



Title: Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes

Related Policies:	■ Somatic Biomarker Testing (Including Liquid Biopsy) for Targeted
	Treatment in Metastatic Colorectal Cancer (KRAS, NRAS, BRAF, NTRK and
	HER2)

Professional / Institutional	
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Populations	Interventions	Comparators	Outcomes
Individuals:	Interventions of	Comparators of	Relevant outcomes
Who are suspected of	interest are:	interest are:	include:
attenuated FAP, MAP, and	 Genetic testing for 	 No genetic 	Overall survival
Lynch syndrome, or are at-risk	APC	testing	 Disease-specific
relatives of patients with FAP			survival
			Test accuracy
			Test validity
Individuals:	Interventions of	Comparators of	Relevant outcomes
Who are suspected of	interest are:	interest are:	include:
attenuated FAP, MAP, and	 Genetic testing for 	 No genetic 	Overall survival
Lynch syndrome	MUTYH after a	testing	

Populations	Interventions	Comparators	Outcomes
	negative <i>APC</i> test result		Disease-specific survivalTest accuracyTest validity
Individuals: • Who are suspected of attenuated FAP, MAP, and Lynch syndrome; CRC; or endometrial cancer meeting clinical criteria for Lynch	Interventions of interest are: • Genetic testing for MMR genes	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity
Individuals: • Who are at-risk relatives of patients with Lynch or family history meeting appropriate criteria, but do not have CRC	Interventions of interest are: • Genetic testing for MMR genes	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity
Individuals: • Who warrant Lynch testing, screen negative on MMR testing, but positive for microsatellite instability (MSI) and lack MSH2 protein expression	Interventions of interest are: • Genetic testing for EPCAM variants	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity
Individuals: • With CRC in whom MLH1 protein is not expressed on immunohistochemical analysis	Interventions of interest are: • Genetic testing for BRAF V600E or MLH1 promoter methylation	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity
 Individuals: Who are suspected of juvenile polyposis syndrome or are atrisk relatives of patients suspected of or diagnosed with JPS 	Interventions of interest are: • Genetic testing for SMAD4 and BMPR1A genes	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity
 Individuals: Who are suspected of Peutz- Jeghers syndrome or are at- risk relatives of patients suspected of or diagnosed with PJS 	Interventions of interest are: • Genetic testing for STK11 gene	Comparators of interest are: No genetic testing	Relevant outcomes include: Overall survival Disease-specific survival Test accuracy Test validity

DESCRIPTION

Genetic testing is available for both those with and those at risk for various types of hereditary cancer. This review evaluates genetic testing for hereditary colorectal cancer (CRC) and polyposis syndromes, including familial adenomatous polyposis (FAP), Lynch syndrome (formerly known as hereditary nonpolyposis colorectal cancer), *MUTYH*-associated polyposis (MAP), Lynch syndrome-related endometrial cancer, juvenile polyposis syndrome (JPS), and Peutz-Jeghers syndrome (PJS)

OBJECTIVE

The objective of this evidence review is to assess whether the use of genetic testing improves the net health outcome in patients with Lynch syndrome and other inherited colon cancer syndromes.

BACKGROUND

I. Hereditary Colorectal Cancers

Currently, 2 types of hereditary colorectal cancers (CRCs) are well-defined: familial adenomatous polyposis (FAP) and Lynch syndrome (formerly hereditary nonpolyposis CRC). Lynch syndrome has been implicated in some endometrial cancers as well.

II. Familial Adenomatous Polyposis and Associated Variants

Familial adenomatous polyposis typically develops by age 16 years and can be identified by the appearance of hundreds to thousands of characteristic, precancerous colon polyps. If left untreated, all affected individuals will develop CRC. The mean age of colon cancer diagnosis in untreated individuals is 39 years. The condition accounts for about 1% of CRC and may also be associated with osteomas of the jaw, skull, and limbs; sebaceous cysts; and pigmented spots on the retina referred to as congenital hypertrophy of the retinal pigment epithelium. Familial adenomatous polyposis associated with these collective extraintestinal manifestations is sometimes referred to as Gardner syndrome. This condition may also be related to central nervous system tumors, referred to as Turcot syndrome.

Germline variants in the adenomatous polyposis coli (*APC*) gene, located on chromosome 5, are responsible for FAP and are inherited in an autosomal dominant manner. Variants in the *APC* gene result in altered protein length in about 80% to 85% of cases of FAP. A specific *APC* gene variant (I1307K) has been found in Ashkenazi Jewish descendants, which may explain a portion of the familial CRC occurring in this population.

A subset of FAP patients may have an attenuated form of FAP, typically characterized by fewer than 100 cumulative colorectal adenomas occurring later in life than in classical FAP. In the attenuated form of FAP, CRC occurs at an average age of 50 to 55 years, but the lifetime risk of CRC remains high (>70% by age 80 years). The risk of extraintestinal cancer is also lower but cumulative lifetime risk remains high (>38%) compared with the general population. Only 30% or fewer of attenuated FAP patients have *APC* variants; some of these patients have variants in the *MUTYH* (formerly *MYH*) gene, and this form of the condition is called *MUTYH*-associated polyposis (MAP). This form of polyposis occurs with a frequency similar to FAP, with some variability among prevalence estimates for both. While clinical features of MAP are similar to FAP or attenuated FAP, a strong multigenerational family history of polyposis is absent.

Biallelic *MUTYH* variants are associated with a cumulative CRC risk of about 80% by age 70, whereas the monoallelic *MUTYH* variant-associated risk of CRC appears to be relatively minimal, although still under debate.^{2,} Thus, inheritance for high-risk CRC predisposition is autosomal recessive in contrast to FAP. When relatively few (ie, between 10 and 99) adenomas are present, and family history is unavailable, the differential diagnosis may include both MAP and Lynch syndrome; genetic testing in this situation could include *APC*, *MUTYH* if *APC* is negative for variants, and screening for variants associated with Lynch syndrome.

It is important to distinguish between classical FAP, attenuated FAP, and MAP (mono- or biallelic) by genetic analysis because recommendations for patient surveillance and cancer prevention vary by syndrome.³,

III. Testing

Genetic testing for APC variants may be considered in the following situations:

- Patients at high-risk, such as those with a family member who tested positive for FAP and have a known *APC* variant.
- Patients undergoing differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome. These patients do not meet the clinical diagnostic criteria for classical FAP and have few adenomatous colonic polyps.
- To confirm FAP in patients with colon cancer with a clinical picture or family history consistent with classical FAP.

IV. Lynch Syndrome

Lynch syndrome is an inherited disorder that results in a higher predisposition to CRC and other malignancies including endometrial and gastric cancer. Lynch syndrome is estimated to account for 3% to 5% of all CRC. People with Lynch syndrome have a 70% to 80% lifetime risk of developing any type of cancer. ^{4,5}, However, the risk varies by genotype. It occurs as a result of germline variants in the mismatch repair (MMR) genes that include *MLH1*, *MSH2*, *MSH6*, and *PMS2*. In approximately 80% of cases, the variants are located in the *MLH1* and *MSH2* genes, while 10% to 12% of variants are located in the *MSH6* gene, and 2% to 3% in the *PMS2* gene. Additionally, variants in 3 additional genes (*MLH3*, *PMS1*, *EX01*) have been implicated with Lynch Syndrome. Notably, in individuals meeting the various clinical criteria for Lynch syndrome, 50% of individuals have a variant in the *MLH1*, *MSH2*, *MSH6*, and *PMS2* genes. The lifetime risk of CRC is nearly 80% in individuals carrying a variant in 1 of these genes.

V. Testing

Preliminary screening of tumor tissue does not identify MMR gene variants but is used to guide subsequent diagnostic testing via DNA analysis for specific variants. Genetic testing or DNA analysis (gene sequencing, deletion, and duplication testing) for the MMR genes involves assessment for *MLH1*, *MSH2*, *MSH6*, and *PMS2* variants. The following are 3 testing strategies.

- Microsatellite instability (MSI) testing (phenotype): Individuals with high MSI either proceed to genetic testing for MLH1, MSH2, MSH6, and PMS2 or to immunohistochemical (IHC) testing.
- IHC testing (phenotype): Individuals with negative staining would proceed to genetic testing for *MLH1*, *MSH2*, *MSH6*, and *PMS2*.

 Modification strategy: Tumor tissue of patients with negative staining for *MLH1* on IHC is tested for the *BRAF* V600E variant to determine methylation status. If the *BRAF* variant is not detected, the individual receives *MLH1* DNA analysis.

The phenotype tests used to identify individuals who may be at a high risk of Lynch syndrome are explained next. The first screening test measures MSI. As a result of variance in the MMR gene family, the MMR protein is either absent or deficient, resulting in an inability to correct DNA replication errors causing MSI. Approximately 80% to 90% of Lynch syndrome CRC tumors have MSI. The National Cancer Institute has recommended screening for 5 markers to detect MSI (Bethesda markers). Microsatellite instability detection in 2 of these markers is considered a positive result or "high probability of MSI."⁶,

The second phenotype screening test is IHC, which involves the staining of tumor tissue for the presence of 4 MMR proteins (MLH1, MSH2, MSH6, PMS2). The absence of 1 or more of these proteins is considered abnormal.

BRAF testing is an optional screening method that may be used in conjunction with IHC testing for *MLH1* to improve efficiency. Methylation analysis of the *MLH1* gene can largely substitute for *BRAF* testing, or be used in combination to improve efficiency slightly.

Both MSI and IHC have a 5% to 10% false-negative rate. Microsatellite instability testing performance depends on the specific MMR variant. Screening with MSI has a sensitivity of about 89% for *MLH1* and *MSH2* and 77% for *MSH6* and a specificity of about 90% for each. The specificity of MSI testing is low because approximately 10% of sporadic CRCs are MSI-positive due to somatic hypermethylation of the *MLH1* promoter. Additionally, some tumors positive for *MSH6* variants are associated with the MSI-low phenotype rather than MSI-high; thus MSI-low should not be a criterion against proceeding to MMR variant testing.^{7,8}, IHC screening has a sensitivity for *MLH1*, *MSH2*, and *MSH6* of about 83% and a specificity of about 90% for each.

Screening of tumor tissue from patients enables genetic testing for a definitive diagnosis of Lynch syndrome and leads to counseling, cancer surveillance (eg, through frequent colonoscopic or endometrial screening examinations), and prophylaxis (eg, risk-reducing colorectal or gynecologic surgeries) for CRC patients, as well as for their family members.

Genetic testing for an MMR gene variant is often limited to *MLH1* and *MSH2* and, if negative, then *MSH6* and *PMS2*. The *BRAF* gene is often mutated in CRC when a particular *BRAF* variant (*V600E*, a change from valine to glutamic acid at amino acid position 600 in the BRAF protein) is present. To date, no *MLH1* gene variants have been reported.^{9,} Therefore, patients negative for MLH1 protein expression by IHC, and therefore potentially positive for an *MLH1* variant, could first be screened for a *BRAF* variant. *BRAF*-positive samples need not be further tested by *MLH1* sequencing. *MLH1* gene methylation largely correlates with the presence of *BRAF* V600E and, in combination with *BRAF* testing, can accurately separate Lynch from sporadic CRC in IHC *MLH1*-negative cases.^{10,}

Novel deletions have been reported to affect the expression of the *MSH2* gene in the absence of an *MSH2* gene variant, and thereby cause Lynch syndrome. In these cases, deletions in *EPCAM*, the gene for the epithelial cell adhesion molecule, are responsible. *EPCAM* testing has been

added to many Lynch syndrome profiles and is conducted only when tumor tissue screening results are MSI-high and/or IHC testing shows a lack of *MSH2* expression, but no *MSH2* variant is found by sequencing. *EPCAM* is found just upstream, in a transcriptional sense, of *MSH2*. Deletions of *EPCAM* that encompass the last 2 exons of the *EPCAM* gene, including the polyadenylation signal that normally ends transcription of DNA into messenger RNA, result in transcriptional "read-through" and subsequent hypermethylation of the nearby and downstream *MSH2* promoter. This hypermethylation prevents normal MSH2 protein expression and leads to Lynch syndrome in a fashion similar to Lynch cases in which an *MSH2* variant prevents *MSH2* gene expression.^{11,}

Distinct from patients with *EPCAM* deletions, rare cases of Lynch syndrome have been reported without detectable germline MMR variants, although IHC testing demonstrated a loss of expression of 1 of the MMR proteins. In at least some of these cases, research has identified germline "epivariants," ie, methylation of promoter regions that control the expression of the MMR genes. 11,12,13, Such methylation may be isolated or be in conjunction with a linked genetic alteration near the affected MMR gene. The germline epivariants may arise de novo or may be heritable in Mendelian or non-Mendelian fashion. This is distinct from some cases of MSI-high sporadic CRC wherein the tumor tissue may show *MLH1* promoter methylation and IHC nonexpression, but the same is not true of germline cells. Clinical testing for Lynch syndrome-related germline epivariants is not routine but may help in exceptional cases.

