%) than in nonclassical FAPs such as attenuated colonic phenotypes (57%) or MAP (33%).

Section Summary: Clinically Valid

The analytic and clinical sensitivity and specificity for *APC* and *MUTYH* are high. About 90% of pathogenic variants in classical FAP are detected by sequencing while 8% to 12% of pathogenic variants are detected by deletion and duplication testing. Among Northern European whites, 85% of pathogenic *MUTYH* variants are detected by the 2 variant test, and 98% of pathogenic *MUTYH* variants are detected by full gene sequencing. The likelihood of detecting an *APC* pathogenic variant is highly dependent on the severity of colonic polyposis and family history. Detection rates are higher in classic polyposis (88%) than in nonclassical FAPs such as attenuated colonic phenotypes (57%) or MAP (33%).

Clinical Useful

A test is clinically useful if use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, or more effective therapy, or avoid unnecessary therapy, or avoid unnecessary testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from randomized controlled trials (RCTs).

No RCTs were identified assessing the clinical utility of genetic testing for FAP and MAP.

Chain of Evidence

Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to demonstrate test performance, no inferences can be made about clinical utility.

Genetic testing of patients requiring a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome may have clinical utility:

- If the test supports the clinical diagnosis of an attenuated disease, the protocol for endoscopic surveillance is affected and, depending on the situation, may avoid more frequent but unnecessary surveillance or necessitates more frequent surveillance.

Genetic testing of at-risk relatives of patients with FAP and/or a known *APC* variant may have clinical utility:

- If, in the absence of genetic testing, the diagnosis of colorectal polyposis in at-risk relatives of patients with FAP and/or a known *APC* variant can only be established by colonoscopy and subsequent histologic examination of removed polyps, which are burdensome.
- If results are negative, the test results may provide release from the intensified screening program resulting in psychological relief.

A TEC Assessment (1998)³⁷ offered the following conclusions:

- Genetic testing for FAP may improve health outcomes by identifying which currently unaffected at-risk family members require intense surveillance or prophylactic colectomy.
- At-risk subjects are considered to be those with greater than 10 adenomatous polyps or close relatives of patients with clinically diagnosed FAP or of patients with an identified *APC* variant.
- The optimal testing strategy is to define the specific genetic variant in an affected family member and then test the unaffected family members to see if they have inherited the same variant.

Table 1 summarizes clinical utility studies assessing genetic testing for FAP.

Testing for the *APC* variant has no role in the evaluation, diagnosis, or treatment of patients with classical FAP where the diagnosis and treatment are based on the clinical presentation.

Table 1. Summary of Clinical Utility Studies for Genetic Testing for FAP

Study	Study Design and Population	Results
Bjork et al (2000) ⁵⁴	Observational:195 confirmed cases of FAP underwent ileorectal anastomosis and followed for, on average, 14 y	Cumulative risk of rectal cancer mortality was 7% at 20 y postsurgery and cumulative mortality was 11.1% at the age

Study	Study Design and Population	Results
Järvinen (1992) ⁵⁵	Observational: 251 individuals from 81 affected families; 76 individuals diagnosed during family screening vs 116 symptomatic individuals with probands	of 70 y, indicating a substantial risk of developing cancer even after surgery 65.5% of symptomatic cases had CRC vs 6.6% cases among those screened during family screening
Vasen et al (1990) ⁵⁶	Observational: CRC rate compared in 230 confirmed FAP cases; 104 symptomatic and 126 at-risk family members identified by screening	47% of symptomatic cases had CRC at a mean age of 35 y vs 4% at 24 y

CRC: colorectal cancer; FAP: familial adenomatous polyposis.

Section Summary: Clinically Useful

Direct evidence of clinical utility for genetic testing of attenuated FAP is not available. Genetic testing of at-risk relatives of patients with FAP and/or a known *APC* variant or those requiring a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from intensified screening program resulting in psychological relief, and may improve health outcomes by identifying currently unaffected at-risk family members who require intense surveillance or prophylactic colectomy.

Lynch Syndrome and CRC Genetic Testing

Clinical Context and Test Purpose

The purpose of genetic testing for Lynch syndrome is to:

- Detect Lynch syndrome in patients diagnosed with colorectal or endometrial cancer
- Identify at-risk relatives of patients with a diagnosed Lynch syndrome and/or a known mismatch repair (MMR) variant and/or positive family history meeting the Amsterdam or Revised Bethesda criteria
- Make a differential diagnosis of attenuated FAP v MAP vs Lynch syndrome.

The questions addressed in this evidence review are: (1) Is there evidence that genetic testing for Lynch syndrome has clinical validity?; and (2) Does genetic testing for Lynch syndrome change patient management in a way that improves outcomes as a result of genetic testing?

The following PICOTS were used to select literature to inform this review.

Patients

The relevant populations of interest are patients diagnosed with colorectal or endometrial cancer or at-risk relatives of patients with a diagnosed Lynch syndrome and/or a known MMR variant and/or positive family history meeting the Amsterdam or Revised Bethesda criteria or those requiring a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome.

Interventions

The relevant intervention is genetic testing for the *MLH1*, *MSH2*, *MSH6*, *PMS2*, *EPCAM*, and/or *BRAFV600E* genes. Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing Lynch syndrome: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be early detection of Lynch syndrome and appropriate and timely interventional strategies (eg, increased surveillance, endoscopic resection, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse effects from that treatment or undertreatment.

Timing

Genetic testing for Lynch syndrome may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual having or developing Lynch syndrome.

Setting

Ordering and interpreting genetic testing may be complex and is best done by experienced specialists such as gastroenterologists. Most patients are likely to be tested in an outpatient setting. Referral for genetic counseling is important for the explanation of genetic disease, heritability, genetic risk, test performance, and possible outcomes.

Study Selection Criteria

For the evaluation of clinical validity of the genetic test, studies that met the following eligibility criterion were considered:

- Reported on the analytic sensitivity and specificity and/or diagnostic yield of the test.

Technically Reliable

Assessment of technical reliability focuses on specific tests and operators and requires review of unpublished and often proprietary information. Review of specific tests, operators, and unpublished data are outside the scope of this evidence review and alternative sources exist. This evidence review focuses on the clinical validity and clinical utility.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

Microsatellite instability (MSI) and immunohistochemical (IHC) screening tests for MMR variants have similar sensitivity and specificity. MSI screening has a sensitivity of about 89% for *MLH1* and *MSH2* and 77% for *MSH6* and a specificity of about 90% for all. IHC screening has sensitivity for *MLH1*, *MSH2*, and *MSH6* of about 83% and a specificity of about 90% for each.

The evidence for Lynch syndrome genetic testing in patients with CRC is based on an evidence report conducted for the Agency for Healthcare Research and Quality by Bonis et al (2007),⁵⁷ a supplemental assessment to that report contracted by the Evaluation of Genomic Applications in Practice and Prevention (EGAPP) Working Group (2009),⁹ and an EGAPP recommendation (2009) for genetic testing in CRC.⁵⁸ Based on the Agency for Healthcare Research and Quality report and supplemental assessment, the EGAPP recommendation concluded the following about genetic testing for MMR variants in patients already diagnosed with CRC:

- Family history, while important information to elicit and consider in each case, has poor sensitivity and specificity as a screening test to determine who should be considered for MMR variant testing and should not be used as a sole determinant or screening test.
- Optional *BRAF* testing can be used to reduce the number of patients, who are negative for *MLH1* expression by IHC, needing *MLH1* gene sequencing, thus improving efficiency without reducing sensitivity for MMR variants.

Moreira et al (2012) compared universal testing of CRC patients with alternative screening approaches.⁵⁹ The alternative screening approaches included using the Bethesda guidelines, the Jerusalem recommendations, and a selective strategy including only those diagnosed with CRC before age 70, or after age 70 if meeting the Bethesda guidelines. In the analysis of 10206 newly diagnosed CRC patients from 4 large cohort studies, MSI testing was used in 2150 patients, and immunostaining was used in 2278 patients, while both MSI and immunostaining were used in 5591 patients. MMR gene variants were found in 312 (3.1%) patients overall. The universal screening approach was superior to the other screening approaches in the population-based cohorts (n=3671 probands). Table 2 summarizes the results of the different screening approaches.

Table 2. Diagnostic Results of the Different Screening Approaches

Screening Approach	Sensitivity (95% CI), %	Specificity (95% CI), %	Diagnostic Yield (95% CI), %
Universal	100 (99.3 to 100)	93 (95 CI, 92.0 to 93.7)	2.2 (95 CI, 1.7 to 2.7)
Bethesda guidelines	87.8 (95 CI, 78.9 to 93.2)	97.5 (95 CI, 96.9 to 98.0)	2.0 (95 CI, 1.5 to 2.4)
Jerusalem recommendations	85.4 (95 CI, 77.1 to 93.6)	96.7 (95 CI, 96.0 to 97.2)	1.9 (95 CI, 1.4 to 2.3)
Selective strategy	95.1 (95 CI, 89.8 to 99.0)	95.5 (95 CI, 94.7 to 96.1)	2.1 (95 CI, 1.6 to 2.6)

CI: confidence interval.

However, the diagnostic yield differences between the screening approaches were small, and the false-positive yield was 2.5% with universal screening. In the selective strategy, 34.8% fewer patients required tumor MMR testing and 28.6% fewer required analyses of MMR variants, resulting in a 4.9% rate of missed Lynch syndrome cases.

Several studies have characterized *EPCAM* deletions, established their correlation with the presence of *EPCAM-MSH2* fusion messenger RNAs (apparently nonfunctional) and with the presence of *MSH2* promoter hypermethylation, and, most importantly, have shown the cosegregation of these *EPCAM* variants with Lynch-like disease in families.¹¹⁻¹⁶ Because studies differ slightly in how patients were selected, the prevalence of these *EPCAM* variants is difficult to estimate but may be in the range of 20% to 40% of patients/families who meet Lynch syndrome criteria, do not have an MMR variant, but have MSI-high tumor tissue. Kempers et al (2011) reported that carriers of an *EPCAM* deletion had a 75% (95% CI, 65% to 85%) cumulative risk of CRC by age 70 years, which did not differ significantly from that of carriers of an *MSH2* deletion (77%; 95% CI, 64% to 90%); mean age at diagnosis was 43 years.⁶⁰ However, the cumulative risk of endometrial cancer was low at 12% (95% CI, 0% to 27%) by age 70 compared with carriers of an *MSH2* variant (51%; 95% CI, 33% to 69%; p<0.001).

Bouzourene et al (2010) analyzed *MLH1* protein abnormalities in 11 patients with sporadic CRC and 16 patients with Lynch syndrome.¹⁰ A *BRAF* variant was not found in any of the Lynch syndrome patients. *MLH1* promoter methylation was only present in 1 Lynch syndrome patient.