Female patients with Lynch syndrome have a predisposition to endometrial cancer. Lynch syndrome is estimated to account for 2% of all endometrial cancers in women and 10% of endometrial cancers in women younger than 50 years of age. Female carriers of the germline variants *MLH1*, *MSH2*, *MSH6*, and *PMS2* have an estimated 40% to 62% lifetime risk of developing endometrial cancer, as well as a 4% to 12% lifetime risk of ovarian cancer.

VI. Population Selection

Various attempts have been made to identify which patients with colon cancer should undergo testing for MMR variants, based primarily on family history and related characteristics using criteria such as the Amsterdam II criteria^{14,} (low sensitivity but high specificity), revised Bethesda guidelines^{15,} (better sensitivity but poorer specificity), and risk prediction models (eg, MMRpro; PREMM5; MMRpredict).^{16,} While family history is an important risk factor and should not be discounted in counseling families, it has poor sensitivity and specificity for identifying Lynch syndrome. Based on this and other evidence, the Evaluation of Genomic Applications in Practice and Prevention Working Group recommended testing all newly diagnosed CRC patients for Lynch syndrome, using a screening strategy based on MSI or IHC (with or without *BRAF*) followed by sequencing in screen-positive patients. This recommendation includes genetic testing for the following types of patients:

- Family members of Lynch syndrome patients with a known MMR variant; family members
 would be tested only for the family variant; those testing positive would benefit from early
 and increased surveillance to prevent future CRC.
- Patients with a differential diagnosis of Lynch syndrome versus attenuated FAP versus
- For Lynch syndrome patients, genetic testing of the proband with CRC likely benefits the proband where Lynch syndrome is identified, and appropriate surveillance for associated

malignancies can be initiated and maintained, benefiting family members by identifying the family variant.

VII. Juvenile Polyposis Syndrome

Juvenile polyposis syndrome (JPS) is an autosomal dominant genetic disorder characterized by the presence of multiple hamartomatous (benign) polyps in the digestive tract. It is rare, with an estimated incidence of 1 in 100,000 to 160,000. Generalized JPS refers to polyps in the upper and lower gastrointestinal tract, and juvenile polyposis coli refers to polyps of the colon and rectum. Those with JPS are at a higher risk for CRC and gastric cancer.^{17,} Approximately 60% of patients with JPS have a germline variant in the *BMPR1A* gene or the *SMAD4* gene.^{18,19,} Approximately 25% of patients have de novo variants.^{20,21,} In most cases, polyps appear in the first decade of life and most patients are symptomatic by age 20 years.^{22,} Rectal bleeding is the most common presenting symptom, occurring in more than half of patients. Other presenting symptoms include prolapsing polyp, melena, pain, iron deficiency anemia, and diarrhea.^{17,21,22,}

As noted, individuals with JPS are at increased risk for CRC and gastric cancer. By 35 years of age, the cumulative risk of CRC is 17% to 22%, which increases to 68% by age 60 years. ^{23,24,} The estimated lifetime risk of gastric cancer is 20% to 30%, with a mean age at diagnosis of 58 years. ^{17,21,23,} Juvenile polyposis syndrome may also be associated with hereditary hemorrhagic telangiectasia. ^{25,} The most common clinical manifestations of hereditary hemorrhagic telangiectasia are telangiectasias of the skin and buccal mucosa, epistaxis, and iron deficiency anemia from bleeding.

VIII. Diagnosis

A clinical diagnosis of JPS is made on the basis of the presence of any 1 of the following: at least 5 juvenile polyps in the colon or multiple juvenile polyps in other parts of the gastrointestinal tract or any number of juvenile polyps in a person with a known family history of juvenile polyps. ^{26,} It is recommended that individuals who meet clinical criteria for JPS undergo genetic testing for a germline variant in the *BMPR1A* and *SMAD4* genes for a confirmatory diagnosis of JPS and to counsel at-risk family members. If there is a known *SMAD4* variant in the family, genetic testing should be performed within the first 6 months of life due to hereditary hemorrhagic telangiectasia risk. ^{27,}

IX. Peutz-Jeghers Syndrome

Peutz-Jeghers syndrome (PJS) is also an autosomal dominant genetic disorder, similar to JPS, and is characterized by the presence of multiple hamartomatous (benign) polyps in the digestive tract, mucocutaneous pigmentation, and an increased risk of gastrointestinal and nongastrointestinal cancers. It is rare, with an estimated incidence of 1 in 8000 to 200,000. In most cases, a germline variant in the *STK11* (*LKB1*) gene is responsible for PJS, which has a high penetrance of over 90% by the age of 30 years. ^{28,29,30}, However, 10% to 20% of individuals with PJS have no family history and are presumed to have PJS due to de novo variants. ³¹, A variant in *STK11* is detected in only 50% to 80% of families with PJS, suggesting that there is a second PJS gene locus.

The reported lifetime risk for any cancer is between 37% and 93% among those diagnosed with PJS with an average age of cancer diagnosis at 42 years. The most common sites for malignancy

are the colon and rectum, followed by breast, stomach, small bowel, and pancreas.^{32,} The estimated lifetime risk of gastrointestinal cancer ranges from 38% to 66%.^{32,} Lifetime cancer risk stratified by organ site is colon and rectum (39%), stomach (29%), small bowel (13%), and pancreas (11% to 36%).

X. Diagnosis

A clinical diagnosis of PJS is made if an individual meets 2 or more of the following criteria: presence of 2 or more histologically confirmed PJ polyps of the small intestine or characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, fingers, or family history of PJS.^{26,} Individuals who meet clinical criteria for PJS should undergo genetic testing for a germline variant in the *STK11* gene for a confirmatory diagnosis of PJS and counseling at-risk family members.

REGULATORY STATUS

Clinical laboratories may develop and validate tests in-house and market them as a laboratory service; laboratory-developed tests must meet the general regulatory standards of the Clinical Laboratory Improvement Amendments (CLIA). Genetic tests reviewed in this evidence review are available under the auspices of the CLIA. Laboratories that offer laboratory-developed tests must be licensed by the CLIA for high-complexity testing. To date, the U.S. Food and Drug Administration has chosen not to require any regulatory review of this test.

POLICY

A. MMR Gene Testing

- 1. Genetic testing for MMR genes (*MLH1*, *MSH2*, *MSH6*, *PMS2*) may be considered **medically necessary** in the following individuals:
 - a. Individuals with colorectal cancer (CRC), with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for Lynch syndrome (see Policy Guidelines).
 - Individuals with endometrial cancer with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for Lynch syndrome (see Policy Guidelines section).
 - c. At-risk relatives (see Policy Guidelines) of individuals with Lynch syndrome with a known pathogenic/likely pathogenic MMR gene variant.
 - d. Individuals with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome. Whether testing begins with APC variants or screening for MMR genes depends on clinical presentation.
 - e. Individuals without CRC but with a family history meeting the Amsterdam criteria, or documentation of 5% or higher predicted risk of the syndrome on a validated risk prediction model (e.g. MMRpro, PREMM5 or MMRpredict), when no affected family members have been tested for MMR variants.
 - 2. Testing for germline MMR gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

B. APC Testing

- Genetic testing for adenosis polyposis coli (APC) may be considered medically necessary in the following individuals:
 - a. At-risk relatives (see Policy Guidelines) of individuals with familial adenomatous polyposis (FAP) and/or a known APC variant.
 - b. Individuals with a differential diagnosis of attenuated FAP vs MUTYH-associated polyposis (MAP) vs Lynch syndrome. Whether testing begins with APC variants or screening for mismatch repair (MMR) variants depends on clinical presentation.
- 2. Genetic testing for *APC* gene variants is **not medically necessary** for colorectal cancer patients with classical FAP for confirmation of the FAP diagnosis.
- 3. Testing for germline *APC* gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

C. MUTYH Testing

- 1. Genetic Testing for *MUTYH* gene variants may be considered **medically necessary** in the following individuals:
 - a. Individuals with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome and a negative result for *APC* gene variants. A family history of no parents or children with FAP is consistent with MAP (autosomal recessive).
- 2. Testing for germline *MUTYH* gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

D. *EPCAM* Testing

- 1. Genetic testing for *EPCAM* gene variants may be considered **medically necessary** when any 1 of the following 3 major criteria is met:
 - a. Individuals with CRC, for the diagnosis of Lynch syndrome (see Policy Guidelines section) when:
 - Tumor tissue shows lack of MSH2 protein expression by immunohistochemistry and the individual is negative for a MSH2 germline variant; OR
 - ii. Tumor tissue shows a high level of microsatellite instability and the individual is negative for a germline variant in *MLH1*, *MSH2*, *MSH6*, and *PMS2*; **OR**
 - b. At-risk relatives (see Policy Guidelines section) of individuals with Lynch syndrome with a known pathogenic/likely pathogenic *EPCAM* variant; **OR**
 - c. Individuals without CRC but with a family history meeting the Amsterdam criteria, or documentation of 5% or higher predicted risk of the syndrome on a validated risk prediction model (e.g. MMRpro, PREMM5 or MMRpredict), when no affected family members have been tested for MMR variants, and when sequencing for MMR variants is negative.
- 2. Testing for germline *EPCAM* gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

E. BRAF V600E OR MLH1 PROMOTER METHYLATION

- Somatic Genetic testing for BRAFV600E or MLH1 promoter methylation may be considered medically necessary to exclude a diagnosis of Lynch syndrome when the MLH1 protein is not expressed in a CRC tumor on immunohistochemical analysis.
- 2. Testing for somatic *BRAF V600E* or *MLH1* promoter methylation to exclude a diagnosis of Lynch syndrome is considered **experimental / investigational** in all other situations.

F. SMAD4 AND BMPR1A TESTING

- Genetic testing for SMAD4 and BMPR1A genes may be considered medically necessary when any one of the following major criteria is met:
 - a. Individuals with a clinical diagnosis of juvenile polyposis syndrome based on the presence of any one of the following:
 - i. at least 5 juvenile polyps in the colon
 - ii. multiple juvenile polyps found throughout the gastrointestinal tract
 - iii. any number of juvenile polyps in an individual with a known family history of juvenile polyps.
 - b. At-risk relative of a patient suspected of or diagnosed with juvenile polyposis syndrome.
- 2. Testing for germline *SMAD4* and *BMPR1A* gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

G. STK11 Testing

- 1. Genetic testing for *STK11* gene variants may be considered **medically necessary** when any one of the following major criteria is met:
 - a. Individuals with a clinical diagnosis of Peutz-Jeghers syndrome based on the presence of any 2 of the following:
 - i. presence of 2 or more histologically confirmed Peutz-Jeghers polyps of the gastrointestinal tract.
 - ii. characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, or fingers
 - iii. family history of Peutz-Jeghers syndrome
 - b. At-risk relative of an individual suspected of or diagnosed with Peutz-Jeghers syndrome.
- 2. Testing for germline *STK11* gene variants for inherited CRC syndromes is considered **experimental / investigational** in all other situations.

H. Other Variants

1. Genetic testing of all other genes for an inherited CRC syndrome is considered **experimental / investigational**.

I. Genetic Counseling

1. Pre-and post-test genetic counseling may be considered **medically necessary** as an adjunct to the genetic testing itself.

POLICY GUIDELINES

A. <u>Testing At-Risk Relatives</u>

Due to the high lifetime risk of cancer of most genetic syndromes discussed in this policy, "at-risk relatives" primarily refers to first-degree relatives. However, some judgment must be permitted, e.g., in the case of a small family pedigree, when extended family members may need to be included in the testing strategy. Family history might include at least 2 second-degree relatives with a Lynch syndrome-related cancer, including at least 1 diagnosed before 50 years of age, or at least 3 second-degree relatives with a Lynch syndrome-related cancer, regardless of age.

B. <u>Targeted Familial Variant Testing</u>

It is recommended that, when possible, initial genetic testing for familial adenomatous polyposis (FAP) or Lynch syndrome be performed in an affected family member, so that testing in unaffected family members can focus on the variant found in the affected family member (see Benefit Application section). If an affected family member is not available for testing, testing should begin with an unaffected family member most closely related to an affected family member.

In many cases, genetic testing for MUTYH gene variants should first target the specific variants Y165C and G382D, which account for more than 80% of variants in white populations, and subsequently, proceed to sequence only as necessary. However, in other ethnic populations, proceeding directly to sequencing is appropriate.

C. Evaluation for Lynch Syndrome

For patients with colorectal cancer (CRC) or endometrial cancer being evaluated for Lynch syndrome, the microsatellite instability (MSI) test or the immunohistochemical (IHC) test with or without BRAF gene variant testing, or methylation testing, should be used as an initial evaluation of tumor tissue before mismatch repair (MMR) gene analysis. Both tests are not necessary. Proceeding to MMR gene sequencing would depend on the results of MSI or IHC testing. In particular, IHC testing may help direct which MMR gene likely contains a variant, if any, and may also provide additional information if MMR genetic testing is inconclusive. For further information on tumor tissue test results, interpretation, and additional testing options, see the NCCN [National Comprehensive Cancer Network] clinical care guidelines on genetic/familial high-risk assessment: colorectal.

When indicated, genetic sequencing for MMR gene variants should begin with MLH1 and MSH2 genes, unless otherwise directed by the results of IHC testing. Standard sequencing methods will not detect large deletions or duplications; when MMR gene variants are expected based on IHC or MSI studies, but none are found by standard sequencing, additional testing for large deletions or duplications is appropriate.

The Amsterdam II Clinical Criteria (all criteria must be fulfilled) are the most stringent criteria for defining families at high risk for Lynch syndrome:

- 1. 3 or more relatives with an associated cancer (colorectal cancer, or cancer of the endometrium, small intestine, ureter or renal pelvis);
- 2. 1 should be a first-degree relative of the other 2;
- 3. 2 or more successive generations affected;

- 4. 1 or more relatives diagnosed before the age of 50 years;
- 5. Familial adenomatous polyposis (FAP) should be excluded in cases of colorectal carcinoma;
- 6. Tumors should be verified by pathologic examination.
- 7. Modifications:
 - a. EITHER: very small families, which cannot be further expanded, can be considered to have HNPCC with only 2 colorectal cancers in first-degree relatives if at least 2 generations have the cancer and at least 1 case of colorectal cancer was diagnosed by the age of 55 years; OR
 - b. In families with 2 first-degree relatives affected by colorectal cancer, the presence of a third relative with an unusual early-onset neoplasm or endometrial cancer is sufficient.

The revised Bethesda Guidelines (fulfillment of any criterion meets guidelines) are less stringent than the Amsterdam criteria and are intended to increase the sensitivity of identifying at-risk families (Umar et al [2004]). The Bethesda guidelines are also considered more useful in identifying which patients with colorectal cancer should have their tumors tested for microsatellite instability and/or immunohistochemistry:

- 1. Colorectal carcinoma (CRC) diagnosed in a patient who is less than 50 years old;
- 2. Presence of synchronous (at the same time) or metachronous (at another time, i.e., a recurrence of) CRC or other HNPCC*—associated tumors, regardless of age;
- 3. CRC with high microsatellite instability histology diagnosed in a patient less than 60 years old;
- CRC diagnosed in 1 or more first-degree relatives with a Lynch syndromeassociated tumor, with one of the cancers being diagnosed before 50 years of age;
- 5. CRC diagnosed in 2 or more first- or second-degree relatives with HNPCC-related tumors, a regardless of age.
- ^a HNPCC-related tumors include colorectal, endometrial, stomach, ovarian, pancreas, ureter and renal pelvis, biliary tract, brain (usually glioblastoma as seen in Turcot syndrome), sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel.
- * HNPCC-related tumors include colorectal, endometrial, stomach, ovarian, pancreas, ureter and renal pelvis, biliary tract, brain (usually glioblastoma as seen in Turcot syndrome), sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel.

Multiple risk prediction models that provide quantitative estimates of the likelihood of an MMR variant are available such MMRpro, PREMM5 or MMRpredict. National Comprehensive Cancer Network guidelines recommend (category 2A) testing for Lynch syndrome in individuals with a 5% or higher predicted risk of the syndrome on these risk prediction models.

D. Genetic Counseling

Genetic counseling is primarily aimed at patients who are at risk for inherited disorders, and experts recommend formal genetic counseling in most cases when genetic testing for an inherited condition is considered. The interpretation of the results of genetic tests and the understanding of risk factors can be very difficult and complex. Therefore, genetic counseling will assist individuals in understanding the possible benefits and harms of genetic

testing, including the possible impact of the information on the individual's family. Genetic counseling may alter the utilization of genetic testing substantially and may reduce inappropriate testing. Genetic counseling should be performed by an individual with experience and expertise in genetic medicine and genetic testing methods.

Please refer to the member's contract benefits in effect at the time of service to determine coverage or non-coverage of these services as it applies to an individual member.

RATIONALE

This evidence review was created using searches of the PubMed database. The most recent literature review was performed through August 5 2025.

Evidence reviews assess whether a medical test is clinically useful. A useful test provides information to make a clinical management decision that improves the net health outcome. That is, the balance of benefits and harms is better when the test is used to manage the condition than when another test or no test is used to manage the condition.

The first step in assessing a medical test is to formulate the clinical context and purpose of the test. The test must be technically reliable, clinically valid, and clinically useful for that purpose. Evidence reviews assess the evidence on whether a test is clinically valid and clinically useful. Technical reliability is outside the scope of these reviews, and credible information on technical reliability is available from other sources.

Genetic Testing for Familial Adenomatous Polyposis and *MUTYH*-Associated Polyposis Clinical Context and Test Purpose

The purpose of genetic testing for familial adenomatous polyposis (FAP) and *MUTYH*-associated polyposis (MAP) is to

- Identify at-risk relatives of individuals with FAP and/or a known adenomatous polyposis coli (*APC*) gene variant.
- Make a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome.

The following PICO was used to select literature to inform this review.

Populations

The relevant population of interest is at-risk relatives of individuals with FAP and/or a known *APC* variant or those who require a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome.

Interventions

The relevant intervention is genetic testing for *APC or MUTYH*. Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing FAP and MAP: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be the early detection of colorectal cancer (CRC) and appropriate and timely interventional strategies (eg, endoscopic resection, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse events from that treatment or undertreatment.

Genetic testing for FAP may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual having or developing FAP.

Study Selection Criteria

For the evaluation of the clinical validity of the genetic test, studies that meet the following eligibility criterion were considered:

Reported on the analytic sensitivity and specificity and/or diagnostic yield of the test.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

Review of Evidence

The evidence review for FAP genetic testing was initially informed by a TEC Assessment (1998).^{33,} Additional information on attenuated FAP and on MAP diagnostic criteria and genetic testing is based on several publications that build on prior, cited research.^{34,35,36,37,}

Clinical sensitivity for classic FAP is about 95%; about 90% of pathogenic variants are detected by sequencing, ^{38,39}, while 8% to 12% of pathogenic variants are detected by deletion and duplication testing. ^{40,41}, Among Northern European whites, 98% of pathogenic *MUTYH* variants are detected by full gene sequencing. ^{42,43},

A comprehensive review of the *APC* pathogenic variant and its association with classical FAP and attenuated FAP and MAP is beyond the scope of this evidence review. The likelihood of detecting an *APC* pathogenic variant is highly dependent on the severity of colonic polyposis^{40,44,45,46,} and family history.^{47,} Detection rates are higher in classic polyposis (88%) than in nonclassical FAPs such as attenuated colonic phenotypes (57%) or MAP (33%).

Clinically Useful

A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, more effective therapy, or avoid unnecessary therapy or testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from randomized controlled trials (RCTs).

No RCTs were identified assessing the clinical utility of genetic testing for FAP and MAP.

Chain of Evidence

Genetic testing of patients requiring a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome may have clinical utility:

• If the test supports the clinical diagnosis of an attenuated disease, the protocol for endoscopic surveillance is affected and, depending on the situation, may avoid more frequent but unnecessary surveillance or necessitates more frequent surveillance.

Genetic testing of at-risk relatives of patients with FAP and/or a known APC variant may have clinical utility:

- If, in the absence of genetic testing, the diagnosis of colorectal polyposis in at-risk
 relatives of patients with FAP and/or a known APC variant can only be established by
 colonoscopy and subsequent histologic examination of removed polyps, which are
 burdensome.
- If results are negative, the test results may provide release from the intensified screening program resulting in psychological relief.

A TEC Assessment (1998)³³, offered the following conclusions:

- Genetic testing for FAP may improve health outcomes by identifying which currently unaffected at-risk family members require intense surveillance or prophylactic colectomy.
- At-risk subjects are considered to be those with greater than 10 adenomatous polyps or close relatives of patients with clinically diagnosed FAP or of patients with an identified APC variant.
- The optimal testing strategy is to define the specific genetic variant in an affected family member and then test the unaffected family members to see if they have inherited the same variant.

Testing for the *APC* variant has no role in the evaluation, diagnosis, or treatment of patients with classical FAP where the diagnosis and treatment are based on the clinical presentation.

Section Summary: Genetic Testing for Familial Adenomatous Polyposis and *MUTYH*-Associated Polyposis

The analytic and clinical sensitivity and specificity for *APC* and *MUTYH* are high. About 90% of pathogenic variants in classical FAP are detected by sequencing while 8% to 12% of pathogenic variants are detected by deletion and duplication testing. Among Northern European whites, 98% of pathogenic *MUTYH* variants are detected by full gene sequencing. The likelihood of detecting an *APC* pathogenic variant is highly dependent on the severity of colonic polyposis and family history. Detection rates are higher in classic polyposis (88%) than in nonclassical FAPs such as attenuated colonic phenotypes (57%) or MAP (33%). Direct evidence of clinical utility for genetic testing of attenuated FAP is not available. Genetic testing of at-risk relatives of patients with FAP and/or a known *APC* variant or those requiring a differential diagnosis of attenuated FAP versus

MAP versus Lynch syndrome may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from an intensified screening program resulting in psychological relief, and improving health outcomes by identifying currently unaffected at-risk family members who require intense surveillance or prophylactic colectomy.

LYNCH SYNDROME AND COLORECTAL CANCER GENETIC TESTING

Clinical Context and Test Purpose

The purpose of genetic testing for Lynch syndrome is to:

- Detect Lynch syndrome in individuals diagnosed with CRC or endometrial cancer,
- Identify at-risk relatives of individuals with a diagnosed Lynch syndrome and/or a known mismatch repair (MMR) variant and/or positive family history meeting Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a risk prediction model,
- Make a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome.

The following PICO was used to select literature to inform this review.

Populations

The relevant populations of interest are individuals diagnosed with CRC or endometrial cancer or at-risk relatives of patients with a diagnosed Lynch syndrome and/or a known MMR variant and/or positive family history meeting Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a risk prediction model, or those requiring a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome.

Interventions

The relevant intervention is genetic testing for the *MLH1*, *MSH2*, *MSH6*, *PMS2*, *EPCAM*, and/or *BRAF* V600E genes. Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing Lynch syndrome: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be early detection of Lynch syndrome and appropriate and timely interventional strategies (eg, increased surveillance, endoscopic resection, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse effects from that treatment or undertreatment.

Genetic testing for Lynch syndrome may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual having or developing Lynch syndrome.

Study Selection Criteria

For the evaluation of the clinical validity of the genetic test, studies that met the following eligibility criterion were considered:

• Reported on the analytic sensitivity and specificity and/or diagnostic yield of the test.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

REVIEW OF EVIDENCE

MMR Genes

Microsatellite instability (MSI) and immunohistochemical (IHC) screening tests for MMR variants have similar sensitivity and specificity. Microsatellite instability screening has a sensitivity of about 89% for *MLH1* and *MSH2* and 77% for *MSH6* and a specificity of about 90% for all. IHC screening has sensitivity for *MLH1*, *MSH2*, and *MSH6* of about 83% and a specificity of about 90% for each.

The evidence for Lynch syndrome genetic testing in patients with CRC is based on an evidence report conducted for the Agency for Healthcare Research and Quality by Bonis et al (2007),^{48,} a supplemental assessment to that report contracted by the Evaluation of Genomic Applications in Practice and Prevention (EGAPP) Working Group (2009),^{9,} and an EGAPP recommendation (2009) for genetic testing in CRC.^{49,} Based on the Agency for Healthcare Research and Quality report and supplemental assessment, the EGAPP recommendation concluded the following about genetic testing for MMR variants in patients already diagnosed with CRC:

- Family history, while important information to elicit and consider in each case, has poor sensitivity and specificity as a screening test to determine who should be considered for MMR variant testing and should not be used as a sole determinant or screening test.
- Optional BRAF testing can be used to reduce the number of patients, who are negative for MLH1 expression by IHC, needing MLH1 gene sequencing, thus improving efficiency without reducing sensitivity for MMR variants.