However, 8 of the 11 sporadic CRC patients had the *BRAF* variant, and all 11 patients were *MLH1* methylated, suggesting patients with *BRAF* variants could be excluded from germline testing for Lynch syndrome. Jin et al (2013) evaluated MMR proteins in 412 newly diagnosed CRC patients.⁶¹ *MLH1* and *PMS2* protein stains were absent in 65 patients who were subsequently tested for *BRAF* variant. Thirty-six (55%) of the 65 patients had the *BRAF*V600E variant, thus eliminating the need for further genetic testing or counseling for Lynch syndrome. Capper et al (2013) reported on a technique of V600E IHC testing for *BRAF* variants on a series of 91 stratified as high MSI CRC patients.⁶² The authors detected *BRAF*-mutated CRC with 100% sensitivity and 98.8% specificity. V600E positive lesions were detected in 21% of *MLH1*-negative CRC patients who could be excluded from MMR germline testing for Lynch syndrome. Therefore, V600E IHC testing for *BRAF* could be an alternative to *MLH1* promoter methylation analysis. To summarize, *BRAF*V600E variant or *MLH1* promoter methylation testing are optional screening methods that may be used when IHC testing shows a loss of *MLH1* protein expression. The presence of *BRAF* V600E or absence of *MLH1* protein expression due to *MLH1* promoter methylation rarely occurs in Lynch syndrome and would eliminate the need for further germline variant analysis for a Lynch syndrome diagnosis.⁶³

The risk of endometrial cancer in MMR variant carriers has been estimated at 34% (95% CI, 17% to 60%) by age 70, and at 8% for ovarian cancer (95% CI, 2% to 39%) by age 70.⁶⁴ Risks do not appear to appreciably increase until after age 40. In a prospective study by Leenen et al (2012), 179 consecutive endometrial cancer patients 70 years of age or younger were analyzed for MSI, using IHC for expression of 4 MMR proteins, MMR gene methylation status, and *BRAF* variants.⁶⁵ Results are presented in Table 3; 92% of patients were older than 50 years of age.

Table 3. Testing Unselected Endometrial Cancer Patients for Lynch Syndrome

Outcomes	N	Percent (95% Confidence Interval)
Microsatellite stable and normal protein staining	137	76
MSI-H and <i>MLH1</i> absent	32	
Sporadic MSI-H	31	17 (13 to 24)
Likely to have Lynch syndrome	11	6 (3 to 11)
Variant-positive	7	
No variant found	3	
Refused further DNA testing	1	

MSI-H: high microsatellite instability.

Clinically Useful

A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, or more effective therapy, or avoid unnecessary therapy, or avoid unnecessary testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from RCTs.

No RCTs were identified assessing the clinical utility of genetic testing for Lynch syndrome.

Chain of Evidence

Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to demonstrate test performance, no inferences can be made about clinical utility.

Genetic testing of patients with colon or endometrial cancer to detect Lynch syndrome has clinical utility:

- To make decisions about the preferred approach for treatment (endoscopic resection, colectomy with ileorectal anastomosis or segmental colectomy).

Genetic testing of at-risk relatives of patients with Lynch syndrome and/or a known MMR variant and/or positive family history meeting the Amsterdam or Revised Bethesda criteria or risk prediction scores has clinical utility:

- If the individuals diagnosed with Lynch syndrome are recommended for screening for Lynch syndrome–associated cancers.
- If, in the absence of genetic testing, the diagnosis of Lynch syndrome in at-risk relatives of patients can only be established by colonoscopy and subsequent histologic examination of excised polyps, which is burdensome.
- If negative test results prompt release from an intensified screening program, thereby reducing in emotional burden.

Genetic testing of patients requiring a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome may have clinical utility:

- If the test supports the clinical diagnosis of Lynch syndrome, the protocol for endoscopic surveillance is affected and, depending on the situation, may avoid more frequent but unnecessary surveillance or necessitates more frequent surveillance.

A chain of evidence can be constructed for the clinical utility of testing all patients with CRC for MMR variants. EGAPP conclusions are summarized next.

1. The chain of evidence from well-designed experimental nonrandomized studies is adequate to demonstrate the clinical utility of testing unaffected (without cancer) first- and second-degree relatives of patients with Lynch syndrome who have a known MMR variant.
2. Seven studies examined how counseling affected testing and surveillance choices among unaffected family members of Lynch syndrome patients.⁶⁶⁻⁷² About half of the relatives received counseling, and 95% of them chose MMR gene variant testing. Among those positive for MMR gene variants, uptake of colonoscopic surveillance beginning at age 20 to 25 years was high at 53% to 100%.

One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed recommended colonic surveillance vs those who did not.

Surveillance and prevention for other Lynch syndrome cancers.

3. The chain of evidence from descriptive studies and expert opinion is inadequate (inconclusive) to demonstrate the clinical utility of testing the probands with Lynch syndrome (ie, the index patient).
 - a. Subtotal colectomy is recommended as an alternative to segmental resection but has not been shown superior in follow-up studies
 - b. Although a small body of evidence suggests that MSI-positive tumors are resistant to 5-fluorouracil and more sensitive to irinotecan than MSI-negative tumors, no alteration in therapy according to MSI status has yet been recommended.
 - c. Surveillance and prevention for other Lynch syndrome cancers:

- i. While invasive and not actively recommended, women may choose hysterectomy with salpingo-oophorectomy to prevent gynecologic cancer. In a retrospective study by Schmeler et al (2006), 315 women who chose this option had no gynecologic cancer over 10 years, whereas about one-third of women who did not have surgery developed endometrial cancer, and 5.5% developed ovarian cancer.⁷³
- ii. In a study by Bouzourene et al (2010), surveillance endometrial biopsy detected endometrial cancer and potentially precancerous conditions at earlier stages in those with Lynch syndrome, but results were not statistically significant, and a survival benefit has yet to be shown.¹⁰ Transvaginal ultrasound is not a highly effective surveillance mechanism for endometrial cancer in patients with Lynch syndrome; however, transvaginal ultrasound in conjunction with endometrial biopsy has been recommended for surveillance.
- iii. Gastroduodenoscopy for gastric cancer surveillance and urine cytology for urinary tract cancer surveillance are recommended based on expert opinion only, in the absence of adequate supporting evidence.

In early documentation of the natural history of CRC in highly selected families with a strong history of hereditary CRC, Fitzgibbons et al (1987) indicated risks of synchronous and metachronous cancers as high as 18% and 24%, respectively, in those with CRC.⁷⁴ As a result, the Cancer Genetic Studies Consortium (1997) recommended that if CRC is diagnosed in patients with an identified variant or a strong family history, a subtotal colectomy with ileorectal anastomosis should be considered as an option to segmental resection.⁷⁵ Although the average risk of a second primary is now estimated to be somewhat lower overall in patients with Lynch syndrome and CRC, effective prevention measures remain imperative. Van Dalen et al (2003) suggested that subtotal colectomy with ileorectal anastomosis markedly reduced the incidence of second surgery for metachronous cancer from 28% to 6% but could not rule out the impact of surveillance.⁷⁶ A 2003 mathematical model comparing total colectomy plus ileorectal anastomosis with hemicolectomy estimated increased life expectancies of 2.3, 1, and 0.3 years for ages 27, 47, and 67, respectively; for stage I cancer, estimated life expectancies for the same ages were 3.4, 1.5, and 0.4, respectively.⁷⁷ Based on this work, the 2006 joint American Society of Clinical Oncology and Society of Surgical Oncology review assessing risk-reducing surgery in hereditary cancers recommended offering both options to patients with Lynch syndrome and CRC, especially those who are younger.⁷⁸ The societies' review also recommended offering Lynch syndrome patients with an index rectal cancer the options of total proctocolectomy with ileal pouch anal anastomosis or anterior proctosigmoidectomy with primary reconstruction. The rationale for total proctocolectomy is the 17% to 45% rate of metachronous colon cancer in the remaining colon after an index rectal cancer in Lynch syndrome patients.

Table 4 summarizes the clinical utility studies assessing genetic testing for Lynch syndrome.

Table 4. Summary of Clinical Validity Studies for Genetic Testing for Lynch Syndrome

Study	Study Design and Population	Results
Yurgelun et al (2012) ⁷⁹	<ul style="list-style-type: none"> • Prospective cohort: Examined uptake of risk-reducing strategies in 40 women at risk for LS-associated endometrial cancer. • Cross-sectional cohort: Examined adoption of risk-reduction strategies using a one-time questionnaire in 77 women at risk of LS-associated endometrial cancer 	<ul style="list-style-type: none"> • In cross-sectional cohort, 58/77 (75%) women reported engaging in endometrial cancer risk-reduction • Proportion of women engaging in endometrial cancer risk-reduction strategy before genetic testing: 26/40 (65%). At 1-y follow-up, 16/16 (100%) MMR variant

Study	Study Design and Population	Results
		carriers were adherent to guidelines for risk-reduction, 9 (56%) of whom had had a prophylactic hysterectomy. By 3 y, 11/16 (69%) MMR variant carriers had a prophylactic hysterectomy. Among women with negative or uninformative genetic test results, none had a prophylactic hysterectomy after testing.
Engel et al (2010) ⁸⁰	Prospective cohort: Assessed efficacy of annual colonoscopic surveillance in 1126 at-risk individuals from families with LS	99 CRCs found in 90 individuals; 71 were diagnosed by surveillance colonoscopies. Median time between CRCs detected through follow-up colonoscopy and preceding colonoscopy was 11.3 mo.
Järvinen et al (2009) ⁸¹	Observational; 609 individuals from 57 LS families; 242 variant-positive and 367 variant-negative followed for cancer incidence over a mean of 11.5 y	No increase in cancer mortality in variant-positive vs -negative individuals; 74 variant-positive individuals had adenomas removed; 48 variant-positive women had prophylactic hysterectomy
Dove-Edwin et al (2005) ⁸²	Prospective observational; 554 individuals from 290 at-risk families with HNPCC or MMR variants followed for 16 y	Estimated 72% decrease in CRC death in screened individuals
De Vos tot Nederveen Cappel et al (2002) ⁸³	Observational; 857 at-risk individuals from 114 HNPCC- or MMR-positive families.	10-y cumulative risk of CRC, 15.7% vs 3.4% for partial vs subtotal colectomy
Syngal et al (1998) ⁸⁴	Decision analysis model: Assessed impact of decision about immediate prophylactic colectomy, delayed colectomy, or endoscopic surveillance at the time of a positive result on genetic testing	Compared with no intervention, all risk-reduction strategies led gains in life expectancy from 13.5 y for surveillance to 15.6 y for prophylactic proctocolectomy at 25 y of age. Also, surveillance led to QALY gain of 3.1 y vs 0.3 y with subtotal colectomy.
Järvinen et al (1995) ⁸⁵ ; Järvinen et al (2000) ⁸⁶	Observational; 252 at-risk individuals from 20 of 22 families with MMR variants invited for colonoscopy screening every 3 y; 133 agreed; 118 declined. Of those who declined, 8 (15%) had screening examinations outside of the study.	<ul style="list-style-type: none"> • Screening vs nonscreening • Incidence of CRC: 4.5% (n=6) vs 11.9% (n=14) (p=0.03) • 6 vs 12 deaths within 10 y (p=0.08)

CRC: colorectal cancer; HNPCC: hereditary nonpolyposis colorectal cancer; LS: Lynch syndrome; MMR: mismatch repair; QALY: quality of life adjusted years.

Kwon et al (2011) developed a Markov Monte Carlo simulation model to compare 6 strategies for Lynch syndrome testing in women with endometrial cancer.⁸⁷ Overall, the results suggested that IHC triage of women at any age who had at least 1 first-degree relative with a Lynch-associated cancer was the most effective strategy for identifying Lynch syndrome and subsequent CRC cases. The model used published prevalence estimates of Lynch syndrome in all endometrial cancer patients of 2% (range, 1%-3%), and of 17% (range, 15%-20%) in endometrial cancer patients with at least 1 first-degree relative with a Lynch-associated cancer. Results are presented in Table 5.