Vos et al (2020) evaluated the yield to detect Lynch syndrome in a prospective cohort of 3602 newly diagnosed CRC cases below age 70.^{50,} The standard testing protocol included IHC or MSI testing, followed by *MLH1* hypermethylation testing. Testing identified *MLH1* hypermethylation in a majority of cases tested (66% of 264). The percentage of MMR deficient CRC explained by hypermethylation increased with age, while the percentage of patients with hereditary CCR decreased with age. Of the 47 patients who underwent genetic testing, 55% (26/47) were determined to have Lynch syndrome. The authors estimated that only 78% of these cases would have been identified by the revised Bethesda guidelines. The percentage by age was 86% (6/7) in those under 40 years, 57% (17/29) in patients aged 40 to 64 years, and 30% (3/10) in patients 65 to 69 years of age and the number needed to test to identify 1 case of Lynch syndrome after prescreening was 1.2 (95% confidence interval [CI], 1.0 to 2.0) in patients under 40 years, 4.1 (95% CI, 3.1 to 5.5) in patients 40 to 64 years of age, and 21 (95% CI, 11 to 43) in CRC patients aged 65 to 69.

Tsuruta et al (2022) performed IHC screening for MMR-related genes (MLH1, MSH2, MSH6, and PMS2) to determine the extent to which Lynch syndrome can be diagnosed in patients with endometrial cancer through universal screening.^{51,} Samples were obtained from 100 patients, and 19 patients with lost results for any of the proteins were identified. The MSI-high phenotype was identified in 16 of 19 patients and MLH1 methylation was identified in 11 of 19 patients. The following were also detected: 2 pathological variants (MSH2 and MSH6), 2 cases of unclassified variant (MSH6), and 1 case of benign variant (PMS2).

EPCAM Testing

Several studies have characterized *EPCAM* deletions, established their correlation with the presence of *EPCAM-MSH2* fusion messenger RNAs (apparently nonfunctional) and with the presence of *MSH2* promoter hypermethylation, and, most importantly, have shown the cosegregation of these *EPCAM* variants with Lynch-like disease in families. ^{11,52,53,54,55,56}, Because studies differ slightly in how patients were selected, the prevalence of these *EPCAM* variants is difficult to estimate but may be in the range of 20% to 40% of patients/families who meet Lynch syndrome criteria, do not have an MMR variant, but have MSI-high tumor tissue. Kempers et al (2011) reported that carriers of an *EPCAM* deletion had a 75% (95% CI, 65% to 85%) cumulative risk of CRC by age 70 years, which did not differ significantly from that of carriers of an *MSH2* deletion (77%; 95% CI, 64% to 90%). The mean age at diagnosis was 43 years. ⁵⁷, However, the cumulative risk of endometrial cancer was low at 12% (95% CI, 0% to 27%) by age 70 compared with carriers of an *MSH2* variant (51%; 95% CI, 33% to 69%; p<.001).

BRAF V600 or MLH1 Promoter Methylation

Jin et al (2013) evaluated MMR proteins in 412 newly diagnosed CRC patients.^{58,} MLH1 and PMS2 protein stains were absent in 65 patients who were subsequently tested for a *BRAF* variant. Thirty-six (55%) of the 65 patients had the *BRAF* V600E variant, thus eliminating the need for further genetic testing or counseling for Lynch syndrome. Capper et al (2013) reported on a technique of V600E IHC testing for *BRAF* variants on a series of 91 stratified as high MSI CRC patients.^{59,} V600E positive lesions were detected in 21% of *MLH1*-negative CRC patients who could be excluded from MMR germline testing for Lynch syndrome. Therefore, V600E IHC testing for *BRAF* could be an alternative to *MLH1* promoter methylation analysis. To summarize, *BRAF* V600E variant or *MLH1* promoter methylation testing are optional screening methods that may be used when IHC testing shows a loss of MLH1 protein expression. The presence of *BRAF* V600E or absence of MLH1 protein expression due to *MLH1* promoter methylation rarely occurs in Lynch syndrome and would eliminate the need for further germline variant analysis for a Lynch syndrome diagnosis.^{60,}

Clinically Useful

A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, more effective therapy, or avoid unnecessary therapy or testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from RCTs.

No RCTs were identified assessing the clinical utility of genetic testing for Lynch syndrome.

Chain of Evidence

Genetic testing of patients with colon or endometrial cancer to detect Lynch syndrome has clinical utility:

 To make decisions about the preferred approach for treatment (endoscopic resection, colectomy with ileorectal anastomosis or segmental colectomy).

Genetic testing of at-risk relatives of patients with Lynch syndrome and/or a known MMR variant and/or positive family history meeting Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a risk prediction model, has clinical utility:

- If the individuals diagnosed with Lynch syndrome are recommended for screening for Lynch syndrome-associated cancers.
- If, in the absence of genetic testing, the diagnosis of Lynch syndrome in at-risk relatives of patients can only be established by colonoscopy and subsequent histologic examination of excised polyps, which is burdensome.
- If negative test results in prompt release from an intensified screening program, thereby reducing an emotional burden.

Genetic testing of patients requiring a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome may have clinical utility:

• If the test supports the clinical diagnosis of Lynch syndrome, the protocol for endoscopic surveillance is affected and, depending on the situation, may avoid more frequent but unnecessary surveillance or necessitates more frequent surveillance.

A chain of evidence can be constructed for the clinical utility of testing all patients with CRC for MMR variants. EGAPP conclusions are summarized next.

- Seven studies examined how counseling affected testing and surveillance choices among unaffected family members of Lynch syndrome patients. 61,62,63,64,65,66,67, About half of the relatives received counseling, and 95% of them chose MMR gene variant testing. Among those positive for MMR gene variants, uptake of colonoscopic surveillance beginning at age 20 to 25 years was high at 53% to 100%.
 - One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed recommended colonic surveillance versus those who did not.
 - Surveillance and prevention for other Lynch syndrome cancers.
- The chain of evidence from descriptive studies and expert opinion is inadequate (inconclusive) to demonstrate the clinical utility of testing the probands with Lynch syndrome (ie, the index patient).
 - Although a small body of evidence suggests that MSI-positive tumors are resistant to 5-fluorouracil and more sensitive to irinotecan than MSI-negative tumors, no alteration in therapy according to MSI status has yet been recommended.
 - Surveillance and prevention for other Lynch syndrome cancers:
 - While invasive and not actively recommended, women may choose hysterectomy with salpingo-oophorectomy to prevent gynecologic cancer.

In a retrospective study by Schmeler et al (2006), 315 women who chose this option had no gynecologic cancer over 10 years, whereas about one-third of women who did not have surgery developed endometrial cancer, and 5.5% developed ovarian cancer.^{68,}

- In a study by Bouzourene et al (2010), surveillance endometrial biopsy detected endometrial cancer and potentially precancerous conditions at earlier stages in those with Lynch syndrome, but results were not statistically significant, and a survival benefit has yet to be shown.^{10,} Transvaginal ultrasound is not a highly effective surveillance mechanism for endometrial cancer in patients with Lynch syndrome; however, transvaginal ultrasound in conjunction with endometrial biopsy has been recommended for surveillance.
- Gastroduodenoscopy for gastric cancer surveillance and urine cytology for urinary tract cancer surveillance are recommended based on expert opinion only, in the absence of adequate supporting evidence.

The Cancer Genetic Studies Consortium (1997) recommended that if CRC is diagnosed in patients with an identified variant or a strong family history, a subtotal colectomy with ileorectal anastomosis should be considered as an option for segmental resection.^{69,} The 2006 joint American Society of Clinical Oncology and Society of Surgical Oncology review assessing risk-reducing surgery in hereditary cancers recommended offering total colectomy plus ileorectal anastomosis or hemicolectomy as options to patients with Lynch syndrome and CRC, especially those who are younger.^{70,} The Societies' review also recommended offering Lynch syndrome patients with an index rectal cancer the options of total proctocolectomy with ileal pouch-anal anastomosis or anterior proctosigmoidectomy with primary reconstruction. The rationale for total proctocolectomy is the 17% to 45% rate of metachronous colon cancer in the remaining colon after an index rectal cancer in Lynch syndrome patients.

The risk of endometrial cancer in MMR variant carriers has been estimated at 34% (95% CI, 17% to 60%) by age 70, and at 8% for ovarian cancer (95% CI, 2% to 39%) by age 70.^{71,} Risks do not appear to appreciably increase until after age 40. Females with Lynch syndrome who choose risk-reducing surgery are encouraged to consider oophorectomy because of the risk of ovarian cancer in Lynch syndrome. In a retrospective cohort study, Obermair et al (2010) found that hysterectomy improved survival among female colon cancer survivors with Lynch syndrome-associated CRC, about 23 would be diagnosed with endometrial cancer within 10 years absent a hysterectomy. Surveillance in Lynch syndrome populations for ovarian cancer has not been demonstrated to be successful at improving survival.^{73,}

Section Summary: Lynch Syndrome and Colorectal Cancer Genetic Testing

Direct evidence of clinical utility for genetic testing for Lynch syndrome is not available. Multiple studies have demonstrated clinical utility in testing unaffected (without cancer) first- and second-degree relatives of patients with Lynch syndrome who have a known MMR variant, in that counseling has been shown to influence testing and surveillance choices among unaffected family members of Lynch syndrome patients. One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed and did not follow recommended colonic surveillance. A positive genetic test

for an MMR gene variant can also lead to changes in the management of other Lynch syndrome malignancies.

GENETIC TESTING FOR JUVENILE POLYPOSIS SYNDROME AND PEUTZ-JEGHERS SYNDROME

Clinical Context and Test Purpose

The purpose of genetic testing for Juvenile Polyposis syndrome (JPS) and Peutz-Jeghers syndrome (PJS) is:

- To confirm a diagnosis of JPS or PJS in individuals suspected of these disorders based on clinical features.
- To identify at-risk relatives of individuals with a confirmed diagnosis of JPS or PJS.

The following PICO was used to select literature to inform this review.

Populations

The relevant populations of interest are individuals with suspected JPS or PJS and individuals who are at-risk relatives of individuals suspected of or diagnosed with JPS or PJS.

Interventions

The relevant intervention is genetic testing for *SMAD4* and *BMPR1* (for JPS) and *STK11* (for PJS). Commercial testing is available from numerous companies.

Comparators

The following practice is currently being used to make decisions about managing JPS and PJS: no genetic testing.

Outcomes

The potential beneficial outcomes of primary interest would be early detection of cancer and appropriate and timely interventional strategies (eg, cancer screening, surgical intervention including polyp resection, gastrectomy, colectomy) to prolong life.

The potential harmful outcomes are those resulting from a false test result. False-positive or false-negative test results can lead to the initiation of unnecessary treatment and adverse events from that treatment or undertreatment.

Genetic testing for *SMAD4* and *BMPR1* (for JPS) and *STK11* (for PJS) may be performed at any point during a lifetime. The necessity for genetic testing is guided by the availability of information that alters the risk of an individual of having or developing JPS and PJS.

Study Selection Criteria

For the evaluation of the clinical validity of the genetic test, studies that met the following eligibility criterion were considered:

• Reported on the diagnostic yield of the test.

Clinically Valid

A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

Review of Evidence

Table 1 summarizes clinical validity studies assessing genetic testing for JPS and PJS.

Table 1. Summary of Clinical Validity Studies Assessing Genetic Testing for JPS and PJS

Study	Study Design and Population	Results
Calva-Cerqueira et al (2009) ^{74,}	Observational; 102 unrelated JPS probands analyzed all of whom met clinical criteria for JPS	SMAD4 and BMPR1A variants detected in 41% (42/102) JPS probands
Aretz et al (2007) ^{75,}	Observational; 80 unrelated patients (65 met clinical criteria for typical JPS; 15 presumed to have JPS) were examined by direct sequencing for <i>SMAD4</i> , <i>BMPR1A</i> , and <i>PTEN</i> variants	SMAD4 and BMPR1A variants detected in 60% of typical JPS patients and none in presumed JPS patients; overall diagnostic yield, 49%
Volikos et al (2006) ^{76,}	Observational; 76 clinically diagnosed with PJS	Detection rate of germline variants was about 80% (59/76)
Aretz et al (2005) ^{77,}	Observational; 71 patients (56 met clinical criteria for PJS; 12 presumed to have PJS)	STK11 variant detected in 52% (37/71)

JPS: juvenile polyposis syndrome; PJS: Peutz-Jeghers syndrome.

Clinical Useful

A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, more effective therapy, or avoid unnecessary therapy or testing.

Direct Evidence

Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from RCTs.

No RCTs were identified assessing the clinical utility of genetic testing for JPS and PJS.

Chain of Evidence

Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to demonstrate test performance, no inferences can be made about clinical utility.

Genetic testing of patients with suspected JPS and PJS has clinical utility:

 To make decisions about a preferred approach for treatment (endoscopic resection, colectomy with ileorectal anastomosis, segmental colectomy).

Genetic testing of individuals who are at-risk relatives of patients suspected of or diagnosed with JPS or PJS has clinical utility:

- If the individuals diagnosed with JPS and PJS are recommended for screening for JPS and PJS-associated cancers.
- If, in the absence of genetic testing, the diagnosis of JPS and PJS in at-risk relatives of
 patients can only be established by colonoscopy and subsequent histologic examination of
 excised polyps, which is burdensome.
- If negative test results in prompt release from an intensified screening program, thereby reducing an emotional burden.