Table 5. Modeling of Endometrial Cancer Screening Strategies for Detecting Lynch Syndrome

Testing Strategy	No. Cases Subject to IHC Triage	No. Identified With Lynch Syndrome	No. Subsequent CRC Cases
Amsterdam II criteria	NA	539	2582
Age <50 y, and at least 1 FDR (Lynch-associated cancer)	NA	530	2470

Testing Strategy	No. Cases Subject to IHC Triage	No. Identified With Lynch Syndrome	No. Subsequent CRC Cases
IHC triage <age 50 y	6285	520	2442
IHC triage <age 60 y	16,226	548	2450
IHC triage at any age; at least 1 FDR with Lynch-associated cancer	5786	755	2442
IHC triage all endometrial cancers	45,000	827	2413

CRC: colorectal cancer; FDR: first-degree relative; IHC: immunohistochemical; NA: not available.

Females with Lynch syndrome who choose risk-reducing surgery are encouraged to consider oophorectomy because of the risk of ovarian cancer in Lynch syndrome. In another retrospective cohort study, Obermair et al (2010) found that hysterectomy improved survival among female colon cancer survivors with Lynch syndrome.⁸⁸ This study also estimated that, for every 100 women diagnosed with Lynch syndrome–associated CRC, about 23 would be diagnosed with endometrial cancer within 10 years absent a hysterectomy. Data on variant-specific risks have suggested that prophylactic gynecologic surgery benefits for carriers of *MSH6* variants may offer less obvious benefits compared with harms, because the lifetime risk of endometrial cancer is lower than for carriers of *MLH1* or *MSH2* variants, and the lifetime risk of ovarian cancer is similar to the risk for the general population.⁶⁴

However, for carriers of the *EPCAM* deletion, 3 studies (2011, 2012) reported on 3 cases of endometrial cancer in 103 female carriers who did not undergo a preventative hysterectomy.^{60,89,90} Women with *EPCAM* deletions consequently have a 1-fold lower lifetime risk of developing endometrial cancer than with carriers with an MMR variant. This might support a clinical management scenario rather than prophylactic surgery.⁸⁹ An alternative to prophylactic surgery is surveillance for endometrial cancer using transvaginal ultrasound and endometrial biopsy. Evidence has suggested that such surveillance significantly reduces the risk of interval cancers, but no evidence as yet has indicated surveillance reduces mortality due to endometrial cancer.⁹¹ Surveillance in Lynch syndrome populations for ovarian cancer has not yet been demonstrated to be successful at improving survival.⁹¹

Section Summary: Clinically Useful

Direct evidence of clinical utility for genetic testing for Lynch syndrome is not available. Multiple studies have demonstrated clinical utility in testing unaffected (without cancer) first- and second-degree relatives of patients with Lynch syndrome who have a known MMR variant, in that counseling has been shown to influence testing and surveillance choices among unaffected family members of Lynch syndrome patients. One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed and did not follow recommended colonic surveillance. A positive genetic test for an MMR gene variant can also lead to changes in the management of other Lynch syndrome malignancies.

Genetic Testing for Juvenile Polyposis Syndrome and Peutz-Jeghers Syndrome

Clinical Context and Test Purpose

The purpose of genetic testing for juvenile polyposis syndrome (JPS) and Peutz-Jeghers syndrome (PJS) is:

- To confirm a diagnosis of JPS or PJS in patients suspected of these disorders based on clinical features

- To identify at-risk relatives of patients with a confirmed diagnosis of JPS or PJS.

The questions addressed in this evidence review are: (1) Is there evidence that genetic testing for patients suspected of JPS and PJS has clinical validity?; and (2) Does genetic testing for JPS and PJS change patient management in a way that improves outcomes as a result of genetic testing?

The following PICOTS were used to select literature to inform this review.

Patients

The relevant populations of interest are patients with suspected JPS or PJS and individuals who are at-risk relatives of patients suspected of or diagnosed with a JPS or PJS.

Interventions

The relevant intervention is genetic testing for *SMAD4* and *BMPRI* (for JPS) and *ASATK11* (for PJS). Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing JPS and PJS: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be early detection of cancer and appropriate and timely interventional strategies (eg, cancer screening, surgical intervention including polyp resection, gastrectomy, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse events from that treatment or undertreatment.

Timing

Genetic testing for *SMAD4* and *BMPRI* (for JPS) and *ASATK11* (for PJS) may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual of having or developing JPS and PJS.

Setting

Ordering and interpreting genetic testing may be complex and is best done by experienced specialists such as gastroenterologists. Most patients are likely to be tested in an outpatient setting. Referral for genetic counseling is important for the explanation of genetic disease, heritability, genetic risk, test performance, and possible outcomes.

Study Selection Criteria

For the evaluation of clinical validity of the genetic test, studies that met the following eligibility criterion were considered:

- Reported on the diagnostic yield of the test.

Technically Reliable

Assessment of technical reliability focuses on specific tests and operators and requires review of unpublished and often proprietary information. Review of specific tests, operators, and unpublished data are outside the scope of this evidence review and alternative sources exist. This evidence review focuses on the clinical validity and clinical utility.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

Table 6 summarizes the clinical utility studies assessing genetic testing for JPS and PJS.

Table 6. Summary of Clinical Validity Studies Assessing Genetic Testing for JPS and PJS

Study	Study Design and Population	Results
Yang et al (2010) ⁹²	Observational; 17 clinically diagnosed children with PJS	<i>STK11</i> variants detected in 29.4% (5/17)
Calva-Cerqueira et al (2009) ⁹³	Observational; 102 unrelated JPS probands analyzed all of whom met clinical criteria for JPS	<i>SMAD4</i> and <i>BMPR1A</i> variants detected in 41% (42/102) JPS probands
Aretz et al (2007) ⁹⁴	Observational; 80 unrelated patients (65 met clinical criteria for typical JPS; 15 presumed to have JPS) were examined by direct sequencing for <i>SMAD4</i> , <i>BMPR1A</i> , and <i>PTEN</i> variants	<i>SMAD4</i> and <i>BMPR1A</i> variants detected in 60% of typical JPS patients and none in presumed JPS patients; overall diagnostic yield, 49%
Volikos et al (2006) ⁹⁵	Observational; 76 clinically diagnosed with PJS	Detection rate of germline variants was about 80% (59/76)
Aretz et al (2005) ⁹⁶	Observational; 71 patients (56 met clinical criteria for PJS; 12 presumed to have PJS)	<i>STK11</i> variant detected in 52% (37/71)

JPS: juvenile polyposis syndrome; PJS: Peutz-Jeghers syndrome.

Section Summary: Clinically Valid

The likelihood of detecting a pathogenic variant is highly dependent on the presence of clinical features and family history. Detection rates for JPS and PJS have been reported to be between 60% and 41% and 29.4% and 80%, respectively.

Clinical Useful

A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, or more effective therapy, or avoid unnecessary therapy, or avoid unnecessary testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from RCTs.

No RCTs were identified assessing the clinical utility of genetic testing for JPS and PJS.

Chain of Evidence

Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to demonstrate test performance, no inferences can be made about clinical utility.

Genetic testing of patients with suspected JPS and PJS has clinical utility:

- To make decisions about a preferred approach for treatment (endoscopic resection, colectomy with ileorectal anastomosis, segmental colectomy).

Genetic testing of individuals who are at-risk relatives of patients suspected of or diagnosed with JPS or PJS has clinical utility:

- If the individuals diagnosed with JPS and PJS are recommended for screening for JPS and PJS-associated cancers.
- If, in the absence of genetic testing, the diagnosis of JPS and PJS in at-risk relatives of patients can only be established by colonoscopy and subsequent histologic examination of excised polyps, which is burdensome.
- If negative test results prompt release from an intensified screening program, thereby reducing in emotional burden.

Table 7 summarizes clinical utility studies assessing genetic testing for JPS and PJS.

Table 7. Summary of Clinical Utility Studies for Genetic Testing for JPS and PJS

Study	Study Design and Population	Results
Aytac et al (2015) ⁹⁷	Observational: 35 patients had germline variants in <i>BMPR1A</i> (8 patients) or <i>SMAD4</i> (27) with a median follow-up of 11 y	No patient was diagnosed with cancer in the <i>BMPR1A</i> group, whereas 4 men with a <i>SMAD4</i> variant developed GI (n=3) or extraintestinal (n=1) cancer. The GI cancer risk in patients with JPS and a <i>SMAD4</i> variant was 11% (3/27).
Resta et al (2013) ⁹⁸	Observational: 119 patients with PJS	Cancer occurred in 31/119 patients (RR for overall cancer risk, 15.1); mean age at first cancer diagnosis was 41 y. Kaplan-Meier estimates for overall cumulative cancer risks were 20%, 43%, 71%, and 89%, at age 40, 50, 60, and 65 y, respectively.
Lier et al (2010) ³⁶	Systematic review: 21 original articles, 20 cohort studies, and 1 meta-analysis (total N=1644 PJS patients)	349 patients developed 384 malignancies at average age of 42 y. Lifetime risk for any cancer varied between 37% and 93% with RRs ranging from 9.9 to 18 vs the general population.
Salloch et al (2009) ⁹⁹	Observational: 31 patients with PJS; <i>STK11</i> variants in 16/22 families	10 carcinomas detected in 6 patients resulting in a cancer risk of 65% up to the age of 65 y; surveillance strategy detected 50% of cancers (n=5) at an early potentially curable stage
Brosens et al (2007) ²⁹	Observational: 84 patients with JPS contributing 1652.2 person-years of follow-up vs general population of the U.S. (SEER data)	RR of CRC was 34.0 (95% CI, 14.4 to 65.7); cumulative life-time risk for CRC was 38.7%; mean age of diagnosis of CRC, 43.9 y

CI: confidence interval; CRC: colorectal cancer; GI: gastrointestinal; JPS: juvenile polyposis syndrome; PJS: Peutz-Jeghers syndrome; RR: relative risk.

Section Summary: Clinically Useful

Direct evidence of the clinical utility for genetic testing of JPS or PJS is not available. Genetic testing of patients with suspected JPS or PJS or individuals who are at-risk relatives of patients suspected of or diagnosed with a polyposis syndrome or PJS may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from intensified screening program resulting in psychological relief, and may improve health outcomes by identifying currently unaffected at-risk family members who require intense surveillance or prophylactic colectomy.

SUMMARY OF EVIDENCE

For individuals who are suspected of attenuated FAP, MAP, and Lynch syndrome who receive genetic testing for *APC*, or are at-risk relatives of patients with FAP who receive genetic testing for *MUTYH* after a negative *APC* test result, the evidence includes a TEC Assessment. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and validity. For patients with an *APC* variant, enhanced surveillance and/or prophylactic treatment will reduce the future incidence of colon cancer and improve health outcomes. A related familial polyposis syndrome, MAP syndrome, is associated with variants in the *MUTYH* gene. Testing for this genetic variant is necessary when the differential diagnosis includes both FAP and MAP because distinguishing between the 2 leads to different management strategies. Depending on presentation, Lynch syndrome may be part of the same differential diagnosis. The evidence is sufficient to determine that the technology results in a meaningful improvement in the net health outcome.