A systematic review of 20 cohort studies with a total of 1644 patients with PJS was published by Lier et al (2010).^{32,} A total of 349 patients developed 384 malignancies at an average age of 42 years. The lifetime risk for any cancer varied between 37% and 93% with relative risks (RRs) ranging from 9.9 to 18 versus the general population.

Section Summary: Genetic Testing for Juvenile Polyposis Syndrome and Peutz-Jeghers Syndrome

The likelihood of detecting a pathogenic variant is highly dependent on the presence of clinical features and family history. Detection rates have been reported to be between 60% and 41% for JPS, and 52% and 80% for PJS. Direct evidence of the clinical utility for genetic testing of JPS or PJS is not available. Genetic testing of patients with suspected JPS or PJS or individuals who are at-risk relatives of patients suspected of or diagnosed with a polyposis syndrome or PJS may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from an intensified screening program resulting in psychological relief, and improving health outcomes by identifying currently unaffected at-risk family members who require intense surveillance or prophylactic colectomy.

SUPPLEMENTAL INFORMATION

The purpose of the following information is to provide reference material. Inclusion does not imply endorsement or alignment with the evidence review conclusions.

Clinical Input From Physician Specialty Societies and Academic Medical Centers While the various physician specialty societies and academic medical centers may collaborate with and make recommendations during this process, through the provision of appropriate reviewers, input received does not represent an endorsement or position statement by the physician specialty societies or academic medical centers, unless otherwise noted.

In response to requests, input was received from 3 physician specialty societies and 3 academic medical centers while this policy was under review in 2009. In general, those providing input agreed with the overall approach described in this policy.

Practice Guidelines and Position Statements

Guidelines or position statements will be considered for inclusion in 'Supplemental Information' if they were issued by, or jointly by, a US professional society, an international society with US representation, or National Institute for Health and Care Excellence (NICE). Priority will be given to guidelines that are informed by a systematic review, include strength of evidence ratings, and include a description of management of conflict of interest.

American College of Gastroenterology

The American College of Gastroenterology (2015) issued practice guidelines for the management of patients with hereditary gastrointestinal cancer syndromes.²¹,

For Lynch syndrome, the College recommended:

"All newly diagnosed colorectal cancers (CRCs) should be evaluated for mismatch repair [MMR] deficiency.

Analysis may be done by immunohistochemical [IHC] testing for the *MLH1/MSH2/MSH6/PMS2* proteins and/or testing for microsatellite instability [MSI]. Tumors that demonstrate loss of *MLH1* should undergo BRAF testing or analysis for *MLH1* promoter hypermethylation.

Individuals who have a personal history of a tumor showing evidence of MMR deficiency (and no demonstrated BRAF variant or hypermethylation of MLH1), a known family variant associated with LS [Lynch syndrome], or a risk of $\geq 5\%$ chance of LS based on risk prediction models should undergo genetic evaluation for LS.⁷⁸,

Genetic testing of patients with suspected LS should include germline variant genetic testing for the *MLH1*, *MSH2*, *MSH6*, *PMS2*, and/or *EPCAM* genes or the altered gene(s) indicated by IHC testing."

For adenomatous polyposis syndromes, the College recommended:

"Familial adenomatous polyposis (FAP)/MUTYH-associated polyposis/attenuated polyposis

Individuals who have a personal history of >10 cumulative colorectal adenomas, a family history of 1 one of the adenomatous polyposis syndromes, or a history of adenomas and FAP-type extracolonic manifestations (duodenal/ampullary adenomas, desmoid tumors, papillary thyroid cancer, congenital hypertrophy of the retinal pigment epithelium, epidermal cysts, osteomas) should undergo assessment for the adenomatous polyposis syndromes.

Genetic testing of patients with suspected adenomatous polyposis syndromes should include *APC* and *MUTYH* gene variant analysis."

For juvenile polyposis syndrome, the College recommended:

"Genetic evaluation of a patient with possible JPS [juvenile polyposis syndrome] should include testing for *SMAD4* and *BMPR1A* mutations"

"Surveillance of the gastrointestinal (GI) tract in affected or at-risk JPS patients should include screening for colon, stomach, and small bowel cancers (strong recommendation, very low quality of evidence).

Colectomy and ileorectal anastomosis or proctocolectomy and ileal pouch-anal anastomosis is indicated for polyp-related symptoms, or when the polyps cannot be managed endoscopically (strong recommendation, low quality of evidence).

Cardiovascular examination for and evaluation for hereditary hemorrhagic telangiectasia should be considered for *SMAD4* mutation carriers (conditional recommendation, very low quality of evidence)."

For Peutz-Jeghers syndrome, the College recommended:

"Genetic evaluation of a patient with possible PJS [Peutz-Jeghers syndrome] should include testing for *STK11* mutations."

"Surveillance in affected or at-risk PJS patients should include monitoring for colon, stomach, small bowel, pancreas, breast, ovary, uterus, cervix, and testes cancers. Risk for lung cancer is increased, but no specific screening has been recommended. It would seem wise to consider annual chest radiograph or chest computed tomography (CT) in smokers (conditional recommendation, low quality of evidence)."

American Society of Clinical Oncology and Society of Surgical Oncology

The American Society of Clinical Oncology (2015) concluded the European Society for Medical Oncology clinical guidelines published in 2013 were based on the most relevant scientific evidence and therefore endorsed them with minor qualifying statements (in bold italics).^{79,} The recommendations as related to genetic testing hereditary CRC syndromes are summarized below:

- "Tumor testing for DNA MMR deficiency with IHC for MMR proteins and/or MSI should be assessed in all CRC patients. As an alternate strategy, tumor testing should be carried out in individuals with CRC younger than 70 years, or those older than 70 years who fulfill any of the revised Bethesda guidelines.
- If loss of MLH1/PMS2 *protein expression* is observed in the tumor, analysis of *BRAF* V600E mutation or analysis of methylation of the *MLH1* promoter should be carried out first to rule out a sporadic case. *If tumor is MMR deficient and somatic* BRAF *mutation is not detected or MLH1 promoter methylation is not identified, testing for germline mutations is indicated*.
- If loss of any of the other proteins (MSH2, MSH6, PMS2) is observed, germline genetic
 testing should be carried out for the genes corresponding to the absent
 proteins (eg, MSH2, MSH6, EPCAM, PMS2, or MLH1).
- Full germline genetic testing *for Lynch syndrome* should include DNA sequencing and large rearrangement analysis.
- Patients with multiple colorectal adenomas should be considered for full germline genetic testing of APC and/or MUTYH.
- Germline testing of MUTYH can be initiated by screening for the most common mutations (G396D, Y179C) in the white population followed by analysis of the entire gene in heterozygotes. Founder mutations among ethnic groups should be taken into account. For nonwhite individuals, full sequencing of MUTYH should be considered."

National Comprehensive Cancer Network

The NCCN guidelines on genetic/familial high-risk assessment of colorectal cancer syndromes (v1.2025v2.2023) are summarized in Table 2.80,

Table 2. Criteria for Evaluation of Lynch Syndrome Based on Personal or Family History of Cancer

Criteria for the Evaluation of Lynch Syndrome

Known LS pathogenic variant in the family

An individual with a LS-related cancer and any of the following:

- Diagnosed <50 v
- Another synchronous or metachronous LS-related cancer^a regardless of age
- 1 first-degree or second-degree relative with LS-related a cancer diagnosed <50 y
- ≥2 first-degree or second-degree relatives with LS-related a cancers regardless of age

Personal history of a tumor with MMR deficiency determined by PCR, NGS, or IHC diagnosed at any age^b

- Family history (on the same side of the family) of any of the following:
- ≥1 first-degree relative with colorectal or endometrial cancer diagnosed <50 y
- ≥1 first-degree relative with colorectal or endometrial cancer and another synchronous or metachronous LS-related cancera
- ≥2 first-degree or second-degree relatives with LS-related cancer, a including ≥1 diagnosed <50 y
- ≥3 first-degree or second-degree relatives with LS-related cancers, a regardless of age

An individual with a \geq 5% risk of having an MMR gene pathogenic variant based on predictive models (ie, PREMM₅, MMRpro, MMRpredict)

- Individuals with a personal history of CRC and/or endometrial cancer with a PREMM5 score of ≥2.5% should be considered for MGPT.
- For individuals without a personal history of CRC and/or endometrial cancer, some data have suggested using a PREMM₅ score threshold of ≥2.5% rather than ≥5% to select individuals for MMR genetic testing. Based on these data, it is reasonable for testing to be done based on the ≥2.5% score result and clinical judgment. Of note, with the lower threshold, there is an increase in sensitivity, but a decrease in specificity.

CRC: colorectal cancer; IHC: immunohistochemisty; LS: Lynch syndrome; MGPT: multi-gene panel testing; MMR: mismatch repair; MSI: microsatellite instability; NGS: next generation sequencing; PCR: polymerase chain reaction.
^a LS-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, urothelial, brain (usually glioblastoma), biliary tract, and small intestinal cancers, as well as sebaceous carcinomas, and keratoacanthomas as seen in Muir-Torre syndrome.

^b The NCCN recommends tumor screening for MMR deficiency for all CRC and endometrial cancers regardless of age at diagnosis. Tumor screening for CRCs for MMR deficiency for purposes of screening for LS is not required if MGPT is chosen as the strategy for screening for LS, but may still be required for CRC therapy selection. Consider tumor screening for MMR deficiency for sebaceous neoplasms as well as the following adenocarcinomas: small bowel, ovarian, gastric, pancreas, biliary tract, brain, bladder, urothelial, and adrenocortical cancers regardless of age at diagnosis. Direct referral for germline testing to rule out LS may be preferred in patients with a strong family history or if diagnosed prior to age 50 y, MSI-H, or loss of MMR protein expression. For patients aged ≥50 at CRC diagnosis, the panel has also recommended to consider germline MGPT evaluation for LS and other hereditary cancer syndromes.

Genetic Testing Recommendations for Lynch Syndrome

Screening of the tumor for defective DNA MMR using IHC and/or MSI is used to identify which patients should undergo mutation testing for Lynch syndrome.^{81,} The NCCN guidelines also indicate that BRAF V600E testing or *MLH1* promoter methylation testing may be used

when *MLH1* is not expressed in the tumor on IHC analysis to exclude a diagnosis of Lynch syndrome.

The NCCN guidelines for colon cancer (v4.2024) recommend that all newly diagnosed patients with colon cancer be tested for MMR or MSI.²⁶,

The NCCN guidelines for uterine neoplasm (v2.2024)also recommend universal screening for MMR genes (and MSI testing if results are equivocal).^{27,} Additionally, the NCCN guidelines recommend screening for Lynch syndrome in all endometrial cancer patients younger than 50 years of age.

The NCCN guidelines for genetic/familial high-risk assessment: colorectal (v2.2023) recommend genetic testing for at-risk family members of patients with positive variants in *MLH1*, *MSH2*, *MSH6*, *PMS2*, and *EPCAM*.⁸⁰, These guidelines also address familial adenomatous polyposis (classical and attenuated) and *MUTYH*-associated polyposis and are consistent with the information provided in this evidence review.

Surveillance Recommendations for Lynch Syndrome

The NCCN guidelines for colon cancer (v4.2024) 26 , and for colorectal cancer (CRC) screening (v1.2024) 81 , recommend CRC patients treated with curative-intent surgery undergo surveillance colonoscopy at 1 year postsurgery and, if normal, again in 3 years, then every 5 years based on findings.

The NCCN guidelines on genetic/familial high-risk assessment for CRC indicate for MLH1, MSH2, and EPCAM variant carriers that surveillance with colonoscopy should begin "at age 20 to 25 years or 2 to 5 years before the earliest colon cancer if it is diagnosed before age 25 years and repeat every 1 to 2 years."⁸⁰,

MSH6 and PMS2 variant carriers should begin surveillance with colonoscopy "at age 30 to 35 years or 2 to 5 years before the earliest colon cancer if it is diagnosed before age 30 years and repeat every 1 to 3 years". 80 ,

Peutz-Jeghers Syndrome and Juvenile Polyposis Syndrome

There are limited data on the efficacy of various screening modalities in juvenile polyposis syndrome (JPS) and Peutz-Jeghers syndrome (PJS). The NCCN cancer risk and surveillance 2 category 2A recommendations for these indications are summarized in Tables 3 and 4.80,

Table 3. Risk and Surveillance Guidelines for Peutz-Jeghers Syndrome

Site	Lifetime Risk, %		Approximate Initiation Age, y
Breast	32 to 54	Mammogram and breast MRI annuallyClinical breast exam every 6 mo	30 y
Colon	39	Colonoscopy every 2 to 3 y; shorter intervals may be indicated based on polyp size, number, and pathology	18 y
Stomach	29	Upper endoscopy every 2 to 3 y; shorter intervals may be indicated based on polyp size, number, and pathology	18 y
Small intestine	13	Small bowel visualization (CT or MRI enterography or video capsule endoscopy) every 2 to 3 y; shorter intervals may be indicated based on polyp size, number, and pathology	18 y
Pancreas	11 to 36	Annual imaging of the pancreas with either EUS or MRI/MRCP (both ideally performed at center of expertise)	30 to 35 y ^a
Cervix (typically minimal deviation adenocarcinoma)	≥10	 Pelvic examination and Pap smear annually Consider total hysterectomy (including uterus and cervix) once completed with childbearing 	18 to 20 y
Uterus	9	Annual pelvic examination with endometrial biopsy if abnormal bleeding	18 to 20 y
Ovary (sex cord tumor with annular tubules)	≥20	Annual pelvic examination with annual pelvic ultrasound	18 to 20 y
Lung	7 to 17	 Provide education about symptoms and smoking cessation No other specific recommendations have been made 	•
Testes (Sertoli cell tumors)	9	Annual testicular exam and observation for feminizing changes	Continued from pediatric screening

CT: computed tomography; EUS: endoscopic ultrasound; MR: magnetic resonance; MRCP: Magnetic resonance cholangiopancreatography; MRI: magnetic resonance imaging.

^aBased on clinical judgment, early initiation age may be considered, such as 10 y younger than the earliest age of onset in the family.

Table 4. Pediatric and Adult Risk and Surveillance Guidelines for Juvenile Polyposis

Syndrome

Site	Lifetime Risk, % for <i>SMAD4/BMPR1A</i> variants	Screening Procedure and Interval	Approximate Initiation Age, y
Colon	up to 50	Adults: Colonoscopy every 1–3 years. Intervals should be based on polyp size, number, and pathology ^a Pediatrics: Colonoscopy every 2–3 years. Intervals should be based on polyp size, number, and pathology ^a	Adults: 18 y Pediatric: 12- 15 y
Stomach	up to 21, especially if multiple gastric polyps present	Adults:Upper endoscopy every 1–3 years. Intervals should be based on polyp size, number, and pathology. ^{a,b} Pediatrics: Upper endoscopy and polypectomy every 2–3 years. Intervals should be based on polyp size, number, and pathology ^a	Adults: 18 y Pediatric: 12- 15 y
Small intestine	Rare, undefined	No recommendations made	
ННТ	22	In individuals with <i>SMAD4</i> variants, screen for vascular lesions associated with HHT	Within first 6 mo of life, or at time of diagnosis

HHT: hereditary hemorrhagic telangectasia.

U.S. Preventive Services Task Force Recommendations

No U.S. Preventive Services Task Force recommendations for genetic testing of Lynch syndrome and other inherited colon cancer syndromes have been identified.

Ongoing and Unpublished Clinical Trials

Some currently ongoing and unpublished trials that might influence this review are listed in Table 5.

^a If polyp burden or polyp-related symptoms (ie, anemia) cannot be controlled endoscopically or prevent optimal surveillance for cancer, consideration should be given to gastrectomy and/or colectomy.

^b While SMAD4 pathogenic variant carriers often have severe upper gastrointestinal tract involvement, BMRP1A pathogenic variant carriers may have a less severe upper gastrointestinal tract phenotype and may merit lengthened surveillance intervals in the absence of polyps. Gastric cancer risk for BMPR1A pathogenic variant carriers may be lower than for SMAD4 pathogenic variant carriers

Table 5. Summary of Key Trials

NCT No.	Trial Name	Planned Enrollment	Completion Date
Ongoing			
NCT02494791	Universal Screening for Lynch Syndrome in Women With Endometrial and Non-Serous Ovarian Cancer	886	July 2025 (status unknown)
NCT04494945	Approaches to Identify and Care for Individuals With Inherited Cancer Syndromes	27500	Jun 2030
NCT06582914	Lynch Syndrome Integrative Epidemiology and Genetics (LINEAGE)	5000	Dec 2054
NCT06501417	EC_ItaLynch: Incorporating Lynch Syndrome Genetic Testing in Standard Medical Care of Patients With Endometrial Cancer (Mainstreaming)	600	Dec 2028
NCT06772844	DNA Methylation Analysis in Stool Samples for Screening of LynchSyndrome-Associated Colorectal Cancer	400	Dec 2028
NCT06863038	Predictive Value of the PREMM5, MMRpredict Models, and the Universal Tumor Screening Strategy for Lynch Syndrome in Vietnam	572	Nov 2027
NCT06989814	Smart Measurement of Circulating Tumor DNA: a Tumoragnostic Computational Tool to Improve Colorectal Cancer Care	50	Feb 2027

NCT: national clinical trial.

CODING

The following codes for treatment and procedures applicable to this policy are included below for informational purposes. This may not be a comprehensive list of procedure codes applicable to this policy.

Inclusion or exclusion of a procedure, diagnosis or device code(s) does not constitute or imply member coverage or provider reimbursement. Please refer to the member's contract benefits in effect at the time of service to determine coverage or non-coverage of these services as it applies to an individual member.

The code(s) listed below are medically necessary ONLY if the procedure is performed according to the "Policy" section of this document.

CPT/HC	CPT/HCPCS		
81201	APC (adenomatous polyposis coli) (e.g., familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; full gene sequence		
81202	APC (adenomatous polyposis coli) (e.g., familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; known familial variants		
81203	APC (adenomatous polyposis coli) (e.g., familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; duplication/deletion variants		
81210	BRAF (rB-Raf proto-oncogene, serine/threonine kinase) (e.g., colon cancer, melanoma), gene analysis, V600 variant(s)		
81288	MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; promoter methylation analysis		
81292	MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis		
81293	MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non- polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants		
81294	MLH1(mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non- polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication; deletion variants		
81295	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis		
81296	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variant		
81297	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (e.g., hereditary non- polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication / deletion variant		
81298	MSH6 (mutS homolog 6 [E. coli]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis		
81299	MSH6 (mutS homolog 6 [E. coli]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants		
81300	MSH6 (mutS homolog 6 [E. coli]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication / deletion variants		

CPT/HC	PCS
81301	Microsatellite instability analysis (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) of markers for mismatch repair deficiency (e.g., BAT25, BAT26), includes comparison of neoplastic and normal tissue, if performed
81317	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis
81318	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants
81319	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants
81403	Molecular pathology procedure, Level 4 (e.g., analysis of single exon by DNA sequence analysis, analysis of >10 amplicons using multiplex PCR in 2 or more independent reactions, mutation scanning or duplication/deletion variants of 2-5 exons)
81435	Hereditary colon cancer-related disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), genomic sequence analysis panel, 5 or more genes, interrogation for sequence variants and copy number variants
0101U	Hereditary colon cancer disorders (e.g., Lynch syndrome, <i>PTEN</i> hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA, and array CGH, with MRNA analytics to resolve variants of unknown significance when indicated (15 genes [sequencing and deletion/duplication], <i>EPCAM</i> and <i>GREM1</i> [deletion/duplication only])
0130U	Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), targeted mRNA sequence analysis panel (APC, CDH1, CHEK2, MLH1, MSH2, MSH6, MUTYH, PMS2, PTEN, and TP53) (List separately in addition to code for primary procedure), (new eff 10/01/2019- panel including many genes discussed in this policy;
0157U	APC (APC regulator of WNT signaling pathway) (e.g., familial adenomatosis polyposis [FAP]) mRNA sequence analysis (List separately in addition to code for primary procedure)
0158U	MLH1 (mutL homolog 1) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)
0159U	MSH2 (mutS homolog 2) (e.g., hereditary colon cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)
0160U	MSH6 (mutS homolog 6) (e.g., hereditary colon cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)
0161U	PMS2 (PMS1 homolog 2, mismatch repair system component) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)

CPT/HCP	CPT/HCPCS	
0162U	Hereditary colon cancer (Lynch syndrome), targeted mRNA sequence analysis panel (MLH1, MSH2, MSH6, PMS2) (List separately in addition to code for primary procedure)	
0238U	Oncology (Lynch syndrome), genomic DNA sequence analysis of MLH1, MSH2, MSH6, PMS2, and EPCAM, including small sequence changes in exonic and intronic regions, deletions, duplications, mobile element insertions, and variants in non-uniquely mappable regions	

REVISIONS	
05-13-2011	Policy added to the bcbsks.com web site.
01-01-2012	In the Coding section:
	Added the new codes: 81210, 81292-81301
04-10-2012	 In the Coding section: Replaced Diagnosis code 183.1 with correct code 183.2. Removed HCPCS codes: S3828, S3829, S3830, S3831 (Deleted codes, effective April 1, 2012.)
01-15-2013	In the Coding section: Added CPT codes: 81401, 81406 Added new CPT codes: 81201, 81202, 81203(Effective 01-01-2013) Removed CPT codes:83890, 83892, 83898, 83902, 83904, 83905, 83906, 83912 (Effective 12-31-2012)
03-26-2013	Updated Description section.
	 In Policy section: In Item I, Note, Amsterdam II Criteria, added "6. Modifications: EITHER: very small families, which cannot be further expanded, can be considered to have HNPCC with only 2 colorectal cancers in first-degree relatives if at least two generations have the cancer and at least one case of colorectal cancer was diagnosed by the age of 55 years; OR: in families with two first-degree relatives affected by colorectal cancer, the presence of a third relative with an unusual early-onset neoplasm or endometrial cancer is sufficient." In Item I, Note, Revised Bethesda Criteria, added "6. Colorectal cancer diagnosed with one or more first-degree relatives with HNPCC-related tumor (colorectal, endometrial, stomach, ovarian, pancreas, bladder, ureter and renal pelvis, biliary tract, brain [usually glioblastoma as seen in Turcot syndrome], sebaceous bland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel), with one of the cancers being diagnosed under age 50 years, OR colorectal cancer diagnosed in two or more first-or second-degree relatives with HNPCC related tumor, regardless of age. (15)" Updated Rationale section.
	Updated Reference section.
08-21-2013	In Coding section:
00 21 2013	Removed CPT code 81210.
	Added ICD-10 Diagnosis codes (Effective October 1, 2014)
01-01-2015	Policy posted 01-16-2015
	In Coding section:
	Added CPT Code: 81288 (Effective January 1, 2015)
03-18-2015	In Title section:

REVISIONS	
	Changed title name from "Genetic Testing for Inherited Susceptibility to Colon
	Cancer, Including Microsatellite Instability Testing"
	Updated Description section.
	In Policy section:
	 Removed Amsterdam II criteria and Revised Bethesda guidelines.
	■ In Policy Guidelines, added items 6-9.
	Updated Rationale section.
	In Coding section:
	 Added CPT Codes 81210, 81317, 81318, 81319, and 81403.
	Removed CPT Code 81406.
	Updated References section.
01-01-2016	In Coding section:
	Revised nomenclature to CPT codes: 81210 and 81401.
02-03-2016	Updated Description section.
	In Policy section:
	 Added statement on genetic counseling to Policy Guidelines.
	Updated Rationale section.
	Updated References section.
	Added Appendix section.
05-25-2016	Updated Description section.
03 23 2010	In Policy section:
	Revised Policy Guideline Item 6.
	Updated Rationale section.
	Updated References section.
11-09-2016	In Policy section:
11 05 2010	 In Item I B, removed "when feasible" and "who meet the revised Bethesda criteria
	(see Policy Guidelines below)" and added "or immunohistochemical (IHC) analysis of
	tumors" and "or endometrial" to read, "Microsatellite instability (MSI) testing or
	immunohistochemical (IHC) analysis of tumors may be considered medically
	necessary as an initial test in persons with colorectal or endometrial cancer in order
	to identify those persons who should proceed with HNPCC variant analysis."
	In Coding section:
	Added CPT codes: 88341, 88342, 88344.
12-08-2017	Updated Description section.
	In Policy section:
	Added new Item I B, "HNPCC genetic testing is considered experimental /
	investigational for all other indications."
	Previous Item I B is now Item I C.
	 Added new Item I D, "MSI testing or IHC analysis of tumors is considered
	experimental / investigational for all other indications."
	 Added new Item II B, "APC genetic testing is considered experimental /
	investigational for all other indications."
	Previous Item II B is now Item II C.
	 Added new Item II D, "MAP genetic testing is considered experimental /
	investigational for all other indications."
	Updated Rationale section.
	In Coding section:
	Removed ICD-9 codes.
	Updated References section.
02-18-2019	Updated Description section.