For individuals who (1) are suspected of attenuated FAP, MAP, and Lynch syndrome, or (2) have colon cancer, or (3) have endometrial cancer and a first-degree relative diagnosed with a Lynch-associated cancer, or (4) are at-risk relatives of patients with Lynch syndrome, or (5) are without colon cancer but with a family history meeting the Amsterdam or Revised Bethesda criteria who receive genetic testing for MMR genes, the evidence includes an Agency for Healthcare Research and Quality report, a supplemental assessment to that report by the Evaluation of Genomic Applications in Practice and Prevention Working Group, and an Evaluation of Genomic Applications in Practice and Prevention recommendation for genetic testing in CRC. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and validity. A chain of evidence from well-designed experimental nonrandomized studies is adequate to demonstrate the clinical utility of testing unaffected (without cancer) first- and second-degree relatives of patients with Lynch syndrome who have a known variant in an MMR gene, in that counseling has been shown to influence testing and surveillance choices among unaffected family members of Lynch syndrome patients. One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed recommended colonic surveillance. A positive genetic test for an MMR variant can also lead to changes in the management of other Lynch syndrome malignancies. The evidence is sufficient to determine that the technology results in a meaningful improvement in the net health outcome.

For individuals who warrant Lynch testing, screen negative on MMR testing, but positive for microsatellite instability and lack MSH2 protein expression who receive genetic testing for *EPCAM* variants, the evidence includes variant prevalence studies and case series. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and validity. Studies have shown an association between *EPCAM* variants and Lynch-like disease in families, and the cumulative risk for CRC is similar to carriers of an *MSH2* variant. Identification of an *EPCAM* variant could lead to changes in management that improve health outcomes. The evidence is sufficient to determine that the technology results in a meaningful improvement in the net health outcome.

For individuals who have CRC in whom MLH1 protein is not expressed on immunohistochemical analysis who receive genetic testing for *BRAFV600E* or *MLH1* promoter methylation, the evidence includes case series. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and validity. Studies have shown, with high sensitivity and specificity, an association between *BRAFV600E* variant and *MLH1* promoter methylation with sporadic CRC. Therefore, this type of testing could eliminate the need for further genetic testing or counseling

for Lynch syndrome. The evidence is sufficient to determine that the technology results in a meaningful improvement in the net health outcome.

For individuals who (1) are suspected of JPS or PJS or (2) are at-risk relatives of patients suspected of or diagnosed with JPS or PJS who receive genetic testing for *SMAD4*, *BMPR1A*, or *STK11* genes, respectively, the evidence includes multiple observational studies. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and validity. Studies have shown, with high sensitivity and specificity, an association between *SMAD4* and *BMPR1A* and *STK11* variants with JPS and PJS, respectively. Direct evidence of clinical utility for genetic testing of a JPS or PJS is not available. Genetic testing may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from intensified screening program resulting in psychological relief, and may improve health outcomes by identifying currently unaffected at-risk family members who require intense surveillance or prophylactic colectomy. The evidence is sufficient to determine that the technology results in a meaningful improvement in the net health outcome.

CLINICAL INPUT RECEIVED THROUGH PHYSICIAN SPECIALTY SOCIETIES AND ACADEMIC MEDICAL CENTERS

While the various physician specialty societies and academic medical centers may collaborate with and make recommendations during this process, through the provision of appropriate reviewers, input received does not represent an endorsement or position statement by the physician specialty societies or academic medical centers, unless otherwise noted.

In response to requests, input was received from 3 physician specialty societies and 3 academic medical centers while this policy was under review in 2009. In general, those providing input were in agreement with the overall approach described in this policy.

PRACTICE GUIDELINES AND POSITION STATEMENTS

National Comprehensive Cancer Network

National Comprehensive Cancer Network (NCCN) guidelines (v.1.2018) for genetic/familial high-risk assessment of colorectal cancer (CRC)³¹ recommend 2 approaches to Lynch syndrome variant screening: (1) all newly diagnosed CRC or (2) all CRC patients diagnosed before age 70 plus those diagnosed at ages 70 and older who meet Bethesda guidelines. Additionally, NCCN guidelines (v.2.2018) recommend screening for Lynch syndrome in all endometrial cancer patients younger than 50 years.¹⁰⁰ Genetic testing is recommended for at-risk family members of patients with positive variants in *MLH1*, *MSH2*, *MSH6*, and *PMS2*. NCCN guidelines also indicate *BRAFV600E* testing or *MLH1* promoter methylation testing may be used when *MLH1* is not expressed in the tumor on immunohistochemical analysis to exclude a diagnosis of Lynch syndrome. These guidelines also address familial adenomatous polyposis (classical and attenuated) and *MUTYH*-associated polyposis and are consistent with the information provided in this evidence review.

NCCN guidelines for colon cancer (v.3.2018)¹⁰¹ and for CRC screening (v.3.2018)¹⁰² recommend CRC patients treated with curative-intent surgery undergo surveillance colonoscopy at 1 year postsurgery and, if normal, again in 3 years, then every 5 years based on findings. However, because of the high likelihood of cancer, colonoscopy is recommended every 1 to 2 years throughout life for patients with Lynch syndrome before cancer diagnosis; and the high likelihood of a second primary cancer is based on a first cancer diagnosis.⁸³ NCCN guidelines on

genetic/familial high-risk assessment for colorectal indicate for *MLH1*, *MSH2*, and *EPCAM* variant carriers that surveillance with colonoscopy should begin “at age 20 to 25 years or 2 to 5 years before the earliest colon cancer if it is diagnosed before age 25 years and repeat every 1 to 2 years.”³¹ “*MSH6* variant carriers should begin surveillance with colonoscopy at age 30 to 35 years, and *PMS2* carriers should begin surveillance at age 35 to 40 years. However, screening may need to be initiated earlier in some families, depending on ages of cancers observed in family members. This screening is recommended every 2 to 3 years until age 40 or 50 years for *MSH6* and *PMS2* variant carriers, respectively, at which time colonoscopy should be performed every 1 to 2 years.” “If the patient is not a candidate for routine surveillance, subtotal colectomy may be considered.”

NCCN guidelines for colon cancer recommend that patients 70 years or younger plus those older than 70 years of age who meet the Bethesda guidelines be tested for the mismatch repair (MMR) protein for possible Lynch syndrome.¹⁰¹ These guidelines also indicate all colon cancer patients should be questioned about family history and considered for risk assessment as per NCCN colorectal screening guidelines. NCCN guidelines for uterine neoplasm also recommend universal screening for MMR genes.¹⁰⁰

There are limited data on the efficacy of various screening modalities in juvenile polyposis syndrome and Peutz-Jeghers syndrome. NCCN cancer risk and surveillance guidelines for these 2 indications are summarized in Tables 8 and 9.³¹

Table 8. Risk and Surveillance Guidelines for Peutz-Jeghers Syndrome (Category 2A Recommendations)

Site	Lifetime Risk, %	Screening Procedure and Interval	Initiation Age, y
Breast	45-50	<ul style="list-style-type: none"> • Mammogram and breast MRI annually • Clinical breast exam every 6 mo 	≈25 y
Colon	39	Colonoscopy every 2-3 y	Late teens
Stomach	29	Upper endoscopy every 2-3 y	Late teens
Small intestine	13	Small bowel visualization (CT or MRI enterography or video capsule endoscopy baseline at 8-10 y with follow-up interval based on findings but at least by age 18, then every 2-3 y, though this may be individualized, or with symptoms)	≈8 to 10 y
Pancreas	11-36	Magnetic resonance cholangiopancreatography with contrast or endoscopic ultrasound every 1-2 h	≈30 to 35 y
Ovary (typically benign sex cord/Sertoli cell tumors)	18-21	<ul style="list-style-type: none"> • Pelvic examination and Pap smear annually • Consider transvaginal ultrasound 	≈18 to 20 y
Cervix (typically cervical adenoma malignum)	10		
Uterus	9		
Testes (typically sex cord/Sertoli cell tumors)		Annual testicular exam and observation for feminizing changes	≈10 y
Lung	15-17	<ul style="list-style-type: none"> • Provide education about symptoms and smoking cessation • No other specific recommendations have been made 	

CT: computed tomography; MRI: magnetic resonance imaging.

Table 9. Risk and Surveillance Guidelines for Juvenile Polyposis Syndrome (Category 2A Recommendations)

Site	Lifetime Risk, %	Screening Procedure and Interval	Initiation Age, y
Colon	40-50	Colonoscopy every year if polyps are found and every 2-3 y if no polyps are found ^a	≈15 y
Stomach	21 if multiple polyps	Upper endoscopy annually if polyps are found and every 2-3 y if no polyps are found ^a	≈15 y
Small intestine	Rare, undefined	No recommendations made	
Pancreas	Rare, undefined	No recommendations made	
HHT	Undefined	In individuals with <i>SMAD4</i> variants, screen for vascular lesions associated with HHT	Within first 56 mo of age

HHT: hereditary hemorrhagic telangiectasia.

^a In families without an identified genetic variants, consider substituting endoscopy every 5 y beginning at age 20 and every 10 y beginning at age 40 y in patients in whom no polyps are found.

American College of Gastroenterology

The American College of Gastroenterology (2015) issued practice guidelines for the management of patients with hereditary gastrointestinal cancer syndromes.²⁶

For Lynch syndrome, the College recommended:

- "All newly diagnosed colorectal cancers (CRCs) should be evaluated for mismatch repair deficiency.
- Analysis may be done by immunohistochemical testing for the *MLH1/MSH2/MSH6/PMS2* proteins and/or testing for microsatellite instability. Tumors that demonstrate loss of *MLH1* should undergo BRAF testing or analysis for *MLH1* promoter hypermethylation.
- Individuals who have a personal history of a tumor showing evidence of mismatch repair deficiency (and no demonstrated BRAF variant or hypermethylation of *MLH1*), a known family variant associated with LS [Lynch syndrome], or a risk of ≥5% chance of LS based on risk prediction models should undergo genetic evaluation for LS.¹⁰³
- Genetic testing of patients with suspected LS should include germline variant genetic testing for the *MLH1, MSH2, MSH6, PMS2*, and/or *EPCAM* genes or the altered gene(s) indicated by IHC testing."

For adenomatous polyposis syndromes, the College recommended:

- "*Familial adenomatous polyposis (FAP)/MUTYH-associated polyposis/attenuated polyposis*
- Individuals who have a personal history of >10 cumulative colorectal adenomas, a family history of one of the adenomatous polyposis syndromes, or a history of adenomas and FAP-type extracolonic manifestations (duodenal/ampullary adenomas, desmoid tumors, papillary thyroid cancer, congenital hypertrophy of the retinal pigment epithelium, epidermal cysts, osteomas) should undergo assessment for the adenomatous polyposis syndromes.
- Genetic testing of patients with suspected adenomatous polyposis syndromes should include *APC* and *MUTYH* gene variant analysis."

For juvenile polyposis syndrome, the College recommended:

- "Genetic evaluation of a patient with possible JPS [juvenile polyposis syndrome] should include testing for *SMAD4* and *BMPRI1A* mutations"

- "Surveillance of the gastrointestinal (GI) tract in affected or at-risk JPS patients should include screening for colon, stomach, and small bowel cancers (strong recommendation, very low quality of evidence).
- Colectomy and ileorectal anastomosis or proctocolectomy and ileal pouch-anal anastomosis is indicated for polyp-related symptoms, or when the polyps cannot be managed endoscopically (strong recommendation, low quality of evidence).
- Cardiovascular examination for and evaluation for hereditary hemorrhagic telangiectasia should be considered for *SMAD4* mutation carriers (conditional recommendation, very low quality of evidence)."

For Peutz-Jeghers syndrome, the College recommended:

- "Genetic evaluation of a patient with possible PJS [Peutz-Jeghers syndrome] should include testing for *STK11* mutations."
- "Surveillance in affected or at-risk PJS patients should include monitoring for colon, stomach, small bowel, pancreas, breast, ovary, uterus, cervix, and testes cancers. Risk for lung cancer is increased, but no specific screening has been recommended. It would seem wise to consider annual chest radiograph or chest computed tomography (CT) in smokers (conditional recommendation, low quality of evidence)."

American Society of Clinical Oncology and Society of Surgical Oncology

In 2015, the American Society of Clinical Oncology concluded that the European Society for Medical Oncology clinical practice guideline published in 2013 were based on the most relevant scientific evidence and therefore endorsed them with minor qualifying statements (in bold italics).¹⁰⁴ The recommendations as relate to genetic testing hereditary CRC syndromes are summarized below:

- "Tumor testing *for DNA mismatch repair (MMR) deficiency* with immunohistochemistry for MMR proteins and/or MSI should be ***assessed*** in all CRC patients. As an alternate strategy, tumor testing should be carried out in individuals with CRC younger than 70 years, or those older than 70 years who fulfill any of the revised Bethesda guidelines.
- If loss of MLH1/PMS2 ***protein expression*** is observed in the tumor, analysis of *BRAF* V600E mutation or analysis of methylation of the *MLH1* promoter should be carried out first to rule out a sporadic case. ***If tumor is MMR deficient and somatic BRAF mutation is not detected or MLH1 promoter methylation is not identified, testing for germline mutations is indicated.***
- If loss of any of the other proteins (MSH2, MSH6, PMS2) is observed, germline genetic testing should be carried out ***for the genes corresponding to the absent proteins (eg, MSH2, MSH6, EPCAM, PMS2, or MLH1).***
- Full germline genetic testing ***for Lynch syndrome*** should include DNA sequencing and large rearrangement analysis...
- Patients with multiple colorectal adenomas should be considered for full germline genetic testing of *APC* and/or *MUTYH*."
- Germline testing of *MUTYH* can be initiated by screening for the most common mutations (*G396D, Y179C*) in the white population followed by analysis of the entire gene in heterozygotes. Founder mutations among ethnic groups should be taken into account. ***For nonwhite individuals, full sequencing of MUTYH should be considered.***"

U.S. PREVENTIVE SERVICES TASK FORCE RECOMMENDATIONS

No U.S. Preventive Services Task Force recommendations for genetic testing of Lynch syndrome and other inherited colon cancer syndromes have been identified.

ONGOING AND UNPUBLISHED CLINICAL TRIALS

Some currently unpublished trials that might influence this review are listed in Table 10.

Table 10. Summary of Key Trials

NCT No.	Trial Name	Planned Enrollment	Completion Date
Unpublished			
NCT01646112	Living in Lynch Syndrome Limbo: Exploring the Meaning of Uncertain Genetic Test Results	34	Feb 2016 (completed)
NCT01850654	Ohio Colorectal Cancer Prevention Initiative: Universal Screening for Lynch Syndrome	4000	Sep 2017

NCT: national clinical trial.

CODING

The following codes for treatment and procedures applicable to this policy are included below for informational purposes. Inclusion or exclusion of a procedure, diagnosis or device code(s) does not constitute or imply member coverage or provider reimbursement. Please refer to the member's contract benefits in effect at the time of service to determine coverage or non-coverage of these services as it applies to an individual member.

CPT/HCPCS

- 81201 APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; full gene sequence
- 81202 APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; known familial variants
- 81203 APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; duplication/deletion variants
- 81210 BRAF (rB-Raf proto-oncogene, serine/threonine kinase) (eg, colon cancer, melanoma), gene analysis, V600 variant(s)
- 81288 MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; promoter methylation analysis
- 81292 MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis
- 81293 MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants
- 81294 MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication; deletion variants
- 81295 MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis

- 81296 MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants
- 81297 MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication / deletion variants
- 81298 MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis
- 81299 MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants
- 81300 MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication / deletion variants
- 81301 Microsatellite instability analysis (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) of markers for mismatch repair deficiency (eg, BAT25, BAT26), includes comparison of neoplastic and normal tissue, if performed
- 81317 PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis
- 81318 PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants
- 81319 PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants
- 81403 Molecular pathology procedure, Level 4 (eg, analysis of single exon by DNA sequence analysis, analysis of >10 amplicons using multiplex PCR in 2 or more independent reactions, variant scanning or duplication/deletion variants of 2-5 exons)

- There are specific CPT codes for genetic testing of *APC*: 81201, 81202, 81203.
- There are specific CPT codes for genetic testing of *MLH1*, *MSH2*, *MSH6*, *PMS2*, and microsatellite instability:
 - *MLH1* genetic testing code range: 81292, 81293, 81294, 81288.
 - *MSH2* genetic testing code range: 81295, 81296, 81297.
 - *MSH2* genetic testing code range: 81298, 81299, 81300.
 - Microsatellite instability analysis (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) of markers for mismatch repair deficiency, (eg, BAT25, BAT26), includes comparison of neoplastic and normal tissue, if performed: 81301.
 - *PMS2* genetic testing code range: 81317, 81318, 81319.
- There is also a specific CPT code for testing of *BRAF*V600 variant(s): 81210.
- The following CPT code includes the testing for *EPCAM*: 81403.
- Genetic testing for colon cancer is not widely available and is most commonly performed by commercial reference labs or research labs dedicated to genetic testing in general.

ICD-10 Diagnoses

- C17.0 Malignant neoplasm of duodenum
- C17.1 Malignant neoplasm of jejunum
- C17.2 Malignant neoplasm of ileum
- C17.3 Meckel's diverticulum, malignant
- C17.8 Malignant neoplasm of overlapping sites of small intestine
- C17.9 Malignant neoplasm of small intestine, unspecified

C18.3	Malignant neoplasm of hepatic flexure
C18.4	Malignant neoplasm of transverse colon
C18.6	Malignant neoplasm of descending colon
C18.7	Malignant neoplasm of sigmoid colon
C18.0	Malignant neoplasm of cecum
C18.1	Malignant neoplasm of appendix
C18.2	Malignant neoplasm of ascending colon
C18.5	Malignant neoplasm of splenic flexure
C18.8	Malignant neoplasm of overlapping sites of colon
C18.9	Malignant neoplasm of colon, unspecified
C19	Malignant neoplasm of rectosigmoid junction
C25.1	Malignant neoplasm of body of pancreas
C25.2	Malignant neoplasm of tail of pancreas
C56.1	Malignant neoplasm of right ovary
C56.2	Malignant neoplasm of left ovary
C56.9	Malignant neoplasm of unspecified ovary
C57.00	Malignant neoplasm of unspecified fallopian tube
C57.01	Malignant neoplasm of right fallopian tube
C57.02	Malignant neoplasm of left fallopian tube
C57.10	Malignant neoplasm of unspecified broad ligament
C57.11	Malignant neoplasm of right broad ligament
C57.12	Malignant neoplasm of left broad ligament
C57.3	Malignant neoplasm of parametrium
C57.20	Malignant neoplasm of unspecified round ligament
C57.21	Malignant neoplasm of right round ligament
C57.22	Malignant neoplasm of left round ligament
C60.1	Malignant neoplasm of glans penis
C71.0	Malignant neoplasm of cerebrum, except lobes and ventricles
C71.1	Malignant neoplasm of frontal lobe
C71.2	Malignant neoplasm of temporal lobe
C71.3	Malignant neoplasm of parietal lobe
C71.4	Malignant neoplasm of occipital lobe
C71.5	Malignant neoplasm of cerebral ventricle
C71.6	Malignant neoplasm of cerebellum
C71.7	Malignant neoplasm of brain stem
C71.8	Malignant neoplasm of overlapping sites of brain
C71.9	Malignant neoplasm of brain, unspecified
D12.0	Benign neoplasm of cecum
D12.1	Benign neoplasm of appendix
D12.2	Benign neoplasm of ascending colon
D12.3	Benign neoplasm of transverse colon
D12.4	Benign neoplasm of descending colon
D12.5	Benign neoplasm of sigmoid colon
D12.6	Benign neoplasm of colon, unspecified
K63.5	Polyp of colon
D12.7	Benign neoplasm of rectosigmoid junction
D12.8	Benign neoplasm of rectum
D12.9	Benign neoplasm of anus and anal canal

D01.0	Carcinoma in situ of colon
D01.1	Carcinoma in situ of rectosigmoid junction
D01.2	Carcinoma in situ of rectum
Z85.038	Personal history of other malignant neoplasm of large intestine
Z85.048	Personal history of other malignant neoplasm of rectum, rectosigmoid junction, and anus
Z80.0	Family history of malignant neoplasm of digestive organs

REVISIONS

05-13-2011	Policy added to the bcbsks.com web site.
01-01-2012	In the Coding section: <ul style="list-style-type: none"> Added the new codes: 81210, 81292-81301
04-10-2012	In the Coding section: <ul style="list-style-type: none"> Replaced Diagnosis code 183.1 with correct code 183.2. Removed HCPCS codes: S3828, S3829, S3830, S3831 (Deleted codes, effective April 1, 2012.)
01-15-2013	In the Coding section: <ul style="list-style-type: none"> Added CPT codes: 81401, 81406 Added new CPT codes: 81201, 81202, 81203(Effective 01-01-2013) Removed CPT codes:83890, 83892, 83898, 83902, 83904, 83905, 83906, 83912 (Effective 12-31-2012)
03-26-2013	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> In Item I, Note, Amsterdam II Criteria, added "6. Modifications: EITHER: very small families, which cannot be further expanded, can be considered to have HNPCC with only 2 colorectal cancers in first-degree relatives if at least two generations have the cancer and at least one case of colorectal cancer was diagnosed by the age of 55 years; OR: in families with two first-degree relatives affected by colorectal cancer, the presence of a third relative with an unusual early-onset neoplasm or endometrial cancer is sufficient." In Item I, Note, Revised Bethesda Criteria, added "6. Colorectal cancer diagnosed with one or more first-degree relatives with HNPCC-related tumor (colorectal, endometrial, stomach, ovarian, pancreas, bladder, ureter and renal pelvis, biliary tract, brain [usually glioblastoma as seen in Turcot syndrome], sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel), with one of the cancers being diagnosed under age 50 years, OR colorectal cancer diagnosed in two or more first-or second-degree relatives with HNPCC related tumor, regardless of age. (15)"
	Updated Rationale section.
	Updated Reference section.
08-21-2013	In Coding section: <ul style="list-style-type: none"> Removed CPT code 81210. Added ICD-10 Diagnosis codes (Effective October 1, 2014)
01-01-2015	Policy posted 01-16-2015
	In Coding section: <ul style="list-style-type: none"> Added CPT Code: 81288 (Effective January 1, 2015)
03-18-2015	In Title section: <ul style="list-style-type: none"> Changed title name from "Genetic Testing for Inherited Susceptibility to Colon Cancer, Including Microsatellite Instability Testing"