 $\textit{Current Procedural Terminology} \ \textcircled{O} \ \text{American Medical Association}. \ \text{All Rights Reserved}.$ Blue Cross and Blue Shield Kansas is an independent licensee of the Blue Cross Blue Shield Association

REVISIONS

In Policy section:

- Removed the previous policy language: "I. Lynch syndrome (also known as Hereditary Non-Polyposis Colorectal Cancer [HNPCC]): A. Genetic testing for HNPCC (MLH1, MSH2, MSH6, PMS2 sequence analysis) is considered medically necessary when one of the following criteria are met: 1. Meets Amsterdam II criteria or revised Bethesda guidelines (see Policy Guidelines below); or 2. A first-* or second-degree** relative with an HNPCC variant (genes MLH1, MSH2, MSH6, PMS2); or 3. Endometrial cancer 50 years of age or younger. B. HNPCC genetic testing is considered experimental / investigational for all other indications. C. Microsatellite instability (MSI) testing or immunohistochemical (IHC) analysis of tumors may be considered medically necessary as an initial test in persons with colorectal or endometrial cancer in order to identify those persons who should proceed with HNPCC variant analysis. D. MSI testing or IHC analysis of tumors is considered experimental / investigational for all other indications. II. Familial Adenomatous Polyposis and associated variance: A. Adenosis polyposis coli (APC) genetic testing is considered medically necessary for either of the following indications: 1. Greater than 10 colonic polyps; or 2. First-degree* relatives diagnosed with familial adenomatous polyposis (FAP) or with a documented APC variant. The specific APC variant should be identified in the affected first-degree relative with FAP prior to testing the member, if feasible. Full sequence APC genetic testing is considered medically necessary only when it is not possible to determine the family variant first. B. APC genetic testing is considered experimental / investigational for all other indications. C. Testing for MYH variants is considered medically necessary for any of the following indications: 1. Personal history of 10 to 20 adenomatous polyposis who have negative APC variant testing and a negative family history for adenomatous polyposis; OR 2. Personal history of 10 to 20 adenomatous polyposis whose family history is consistent with recessive inheritance (i.e., family history is positive only for sibling[s]); OR 3. Asymptomatic siblings of individuals with known MYH polyposis. D. MAP genetic testing is considered experimental / investigational for all other indications. *First-degree relatives are parents, siblings, and offspring. ** Seconddegree relatives are aunts, uncles, grandparents, niece, nephews or half-siblings. ▲ Hereditary nonpolyposis colorectal cancer (HNPCC)-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, ureter and renal pelvis, brain (usually glioblastoma as seen in Turcot syndrome), and small intestinal cancers, as well as sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome."
- Added new policy language, "A. MMR Gene Testing 1. Genetic testing for MMR genes (MLH1, MSH2, MSH6, PMS2) may be considered medically necessary in the following patients: a) Patients with colorectal cancer (CRC), for the diagnosis of Lynch syndrome (see Policy Guidelines. b) Patients with endometrial cancer and a first-degree relative diagnosed with a Lynch-associated cancer (see Policy Guidelines), for the diagnosis of Lynch syndrome. c) At-risk relatives (see Policy Guidelines) of patients with Lynch syndrome with a known MMR gene variant. d) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome. Whether testing begins with APC variants or screening for MMR genes depends on clinical presentation. e) Patients without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, when no affected family members have been tested for MMR variants. B. APC Testing 1. Genetic testing for adenosis polyposis coli (APC) may be considered medically necessary in the following patients: a) At-risk relatives (see Policy Guidelines) of patients with familial adenomatous polyposis (FAP) and/or a known APC variant. b) Patients with a differential diagnosis

REVISIONS

of attenuated FAP vs MUTYH-associated polyposis (MAP) vs Lynch syndrome. Whether testing begins with APC variants or screening for mismatch repair (MMR) variants depends on clinical presentation. 2. Genetic testing for APC gene variants is not medically necessary for colorectal cancer patients with classical FAP for confirmation of the FAP diagnosis. C. MUTYH Testing 1. Testing for *MUTYH* gene variants may be considered medically necessary in the following patients: a) Patients with a differential diagnosis of attenuated FAP vs MAP vs Lynch syndrome and a negative result for APC gene variants. A family history of no parents or children with FAP is consistent with MAP (autosomal recessive). D. EPCAM Testing 1. Genetic testing for *EPCAM* gene variants may be considered medically necessary when any one of the following 3 major criteria (solid bullets) is met: 1) Patients with CRC, for the diagnosis of Lynch syndrome (see Policy Guidelines section) when: i. Tumor tissue shows lack of MSH2 protein expression by immunohistochemistry and patient is negative for a MSH2 germline variant; OR ii. Tumor tissue shows a high level of microsatellite instability and patient is negative for a germline variant in MLH1, MSH2, MSH6, and PMS2; OR b) At-risk relatives (see Policy Guidelines section) of patients with Lynch syndrome with a known EPCAM variant; OR c) Patients without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, when no affected family members have been tested for MMR variants, and when sequencing for MMR variants is negative. E. BRAF V600E or MLH1 promoter methylation 1. Genetic testing for BRAF V600E or MLH1 promoter methylation may be considered medically necessary to exclude a diagnosis of Lynch syndrome when the MLH1 protein is not expressed in a CRC tumor on immunohistochemical analysis. F. SMAD4 and BMPR1A Testing 1. Genetic testing for SMAD4 and BMPR1A gene variants may be considered medically necessary when any one of the following major criteria (solid bullets) is met: a) Patients with a clinical diagnosis of juvenile polyposis syndrome based on the presence of any one of the following: i. at least 3 to 5 juvenile polyps in the colon ii. multiple juvenile polyps in other parts of the gastrointestinal tract iii. any number of juvenile polyps in a person with a known family history of juvenile polyps. b) At-risk relative of a patient suspected of or diagnosed with juvenile polyposis syndrome. G. STK11 Testing 1. Genetic testing for STK11 gene variants may be considered medically necessary when any one of the following major criteria (solid bullets) is met: a) Patients with a clinical diagnosis of Peutz-Jeghers syndrome based on the presence of any 2 of the following: i. presence of 2 or more histologically confirmed Peutz-Jeghers polyps of the small intestine ii. characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, or fingers iii. family history of Peutz-Jeghers syndrome b) At-risk relative of a patient suspected of or diagnosed with Peutz-Jeahers syndrome, H. Genetic testing for all other gene variants for Lynch syndrome or CRC is considered experimental / investigational."

Updated Policy Guidelines.

Updated Rationale section.

In Coding section:

- Removed CPT codes: 81401, 88341, 88342, 88344.
- Updated coding bullets.

Updated References section.

Removed Appendix section.

04-24-2019

In Policy section:

REVISIONS	
1(201010	■ In Item A 1 e, removed "or Revised Bethesda" to read, "Patients without CRC but
	with a family history meeting the Amsterdam criteria, when no affected family members have been tested for MMR variants."
	In Item D 1 c, removed "or Revised Bethesda" and "and when sequencing for MMR
	variants is negative" to read, "Patients without CRC but with a family history meeting
	the Amsterdam criteria, when no affected family members have been tested for
07.04.0040	MMR variants."
07-01-2019	In Coding section:
07.00.2020	Added new CPT code: 0101U Published 07-09-2020. Effective 06-15-2020
07-09-2020	
	Updated Description section Updated Coding section-
	Removed: CPT 81403
	A LL LODT 04405 04405 0400U
	 Added CP1: 81435, 81436, 01300 Removed ICD 10: C17.0, C17.1,C17.2, C17.3, C17.8,C17.9, , C25.1, C25.2,
	C56.1, C56.2, C56.9, C57.00, C579.01, C57.02, C57.10, C57.11, C57.12, C57.3,
	C5.20, C57.21, C57.22, C60.1, C71.0, C71.1, C71.2, C71.3, C71.4, C71.5, C71.6,
	C717.7, C71.8, C71.9
	 Added ICD 10: D01.3, D01.4, D01.5, D01.6, D01.7, D01.8, D01.9, Q85.8, Z31.5
	Updated Rationale section
	Updated Reference section
05-21-2021	Updated Description section
	In Policy Section:
	Added the following:
	o In Item A.1.e and Item D.1.c, "or documentation of 5% or higher predicted risk of
	the syndrome on a validated risk prediction model (e.g. MMRpro, PREMM5 or MMRpredict),"
	o In Item C.1, "Genetic"
	o In policy guideline 1, "A family history might include at least 2 second-degree
	relatives with a Lynch syndrome-related cancer, including at least 1 diagnosed
	before 50 years of age, or at least 3 second-degree relatives with a Lynch
	syndrome-related cancer, regardless of age."
	 In policy guideline C., "* HNPCC-related tumors include colorectal, endometrial,
	stomach, ovarian, pancreas, ureter and renal pelvis, biliary tract, brain (usually
	glioblastoma as seen in Turcot syndrome), sebaceous gland adenomas and
	keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel.
	Multiple risk prediction models that provide quantitative estimates of the likelihood
	of an MMR variant are available such MMRpro, PREMM5 (Kastrinos et al [2017]), or
	MMRpredict. National Comprehensive Cancer Network guidelines recommend
	(category 2A) testing for Lynch syndrome in individuals with a 5% or higher
	predicted risk of the syndrome on these risk prediction models."
	Replaced the following: In Them 5.1.4.1. "3 to" with "5".
	o In Item F.1.A.1, "3 to" with "5"
	o In Item F.1.A.2, "in other parts of" with "found throughout"
	 In Item G.1.A.1, "small intestine" with "gastrointestinal tract" Policy quideline A, "majority of the" with "most"
	 Policy guideline A, "majority of the" with "most" Removed the following:
	To Thomas Educad Cd Weelid hollate)
	 In Items F.1 and G.1, "(Solid bullets) In policy guidelines removed the genetic nomenclature update section and the
	following:
	ronowing.

REVISIONS

A. Several Clinical Laboratory Improvement Amendments (CLIA)–licensed clinical laboratories offer MMR gene variant testing for Lynch syndrome. For example, the GeneTests website (available online at:

http://www.ncbi.nlm.nih.gov/sites/GeneTests/lab/clinical_disease_id/2622?db) lists 32 U.S.-located laboratories that offer this service. In at least 1 laboratory, Lynch syndrome variant testing is packaged under 1 copyrighted name. The COLARIS® test from Myriad Genetic Laboratories includes sequence analysis of MLH1, MSH2, MSH6, and PMS2; large rearrangement analysis for MLH1, MSH2, PMS2, and MSH6 large deletions/duplications; and analysis for large deletions in the EPCAM gene near MSH2. Note that there may be 2 versions of this test, the COLARIS (excludes PMS2 testing) and COLARIS Update (includes PMS2 testing). Individualized testing (e.g., targeted testing for a family variant) can also be requested. The COLARISPLUS test includes full sequence analysis of MLH1, MSH2, MSH6, PMS2, and MYH genes and rearrangement analysis of MLH1, MSH2, MSH6, MYH, and EPCAM by microarray comparative genomic hybridization analysis, and multiplex ligation-dependent probe amplification analysis for PMS2.

B. Similarly, GeneTests lists 15 U.S.-based CLIA-licensed clinical laboratories that provide APC variant testing and 14 that provide MUTYH variant testing. The COLARIS® AP test from Myriad Genetic Laboratories includes DNA sequencing analysis of the APC and MUTYH genes, as well as analysis of large rearrangements in the APC gene that are not detected by DNA sequencing.