	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> Removed Amsterdam II criteria and Revised Bethesda guidelines. In Policy Guidelines, added items 6-9.
	Updated Rationale section.
	In Coding section: <ul style="list-style-type: none"> Added CPT Codes 81210, 81317, 81318, 81319, and 81403. Removed CPT Code 81406.
	Updated References section.
01-01-2016	In Coding section: <ul style="list-style-type: none"> Revised nomenclature to CPT codes: 81210 and 81401.
02-03-2016	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> Added statement on genetic counseling to Policy Guidelines.
	Updated Rationale section.
	Updated References section.
	Added Appendix section.
05-25-2016	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> Revised Policy Guideline Item 6.
	Updated Rationale section.
	Updated References section.
11-09-2016	In Policy section: <ul style="list-style-type: none"> In Item I B, removed "when feasible" and "who meet the revised Bethesda criteria (see Policy Guidelines below)" and added "or immunohistochemical (IHC) analysis of tumors" and "or endometrial" to read, "Microsatellite instability (MSI) testing or immunohistochemical (IHC) analysis of tumors may be considered medically necessary as an initial test in persons with colorectal or endometrial cancer in order to identify those persons who should proceed with HNPCC variant analysis."
	In Coding section: <ul style="list-style-type: none"> Added CPT codes: 88341, 88342, 88344.
12-08-2017	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> Added new Item I B, "HNPCC genetic testing is considered experimental / investigational for all other indications." Previous Item I B is now Item I C. Added new Item I D, "MSI testing or IHC analysis of tumors is considered experimental / investigational for all other indications." Added new Item II B, "APC genetic testing is considered experimental / investigational for all other indications." Previous Item II B is now Item II C. Added new Item II D, "MAP genetic testing is considered experimental / investigational for all other indications."
	Updated Rationale section.
	In Coding section: <ul style="list-style-type: none"> Removed ICD-9 codes.
	Updated References section.
02-18-2019	Updated Description section.
	In Policy section: <ul style="list-style-type: none"> Removed the previous policy language: "I. Lynch syndrome (also known as Hereditary Non-Polyposis Colorectal Cancer [HNPCC]): A. Genetic testing for HNPCC (MLH1, MSH2, MSH6, PMS2 sequence analysis) is considered medically necessary when one

	<p>of the following criteria are met: 1. Meets Amsterdam II criteria or revised Bethesda guidelines (see Policy Guidelines below); or 2. A first-* or second-degree** relative with an HNPCC variant (genes <i>MLH1</i>, <i>MSH2</i>, <i>MSH6</i>, <i>PMS2</i>); or 3. Endometrial cancer 50 years of age or younger. B. HNPCC genetic testing is considered experimental / investigational for all other indications. C. Microsatellite instability (MSI) testing or immunohistochemical (IHC) analysis of tumors may be considered medically necessary as an initial test in persons with colorectal or endometrial cancer in order to identify those persons who should proceed with HNPCC variant analysis. D. MSI testing or IHC analysis of tumors is considered experimental / investigational for all other indications. II. Familial Adenomatous Polyposis and associated variance: A. Adenosis polyposis coli (<i>APC</i>) genetic testing is considered medically necessary for either of the following indications: 1. Greater than 10 colonic polyps; or 2. First-degree* relatives diagnosed with familial adenomatous polyposis (FAP) or with a documented <i>APC</i> variant. The specific <i>APC</i> variant should be identified in the affected first-degree relative with FAP prior to testing the member, if feasible. Full sequence <i>APC</i> genetic testing is considered medically necessary only when it is not possible to determine the family variant first. B. <i>APC</i> genetic testing is considered experimental / investigational for all other indications. C. Testing for <i>MYH</i> variants is considered medically necessary for any of the following indications: 1. Personal history of 10 to 20 adenomatous polyposis who have negative <i>APC</i> variant testing and a negative family history for adenomatous polyposis; OR 2. Personal history of 10 to 20 adenomatous polyposis whose family history is consistent with recessive inheritance (ie, family history is positive only for sibling[s]); OR 3. Asymptomatic siblings of individuals with known <i>MYH</i> polyposis. D. <i>MAP</i> genetic testing is considered experimental / investigational for all other indications. *First-degree relatives are parents, siblings, and offspring. ** Second-degree relatives are aunts, uncles, grandparents, niece, nephews or half-siblings. ^ Hereditary nonpolyposis colorectal cancer (HNPCC)-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, ureter and renal pelvis, brain (usually glioblastoma as seen in Turcot syndrome), and small intestinal cancers, as well as sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome.”</p> <ul style="list-style-type: none"> ▪ Added new policy language, “A. MMR Gene Testing 1. Genetic testing for MMR genes (<i>MLH1</i>, <i>MSH2</i>, <i>MSH6</i>, <i>PMS2</i>) may be considered medically necessary in the following patients: a) Patients with colorectal cancer (CRC), for the diagnosis of Lynch syndrome (see Policy Guidelines. b) Patients with endometrial cancer and a first-degree relative diagnosed with a Lynch-associated cancer (see Policy Guidelines), for the diagnosis of Lynch syndrome. c) At-risk relatives (see Policy Guidelines) of patients with Lynch syndrome with a known MMR gene variant. d) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome. Whether testing begins with <i>APC</i> variants or screening for MMR genes depends on clinical presentation. e) Patients without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, when no affected family members have been tested for MMR variants. B. <i>APC</i> Testing 1. Genetic testing for adenosis polyposis coli (<i>APC</i>) may be considered medically necessary in the following patients: a) At-risk relatives (see Policy Guidelines) of patients with familial adenomatous polyposis (FAP) and/or a known <i>APC</i> variant. b) Patients with a differential diagnosis of attenuated FAP vs <i>MUTYH</i>-associated polyposis (MAP) vs Lynch syndrome. Whether testing begins with <i>APC</i> variants or screening for mismatch repair (MMR) variants depends on clinical presentation. 2. Genetic testing for <i>APC</i> gene variants is not medically necessary for colorectal cancer patients with classical FAP for confirmation of the FAP diagnosis. C. <i>MUTYH</i> Testing 1. Testing for <i>MUTYH</i> gene variants may be considered medically necessary in the following patients: a) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome and a negative result for <i>APC</i> gene variants. A family history of no parents or children with FAP is consistent with
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	<p>MAP (autosomal recessive). D. <i>EPCAM</i> Testing 1. Genetic testing for <i>EPCAM</i> gene variants may be considered medically necessary when any one of the following 3 major criteria (solid bullets) is met: 1) Patients with CRC, for the diagnosis of Lynch syndrome (see Policy Guidelines section) when: i. Tumor tissue shows lack of MSH2 protein expression by immunohistochemistry and patient is negative for a <i>MSH2</i> germline variant; OR ii. Tumor tissue shows a high level of microsatellite instability and patient is negative for a germline variant in <i>MLH1</i>, <i>MSH2</i>, <i>MSH6</i>, and <i>PMS2</i>; OR b) At-risk relatives (see Policy Guidelines section) of patients with Lynch syndrome with a known <i>EPCAM</i> variant; OR c) Patients without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, when no affected family members have been tested for MMR variants, and when sequencing for MMR variants is negative. E. <i>BRAF</i>V600E or <i>MLH1</i> promoter methylation 1. Genetic testing for <i>BRAF</i> V600E or <i>MLH1</i> promoter methylation may be considered medically necessary to exclude a diagnosis of Lynch syndrome when the MLH1 protein is not expressed in a CRC tumor on immunohistochemical analysis. F. <i>SMAD4</i> and <i>BMPR1A</i> Testing 1. Genetic testing for <i>SMAD4</i> and <i>BMPR1A</i> gene variants may be considered medically necessary when any one of the following major criteria (solid bullets) is met: a) Patients with a clinical diagnosis of juvenile polyposis syndrome based on the presence of any one of the following: i. at least 3 to 5 juvenile polyps in the colon ii. multiple juvenile polyps in other parts of the gastrointestinal tract iii. any number of juvenile polyps in a person with a known family history of juvenile polyps. b) At-risk relative of a patient suspected of or diagnosed with juvenile polyposis syndrome. G. <i>STK11</i> Testing 1. Genetic testing for <i>STK11</i> gene variants may be considered medically necessary when any one of the following major criteria (solid bullets) is met: a) Patients with a clinical diagnosis of Peutz-Jeghers syndrome based on the presence of any 2 of the following: i. presence of 2 or more histologically confirmed Peutz-Jeghers polyps of the small intestine ii. characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, or fingers iii. family history of Peutz-Jeghers syndrome b) At-risk relative of a patient suspected of or diagnosed with Peutz-Jeghers syndrome. H. Genetic testing for all other gene variants for Lynch syndrome or CRC is considered experimental / investigational.”</p> <ul style="list-style-type: none"> ▪ Updated Policy Guidelines.
	Updated Rationale section.
	<p>In Coding section:</p> <ul style="list-style-type: none"> ▪ Removed CPT codes: 81401, 88341, 88342, 88344. ▪ Updated coding bullets.
	Updated References section.
	Removed Appendix section.
04-24-2019	<p>In Policy section:</p> <ul style="list-style-type: none"> ▪ In Item A 1 e, removed “or Revised Bethesda” to read, “Patients without CRC but with a family history meeting the Amsterdam criteria, when no affected family members have been tested for MMR variants.” ▪ In Item D 1 c, removed “or Revised Bethesda” and “and when sequencing for MMR variants is negative” to read, “Patients without CRC but with a family history meeting the Amsterdam criteria, when no affected family members have been tested for MMR variants.”

REFERENCES

1. Vogt S, Jones N, Christian D, et al. Expanded extracolonic tumor spectrum in MUTYH-associated polyposis. *Gastroenterology*. Dec 2009;137(6):1976-1985 e1971-1910. PMID 19732775

2. Balmana J, Castells A, Cervantes A. Familial colorectal cancer risk: ESMO Clinical Practice Guidelines. *Ann Oncol*. May 2010;21(Suppl 5):v78-81. PMID 20555108
3. Gala M, Chung DC. Hereditary colon cancer syndromes. *Semin Oncol*. Aug 2011;38(4):490-499. PMID 21810508
4. Quehenberger F, Vasen HF, van Houwelingen HC. Risk of colorectal and endometrial cancer for carriers of mutations of the hMLH1 and hMSH2 gene: correction for ascertainment. *J Med Genet*. Jun 2005;42(6):491-496. PMID 15937084
5. Guindalini RS, Win AK, Gulden C, et al. Mutation spectrum and risk of colorectal cancer in African American families with Lynch syndrome. *Gastroenterology*. Nov 2015;149(6):1446-1453. PMID 26248088
6. Sinn DH, Chang DK, Kim YH, et al. Effectiveness of each Bethesda marker in defining microsatellite instability when screening for Lynch syndrome. *Hepatogastroenterology*. May-Jun 2009;56(91-92):672-676. PMID 19621678
7. Wu Y, Berends MJ, Mensink RG, et al. Association of hereditary nonpolyposis colorectal cancer-related tumors displaying low microsatellite instability with MSH6 germline mutations. *Am J Hum Genet*. Nov 1999;65(5):1291-1298. PMID 10521294
8. Goel A, Nagasaka T, Spiegel J, et al. Low frequency of Lynch syndrome among young patients with non-familial colorectal cancer. *Clin Gastroenterol Hepatol*. Nov 2010;8(11):966-971. PMID 20655395
9. Palomaki GE, McClain MR, Melillo S, et al. EGAPP supplementary evidence review: DNA testing strategies aimed at reducing morbidity and mortality from Lynch syndrome. *Genet Med*. Jan 2009;11(1):42-65. PMID 19125127
10. Bouzourene H, Hutter P, Losi L, et al. Selection of patients with germline MLH1 mutated Lynch syndrome by determination of MLH1 methylation and BRAF mutation. *Fam Cancer*. Jun 2010;9(2):167-172. PMID 19949877
11. Niessen RC, Hofstra RM, Westers H, et al. Germline hypermethylation of MLH1 and EPCAM deletions are a frequent cause of Lynch syndrome. *Genes Chromosomes Cancer*. Aug 2009;48(8):737-744. PMID 19455606
12. Kloor M, Voigt AY, Schackert HK, et al. Analysis of EPCAM protein expression in diagnostics of Lynch syndrome. *J Clin Oncol*. Jan 10 2011;29(2):223-227. PMID 21115857
13. Kuiper RP, Vissers LE, Venkatachalam R, et al. Recurrence and variability of germline EPCAM deletions in Lynch syndrome. *Hum Mutat*. Apr 2011;32(4):407-414. PMID 21309036
14. Kovacs ME, Papp J, Szentirmay Z, et al. Deletions removing the last exon of TACSTD1 constitute a distinct class of mutations predisposing to Lynch syndrome. *Hum Mutat*. Feb 2009;30(2):197-203. PMID 19177550
15. Ligtenberg MJ, Kuiper RP, Chan TL, et al. Heritable somatic methylation and inactivation of MSH2 in families with Lynch syndrome due to deletion of the 3' exons of TACSTD1. *Nat Genet*. Jan 2009;41(1):112-117. PMID 19098912
16. Rumilla K, Schowalter KV, Lindor NM, et al. Frequency of deletions of EPCAM (TACSTD1) in MSH2-associated lynch syndrome cases. *J Mol Diagn*. Jan 2011;13(1):93-99. PMID 21227399
17. Hesson LB, Hitchins MP, Ward RL. Epimutations and cancer predisposition: importance and mechanisms. *Curr Opin Genet Dev*. Jun 2010;20(3):290-298. PMID 20359882
18. Hitchins MP. Inheritance of epigenetic aberrations (constitutional epimutations) in cancer susceptibility. *Adv Genet*. Oct 2010;70:201-243. PMID 20920750
19. Vasen HF, Watson P, Mecklin JP, et al. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. *Gastroenterology*. Jun 1999;116(6):1453-1456. PMID 10348829
20. Umar A, Boland CR, Terdiman JP, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. *J Natl Cancer Inst*. Feb 18 2004;96(4):261-268. PMID 14970275
21. Kastrinos F, Uno H, Ukaegbu C, et al. Development and validation of the PREMM5 model for comprehensive risk assessment of Lynch syndrome. *J Clin Oncol*. Jul 01 2017;35(19):2165-2172. PMID 28489507

22. Latchford AR, Neale K, Phillips RK, et al. Juvenile polyposis syndrome: a study of genotype, phenotype, and long-term outcome. *Dis Colon Rectum*. Oct 2012;55(10):1038-1043. PMID 22965402
23. Howe JR, Roth S, Ringold JC, et al. Mutations in the SMAD4/DPC4 gene in juvenile polyposis. *Science*. May 15 1998;280(5366):1086-1088. PMID 9582123
24. Fogt F, Brown CA, Badizadegan K, et al. Low prevalence of loss of heterozygosity and SMAD4 mutations in sporadic and familial juvenile polyposis syndrome-associated juvenile polyps. *Am J Gastroenterol*. Oct 2004;99(10):2025-2031. PMID 15447767
25. Burger B, Uhlhaas S, Mangold E, et al. Novel de novo mutation of MADH4/SMAD4 in a patient with juvenile polyposis. *Am J Med Genet*. Jul 1 2002;110(3):289-291. PMID 12116240
26. Syngal S, Brand RE, Church JM, et al. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. *Am J Gastroenterol*. Feb 2015;110(2):223-262; quiz 263. PMID 25645574
27. Grotsky HW, Rickert RR, Smith WD, et al. Familial juvenile polyposis coli. A clinical and pathologic study of a large kindred. *Gastroenterology*. Mar 1982;82(3):494-501. PMID 7054044
28. Schreiber IR, Baker M, Amos C, et al. The hamartomatous polyposis syndromes: a clinical and molecular review. *Am J Gastroenterol*. Feb 2005;100(2):476-490. PMID 15667510
29. Brosens LA, van Hattem A, Hylind LM, et al. Risk of colorectal cancer in juvenile polyposis. *Gut*. Jul 2007;56(7):965-967. PMID 17303595
30. Gallione CJ, Repetto GM, Legius E, et al. A combined syndrome of juvenile polyposis and hereditary haemorrhagic telangiectasia associated with mutations in MADH4 (SMAD4). *Lancet*. Mar 13 2004;363(9412):852-859. PMID 15031030
31. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High-Risk Assessment: Colorectal. Version 1.2018. http://www.nccn.org/professionals/physician_gls/pdf/genetics_colon.pdf. Accessed August 15, 2018.
32. Olschwang S, Markie D, Seal S, et al. Peutz-Jeghers disease: most, but not all, families are compatible with linkage to 19p13.3. *J Med Genet*. Jan 1998;35(1):42-44. PMID 9475093
33. Jenne DE, Reimann H, Nezu J, et al. Peutz-Jeghers syndrome is caused by mutations in a novel serine threonine kinase. *Nat Genet*. Jan 1998;18(1):38-43. PMID 9425897
34. Hemminki A, Markie D, Tomlinson I, et al. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. *Nature*. Jan 8 1998;391(6663):184-187. PMID 9428765
35. Hernan I, Roig I, Martin B, et al. De novo germline mutation in the serine-threonine kinase STK11/LKB1 gene associated with Peutz-Jeghers syndrome. *Clin Genet*. Jul 2004;66(1):58-62. PMID 15200509
36. van Lier MG, Wagner A, Mathus-Vliegen EM, et al. High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. *Am J Gastroenterol*. Jun 2010;105(6):1258-1264; author reply 1265. PMID 20051941
37. Blue Cross and Blue Shield Association Technology Evaluation Center (TEC). Genetic Testing for Inherited Susceptibility to Colorectal Cancer: Part I – Adenomatous Polyposis Coli Gene Mutations. *TEC Assessments*. 1998;Volume 13:Tab 10. PMID
38. Jasperson KW, Patel SG, Ahnen DJ. APC-Associated Polyposis Conditions. In: Pagon RA, Adam MP, Ardinger HH, et al., eds. *GeneReviews*. Seattle, WA: University of Washington; 2017.
39. Kastrinos F, Syngal S. Recently identified colon cancer predispositions: MYH and MSH6 mutations. *Semin Oncol*. Oct 2007;34(5):418-424. PMID 17920897
40. Lefevre JH, Parc Y, Svrcek M, et al. APC, MYH, and the correlation genotype-phenotype in colorectal polyposis. *Ann Surg Oncol*. Apr 2009;16(4):871-877. PMID 19169759
41. Avezzu A, Agostini M, Pucciarelli S, et al. The role of MYH gene in genetic predisposition to colorectal cancer: another piece of the puzzle. *Cancer Lett*. Sep 18 2008;268(2):308-313. PMID 18495334
42. Balaguer F, Castellvi-Bel S, Castells A, et al. Identification of MYH mutation carriers in colorectal cancer: a multicenter, case-control, population-based study. *Clin Gastroenterol Hepatol*. Mar 2007;5(3):379-387. PMID 17368238
43. Lagarde A, Rouleau E, Ferrari A, et al. Germline APC mutation spectrum derived from 863 genomic variations identified through a 15-year medical genetics service to French patients with FAP. *J Med Genet*. Oct 2010;47(10):721-722. PMID 20685668

44. Aretz S, Stienen D, Uhlhaas S, et al. Large submicroscopic genomic APC deletions are a common cause of typical familial adenomatous polyposis. *J Med Genet.* Feb 2005;42(2):185-192. PMID 15689459
45. Bunyan DJ, Eccles DM, Sillibourne J, et al. Dosage analysis of cancer predisposition genes by multiplex ligation-dependent probe amplification. *Br J Cancer.* Sep 13 2004;91(6):1155-1159. PMID 15475941
46. Aretz S, Genuardi M, Hes FJ. Clinical utility gene card for: MUTYH-associated polyposis (MAP), autosomal recessive colorectal adenomatous polyposis, multiple colorectal adenomas, multiple adenomatous polyps (MAP) - update 2012. *Eur J Hum Genet.* Jan 2013;21(1). PMID 22872101
47. Inra JA, Steyerberg EW, Grover S, et al. Racial variation in frequency and phenotypes of APC and MUTYH mutations in 6,169 individuals undergoing genetic testing. *Genet Med.* Oct 2015;17(10):815-821. PMID 25590978
48. Out AA, Tops CM, Nielsen M, et al. Leiden Open Variation Database of the MUTYH gene. *Hum Mutat.* Nov 2010;31(11):1205-1215. PMID 20725929
49. Nielsen M, Lynch H, Infante E, et al. MUTYH-Associated Polyposis. In: Pagon RA, Adam MP, Ardinger HH, eds. *GeneReviews* Seattle, WA: University of Washington; 2012.
50. Sieber OM, Lamlum H, Crabtree MD, et al. Whole-gene APC deletions cause classical familial adenomatous polyposis, but not attenuated polyposis or "multiple" colorectal adenomas. *Proc Natl Acad Sci U S A.* Mar 05 2002;99(5):2954-2958. PMID 11867715
51. Aretz S, Uhlhaas S, Goergens H, et al. MUTYH-associated polyposis: 70 of 71 patients with biallelic mutations present with an attenuated or atypical phenotype. *Int J Cancer.* Aug 15 2006;119(4):807-814. PMID 16557584
52. Michils G, Tejpar S, Thoelen R, et al. Large deletions of the APC gene in 15% of mutation-negative patients with classical polyposis (FAP): a Belgian study. *Hum Mutat.* Feb 2005;25(2):125-134. PMID 15643602
53. Truta B, Allen BA, Conrad PG, et al. A comparison of the phenotype and genotype in adenomatous polyposis patients with and without a family history. *Fam Cancer.* Jun 2005;4(2):127-133. PMID 15951963
54. Bjork JA, Akerbrant HI, Iselius LE, et al. Risk factors for rectal cancer morbidity and mortality in patients with familial adenomatous polyposis after colectomy and ileorectal anastomosis. *Dis Colon Rectum.* Dec 2000;43(12):1719-1725. PMID 11156457
55. Jarvinen HJ. Epidemiology of familial adenomatous polyposis in Finland: impact of family screening on the colorectal cancer rate and survival. *Gut.* Mar 1992;33(3):357-360. PMID 1314763
56. Vasen HF, Griffioen G, Offerhaus GJ, et al. The value of screening and central registration of families with familial adenomatous polyposis. A study of 82 families in The Netherlands. *Dis Colon Rectum.* Mar 1990;33(3):227-230. PMID 2155763
57. Bonis PA, Trikalinos TA, Chung M, et al. *Hereditary Nonpolyposis Colorectal Cancer: Diagnostic Strategies and Their Implications (Evidence Report/Technology Assessment No. 150)*. Rockville, MD: Agency for Healthcare Research and Quality; 2007.
58. Evaluation of Genomic Applications in Practice and Prevention (EGAPP) Working Group. Recommendations from the EGAPP Working Group: genetic testing strategies in newly diagnosed individuals with colorectal cancer aimed at reducing morbidity and mortality from Lynch syndrome in relatives. *Genet Med.* Jan 2009;11(1):35-41. PMID 19125126
59. Moreira L, Balaguer F, Lindor N, et al. Identification of Lynch syndrome among patients with colorectal cancer. *JAMA.* Oct 17 2012;308(15):1555-1565. PMID 23073952
60. Kempers MJ, Kuiper RP, Ockeloen CW, et al. Risk of colorectal and endometrial cancers in EPCAM deletion-positive Lynch syndrome: a cohort study. *Lancet Oncol.* Jan 2011;12(1):49-55. PMID 21145788
61. Jin M, Hampel H, Zhou X, et al. BRAF V600E mutation analysis simplifies the testing algorithm for Lynch syndrome. *Am J Clin Pathol.* Aug 2013;140(2):177-183. PMID 23897252
62. Capper D, Voigt A, Bozukova G, et al. BRAF V600E-specific immunohistochemistry for the exclusion of Lynch syndrome in MSI-H colorectal cancer. *Int J Cancer.* Oct 1 2013;133(7):1624-1630. PMID 23553055

63. Kastrinos F, Syngal S. Screening patients with colorectal cancer for Lynch syndrome: what are we waiting for? *J Clin Oncol*. Apr 1 2012;30(10):1024-1027. PMID 22355054
64. Bonadona V, Bonaiti B, Olschwang S, et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. *JAMA*. Jun 8 2011;305(22):2304-2310. PMID 21642682
65. Leenen CH, van Lier MG, van Doorn HC, et al. Prospective evaluation of molecular screening for Lynch syndrome in patients with endometrial cancer \leq 70 years. *Gynecol Oncol*. May 2012;125(2):414-420. PMID 22306203
66. Hampel H, Frankel WL, Martin E, et al. Screening for the Lynch syndrome (hereditary nonpolyposis colorectal cancer). *N Engl J Med*. May 05 2005;352(18):1851-1860. PMID 15872200
67. Aktan-Collan K, Mecklin JP, Jarvinen H, et al. Predictive genetic testing for hereditary non-polyposis colorectal cancer: uptake and long-term satisfaction. *Int J Cancer*. Jan 20 2000;89(1):44-50. PMID 10719730
68. Aktan-Collan K, Haukkala A, Pylvanainen K, et al. Direct contact in inviting high-risk members of hereditary colon cancer families to genetic counselling and DNA testing. *J Med Genet*. Nov 2007;44(11):732-738. PMID 17630403
69. Stanley AJ, Gaff CL, Aittomaki AK, et al. Value of predictive genetic testing in management of hereditary non-polyposis colorectal cancer (HNPCC). *Med J Aust*. Apr 03 2000;172(7):313-316. PMID 10844916
70. Hadley DW, Jenkins J, Dimond E, et al. Genetic counseling and testing in families with hereditary nonpolyposis colorectal cancer. *Arch Intern Med*. Mar 10 2003;163(5):573-582. PMID 12622604
71. Lerman C, Hughes C, Trock BJ, et al. Genetic testing in families with hereditary nonpolyposis colon cancer. *JAMA*. May 05 1999;281(17):1618-1622. PMID 10235155
72. Codori AM, Petersen GM, Miglioretti DL, et al. Attitudes toward colon cancer gene testing: factors predicting test uptake. *Cancer Epidemiol Biomarkers Prev*. Apr 1999;8(4 Pt 2):345-351. PMID 10207639
73. Schmeler KM, Lynch HT, Chen LM, et al. Prophylactic surgery to reduce the risk of gynecologic cancers in the Lynch syndrome. *N Engl J Med*. Jan 19 2006;354(3):261-269. PMID 16421367
74. Fitzgibbons RJ, Jr., Lynch HT, Stanislav GV, et al. Recognition and treatment of patients with hereditary nonpolyposis colon cancer (Lynch syndromes I and II). *Ann Surg*. Sep 1987;206(3):289-295. PMID 3632093
75. Burke W, Petersen G, Lynch P, et al. Recommendations for follow-up care of individuals with an inherited predisposition to cancer. I. Hereditary nonpolyposis colon cancer. Cancer Genetics Studies Consortium. *JAMA*. Mar 19 1997;277(11):915-919. PMID 9062331
76. Van Dalen R, Church J, McGannon E, et al. Patterns of surgery in patients belonging to amsterdam-positive families. *Dis Colon Rectum*. May 2003;46(5):617-620. PMID 12792437
77. de Vos tot Nederveen Cappel WH, Buskens E, van Duijvendijk P, et al. Decision analysis in the surgical treatment of colorectal cancer due to a mismatch repair gene defect. *Gut*. Dec 2003;52(12):1752-1755. PMID 14633956
78. Guillem JG, Wood WC, Moley JF, et al. ASCO/SSO review of current role of risk-reducing surgery in common hereditary cancer syndromes. *J Clin Oncol*. Oct 1 2006;24(28):4642-4660. PMID 17008706
79. Yurgelun MB, Mercado R, Rosenblatt M, et al. Impact of genetic testing on endometrial cancer risk-reducing practices in women at risk for Lynch syndrome. *Gynecol Oncol*. Dec 2012;127(3):544-551. PMID 22940489
80. Engel C, Rahner N, Schulmann K, et al. Efficacy of annual colonoscopic surveillance in individuals with hereditary nonpolyposis colorectal cancer. *Clin Gastroenterol Hepatol*. Feb 2010;8(2):174-182. PMID 19835992
81. Järvinen HJ, Renkonen-Sinisalo L, Aktan-Collan K, et al. Ten years after mutation testing for Lynch syndrome: cancer incidence and outcome in mutation-positive and mutation-negative family members. *J Clin Oncol*. Oct 01 2009;27(28):4793-4797. PMID 19720893
82. Dove-Edwin I, Sasieni P, Adams J, et al. Prevention of colorectal cancer by colonoscopic surveillance in individuals with a family history of colorectal cancer: 16 year, prospective, follow-up study. *BMJ*. Nov 05 2005;331(7524):1047. PMID 16243849

83. de Vos tot Nederveen Cappel WH, Nagengast FM, Griffioen G, et al. Surveillance for hereditary nonpolyposis colorectal cancer: a long-term study on 114 families. *Dis Colon Rectum*. Dec 2002;45(12):1588-1594. PMID 12473880
84. Syngal S, Weeks JC, Schrag D, et al. Benefits of colonoscopic surveillance and prophylactic colectomy in patients with hereditary nonpolyposis colorectal cancer mutations. *Ann Intern Med*. Nov 15 1998;129(10):787-796. PMID 9841584
85. Järvinen HJ, Mecklin JP, Sistonen P. Screening reduces colorectal cancer rate in families with hereditary nonpolyposis colorectal cancer. *Gastroenterology*. May 1995;108(5):1405-1411. PMID 7729632
86. Järvinen HJ, Aarnio M, Mustonen H, et al. Controlled 15-year trial on screening for colorectal cancer in families with hereditary nonpolyposis colorectal cancer. *Gastroenterology*. May 2000;118(5):829-834. PMID 10784581
87. Kwon JS, Scott JL, Gilks CB, et al. Testing women with endometrial cancer to detect Lynch syndrome. *J Clin Oncol*. Jun 1 2011;29(16):2247-2252. PMID 21537049
88. Obermair A, Youlden DR, Young JP, et al. Risk of endometrial cancer for women diagnosed with HNPCC-related colorectal carcinoma. *Int J Cancer*. Dec 1 2010;127(11):2678-2684. PMID 20533284
89. Grandval P, Baert-Desurmont S, Bonnet F, et al. Colon-specific phenotype in Lynch syndrome associated with EPCAM deletion. *Clin Genet*. Jul 2012;82(1):97-99. PMID 22243433
90. Lynch HT, Riegert-Johnson DL, Snyder C, et al. Lynch syndrome-associated extracolonic tumors are rare in two extended families with the same EPCAM deletion. *Am J Gastroenterol*. Oct 2011;106(10):1829-1836. PMID 21769135
91. Auranen A, Joutsiniemi T. A systematic review of gynecological cancer surveillance in women belonging to hereditary nonpolyposis colorectal cancer (Lynch syndrome) families. *Acta Obstet Gynecol Scand*. May 2011;90(5):437-444. PMID 21306348
92. Yang HR, Ko JS, Seo JK. Germline mutation analysis of STK11 gene using direct sequencing and multiplex ligation-dependent probe amplification assay in Korean children with Peutz-Jeghers syndrome. *Dig Dis Sci*. Dec 2010;55(12):3458-3465. PMID 20393878
93. Calva-Cerqueira D, Chinnathambi S, Pechman B, et al. The rate of germline mutations and large deletions of SMAD4 and BMPR1A in juvenile polyposis. *Clin Genet*. Jan 2009;75(1):79-85. PMID 18823382
94. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic deletions and a genotype phenotype update in 80 unrelated families with juvenile polyposis syndrome. *J Med Genet*. Nov 2007;44(11):702-709. PMID 17873119
95. Volikos E, Robinson J, Aittomaki K, et al. LKB1 exonic and whole gene deletions are a common cause of Peutz-Jeghers syndrome. *J Med Genet*. May 2006;43(5):e18. PMID 16648371
96. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic STK11 deletions in Peutz-Jeghers syndrome. *Hum Mutat*. Dec 2005;26(6):513-519. PMID 16287113
97. Aytac E, Sulu B, Heald B, et al. Genotype-defined cancer risk in juvenile polyposis syndrome. *Br J Surg*. Jan 2015;102(1):114-118. PMID 25389115
98. Resta N, Pierannunzio D, Lenato GM, et al. Cancer risk associated with STK11/LKB1 germline mutations in Peutz-Jeghers syndrome patients: results of an Italian multicenter study. *Dig Liver Dis*. Jul 2013;45(7):606-611. PMID 23415580
99. Salloch H, Reinacher-Schick A, Schulmann K, et al. Truncating mutations in Peutz-Jeghers syndrome are associated with more polyps, surgical interventions and cancers. *Int J Colorectal Dis*. Jan 2010;25(1):97-107. PMID 19727776
100. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Uterine Neoplasms. Version 2.2018. http://www.nccn.org/professionals/physician_gls/pdf/uterine.pdf. Accessed August 16, 2018.
101. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colon Cancer. Version 3.2018. http://www.nccn.org/professionals/physician_gls/pdf/colon.pdf. Accessed August 16, 2018.
102. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colorectal Cancer Screening. Version 1.2018.

http://www.nccn.org/professionals/physician_gls/pdf/colorectal_screening.pdf. Accessed August 16, 2018.

103. Kastrinos F, Steyerberg EW, Mercado R, et al. The PREMM(1,2,6) model predicts risk of MLH1, MSH2, and MSH6 germline mutations based on cancer history. *Gastroenterology*. Jan 2011;140(1):73-81. PMID 20727894
104. Stoffel EM, Mangu PB, Gruber SB, et al. Hereditary colorectal cancer syndromes: American Society of Clinical Oncology Clinical Practice Guideline endorsement of the familial risk-colorectal cancer: European Society for Medical Oncology Clinical Practice Guidelines. *J Clin Oncol*. Jan 10 2015;33(2):209-217. PMID 25452455

Other References

1. Blue Cross and Blue Shield of Kansas Surgery Liaison, August 2010; August 2011; August 2018; January 2019.