Updated Rationale section

In Coding section:

- Added Codes: 0157U, 0158U, 0159U, 0160U, 0161U, 0162U, 0238U
- Added ICD-10 diagnosis code Z85.040
- Removed ICD-10 diagnosis code D01.8

Updated References section

12-05-2021

Updated Description Section

Updated Policy Section

- A1a deleted "for the diagnosis" and replaced with "with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for"
- A1b Changed to read "Patients with endometrial cancer with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for Lynch syndrome (see Policy Guidelines section)"
- Added section A2 Testing for germline MMR gene variants for inherited CRC syndromes is considered experimental / investigational in all other situations
- Added section B4 Testing for germline APC gene variants for inherited CRC syndromes is considered experimental / investigational in all other situations.
- Added section C2 Testing for germline MUTYH gene variants for inherited CRC syndromes is considered experimental / investigational in all other situations
- Added section D2 added Testing for germline EPCAM gene variants for inherited CRC syndromes is considered experimental / investigational in all other situations.
- Added section E2 Testing for somatic BRAF V600E or MLH1 promoter methylation to exclude a diagnosis of Lynch syndrome is considered experimental / investigational in all other situations.
- Added section F2 Testing for germline SMAD4 and BMPR1A gene variants for inherited CRC syndromes is considered experimental / investigational in all other situations

Updated Rationale Section

REVISIONS	
	Updated Reference Section
12-09-2022	Updated Description Section
12 03 2022	Updated Policy Section
	 Section A1c added "pathogenic/likely pathogenic" to statement. Now reads: "At-
	risk relatives (see Policy Guidelines) of individuals with Lynch syndrome with a
	known pathogenic/likely pathogenic MMR gene variant."
	 Section D1b added "pathogenic/likely pathogenic" to statement. Now reads:
	"At-risk relatives (see Policy Guidelines section) of individuals with Lynch
	syndrome with a known pathogenic/likely pathogenic <i>EPCAM</i> variant; OR"
	 Section D1c added "and when sequencing for MMR variants is negative." to
	statement. Now reads: "Individuals without CRC but with a family history
	meeting the Amsterdam criteria, or documentation of 5% or higher predicted
	risk of the syndrome on a validated risk prediction model (e.g. MMRpro,
	PREMM5 or MMRpredict), when no affected family members have been tested
	for MMR variants, and when sequencing for MMR variants is negative."
	 Section E1 added "somatic" in front of Genetic testing for BRAF V600E
	 Section F1 removed "variants" after genes. Now reads: "Genetic testing for
	SMAD4 and BMPR1A genes may be considered medically necessary when any
	one of the following major criteria is met:"
	Updated Policy Guideline Section
	 Updated Genetic Counseling Section to read: "Genetic counseling is primarily
	aimed at patients who are at risk for inherited disorders, and experts
	recommend formal genetic counseling in most cases when genetic testing for an
	inherited condition is considered. The interpretation of the results of genetic
	tests and the understanding of risk factors can be very difficult and complex.
	Therefore, genetic counseling will assist individuals in understanding the possible
	benefits and harms of genetic testing, including the possible impact of the
	information on the individual's family. Genetic counseling may alter the
	utilization of genetic testing substantially and may reduce inappropriate testing.
	Genetic counseling should be performed by an individual with experience and
	expertise in genetic medicine and genetic testing methods."
	Updated Rationale Section
	Updated Coding Section
	 Converted ICD-10 codes to ranges
	Updated References Section
10-24-2023	Updated Description Section
	Updated Rationale Section
	Updated Coding Section
	 Removed ICD-10 codes
	Updated References Section
11-20-2024	Updated Description Section
	Updated Rationale Section
	Updated References Section
01-01-2025	Updated Coding Section
	Revised nomenclature for 81435 (eff. 01-01-2025)
	The vised from chalacter for of 155 (cm of of 2025)
	Removed deleted code 81436 (eff. 01-01-2025)
11-26-2025	
11-26-2025	Removed deleted code 81436 (eff. 01-01-2025)

REFERENCES

- Vogt S, Jones N, Christian D, et al. Expanded extracolonic tumor spectrum in MUTYHassociated polyposis. Gastroenterology. Dec 2009; 137(6): 1976-85.e1-10. PMID 19732775
- 2. Balmaña J, Castells A, Cervantes A. Familial colorectal cancer risk: ESMO Clinical Practice Guidelines. Ann Oncol. May 2010; 21 Suppl 5: v78-81. PMID 20555108
- 3. Gala M, Chung DC. Hereditary colon cancer syndromes. Semin Oncol. Aug 2011; 38(4): 490-9. PMID 21810508
- 4. Quehenberger F, Vasen HF, van Houwelingen HC. Risk of colorectal and endometrial cancer for carriers of mutations of the hMLH1 and hMSH2 gene: correction for ascertainment. J Med Genet. Jun 2005; 42(6): 491-6. PMID 15937084
- 5. Guindalini RS, Win AK, Gulden C, et al. Mutation spectrum and risk of colorectal cancer in African American families with Lynch syndrome. Gastroenterology. Nov 2015; 149(6): 1446-53. PMID 26248088
- 6. Sinn DH, Chang DK, Kim YH, et al. Effectiveness of each Bethesda marker in defining microsatellite instability when screening for Lynch syndrome. Hepatogastroenterology. 2009; 56(91-92): 672-6. PMID 19621678
- 7. Wu Y, Berends MJ, Mensink RG, et al. Association of hereditary nonpolyposis colorectal cancer-related tumors displaying low microsatellite instability with MSH6 germline mutations. Am J Hum Genet. Nov 1999; 65(5): 1291-8. PMID 10521294
- 8. Goel A, Nagasaka T, Spiegel J, et al. Low frequency of Lynch syndrome among young patients with non-familial colorectal cancer. Clin Gastroenterol Hepatol. Nov 2010; 8(11): 966-71. PMID 20655395
- 9. Palomaki GE, McClain MR, Melillo S, et al. EGAPP supplementary evidence review: DNA testing strategies aimed at reducing morbidity and mortality from Lynch syndrome. Genet Med. Jan 2009; 11(1): 42-65. PMID 19125127
- 10. Bouzourene H, Hutter P, Losi L, et al. Selection of patients with germline MLH1 mutated Lynch syndrome by determination of MLH1 methylation and BRAF mutation. Fam Cancer. Jun 2010; 9(2): 167-72. PMID 19949877
- 11. Niessen RC, Hofstra RM, Westers H, et al. Germline hypermethylation of MLH1 and EPCAM deletions are a frequent cause of Lynch syndrome. Genes Chromosomes Cancer. Aug 2009; 48(8): 737-44. PMID 19455606
- 12. Hesson LB, Hitchins MP, Ward RL. Epimutations and cancer predisposition: importance and mechanisms. Curr Opin Genet Dev. Jun 2010; 20(3): 290-8. PMID 20359882
- 13. Hitchins MP. Inheritance of epigenetic aberrations (constitutional epimutations) in cancer susceptibility. Adv Genet. 2010; 70: 201-43. PMID 20920750
- 14. Vasen HF, Watson P, Mecklin JP, et al. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. Gastroenterology. Jun 1999; 116(6): 1453-6. PMID 10348829
- 15. Umar A, Boland CR, Terdiman JP, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. J Natl Cancer Inst. Feb 18 2004; 96(4): 261-8. PMID 14970275
- 16. Kastrinos F, Uno H, Ukaegbu C, et al. Development and Validation of the PREMM 5 Model for Comprehensive Risk Assessment of Lynch Syndrome. J Clin Oncol. Jul 01 2017; 35(19): 2165-2172. PMID 28489507

- 17. Latchford AR, Neale K, Phillips RK, et al. Juvenile polyposis syndrome: a study of genotype, phenotype, and long-term outcome. Dis Colon Rectum. Oct 2012; 55(10): 1038-43. PMID 22965402
- 18. Howe JR, Roth S, Ringold JC, et al. Mutations in the SMAD4/DPC4 gene in juvenile polyposis. Science. May 15 1998; 280(5366): 1086-8. PMID 9582123
- 19. Fogt F, Brown CA, Badizadegan K, et al. Low prevalence of loss of heterozygosity and SMAD4 mutations in sporadic and familial juvenile polyposis syndrome-associated juvenile polyps. Am J Gastroenterol. Oct 2004; 99(10): 2025-31. PMID 15447767
- Burger B, Uhlhaas S, Mangold E, et al. Novel de novo mutation of MADH4/SMAD4 in a patient with juvenile polyposis. Am J Med Genet. Jul 01 2002; 110(3): 289-91. PMID 12116240
- 21. Syngal S, Brand RE, Church JM, et al. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol. Feb 2015; 110(2): 223-62; quiz 263. PMID 25645574
- 22. Grotsky HW, Rickert RR, Smith WD, et al. Familial juvenile polyposis coli. A clinical and pathologic study of a large kindred. Gastroenterology. Mar 1982; 82(3): 494-501. PMID 7054044
- 23. Schreibman IR, Baker M, Amos C, et al. The hamartomatous polyposis syndromes: a clinical and molecular review. Am J Gastroenterol. Feb 2005; 100(2): 476-90. PMID 15667510
- 24. Brosens LA, van Hattem A, Hylind LM, et al. Risk of colorectal cancer in juvenile polyposis. Gut. Jul 2007; 56(7): 965-7. PMID 17303595
- 25. Gallione CJ, Repetto GM, Legius E, et al. A combined syndrome of juvenile polyposis and hereditary haemorrhagic telangiectasia associated with mutations in MADH4 (SMAD4). Lancet. Mar 13 2004; 363(9412): 852-9. PMID 15031030
- 26. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colon Cancer. Version 4.2025. http://www.nccn.org/professionals/physician_gls/pdf/colon.pdf. Accessed August 05, 2025
- 27. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Uterine Neoplasms. Version 3.2025. https://www.nccn.org/professionals/physician_gls/pdf/uterine.pdf Accessed August 8, 2025
- 28. Olschwang S, Markie D, Seal S, et al. Peutz-Jeghers disease: most, but not all, families are compatible with linkage to 19p13.3. J Med Genet. Jan 1998; 35(1): 42-4. PMID 9475093
- 29. Jenne DE, Reimann H, Nezu J, et al. Peutz-Jeghers syndrome is caused by mutations in a novel serine threonine kinase. Nat Genet. Jan 1998; 18(1): 38-43. PMID 9425897
- 30. Hemminki A, Markie D, Tomlinson I, et al. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. Nature. Jan 08 1998; 391(6663): 184-7. PMID 9428765
- 31. Hernan I, Roig I, Martin B, et al. De novo germline mutation in the serine-threonine kinase STK11/LKB1 gene associated with Peutz-Jeghers syndrome. Clin Genet. Jul 2004; 66(1): 58-62. PMID 15200509
- 32. van Lier MG, Wagner A, Mathus-Vliegen EM, et al. High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. Am J Gastroenterol. Jun 2010; 105(6): 1258-64; author reply 1265. PMID 20051941

- 33. Blue Cross and Blue Shield Association Technology Evaluation Center (TEC). Genetic Testing for Inherited Susceptibility to Colorectal Cancer: Part I Adenomatous Polyposis Coli Gene Mutations. TEC Assessments. 1998; Volume 13:Tab 10. PMID
- 34. Kastrinos F, Syngal S. Recently identified colon cancer predispositions: MYH and MSH6 mutations. Semin Oncol. Oct 2007; 34(5): 418-24. PMID 17920897
- 35. Lefevre JH, Parc Y, Svrcek M, et al. APC, MYH, and the correlation genotype-phenotype in colorectal polyposis. Ann Surg Oncol. Apr 2009; 16(4): 871-7. PMID 19169759
- 36. Avezzù A, Agostini M, Pucciarelli S, et al. The role of MYH gene in genetic predisposition to colorectal cancer: another piece of the puzzle. Cancer Lett. Sep 18 2008; 268(2): 308-13. PMID 18495334
- 37. Balaguer F, Castellví-Bel S, Castells A, et al. Identification of MYH mutation carriers in colorectal cancer: a multicenter, case-control, population-based study. Clin Gastroenterol Hepatol. Mar 2007; 5(3): 379-87. PMID 17368238
- 38. Jasperson KW, Patel SG, Ahnen DJ. APC-Associated Polyposis Conditions. In: Pagon RA, Adam MP, Ardinger HH, et al., eds. GeneReviews. Seattle, WA: University of Washington; 2017.
- 39. Lagarde A, Rouleau E, Ferrari A, et al. Germline APC mutation spectrum derived from 863 genomic variations identified through a 15-year medical genetics service to French patients with FAP. J Med Genet. Oct 2010; 47(10): 721-2. PMID 20685668
- 40. Aretz S, Stienen D, Uhlhaas S, et al. Large submicroscopic genomic APC deletions are a common cause of typical familial adenomatous polyposis. J Med Genet. Feb 2005; 42(2): 185-92. PMID 15689459
- 41. Bunyan DJ, Eccles DM, Sillibourne J, et al. Dosage analysis of cancer predisposition genes by multiplex ligation-dependent probe amplification. Br J Cancer. Sep 13 2004; 91(6): 1155-9. PMID 15475941
- 42. Out AA, Tops CM, Nielsen M, et al. Leiden Open Variation Database of the MUTYH gene. Hum Mutat. Nov 2010; 31(11): 1205-15. PMID 20725929
- 43. Nielsen M, Lynch H, Infante E, et al. MUTYH-Associated Polyposis. In: Pagon RA, Adam MP, Ardinger HH, eds. GeneReviews Seattle, WA: University of Washington; 2012.
- 44. Sieber OM, Lamlum H, Crabtree MD, et al. Whole-gene APC deletions cause classical familial adenomatous polyposis, but not attenuated polyposis or "multiple" colorectal adenomas. Proc Natl Acad Sci U S A. Mar 05 2002; 99(5): 2954-8. PMID 11867715
- 45. Aretz S, Uhlhaas S, Goergens H, et al. MUTYH-associated polyposis: 70 of 71 patients with biallelic mutations present with an attenuated or atypical phenotype. Int J Cancer. Aug 15 2006; 119(4): 807-14. PMID 16557584
- 46. Michils G, Tejpar S, Thoelen R, et al. Large deletions of the APC gene in 15% of mutationnegative patients with classical polyposis (FAP): a Belgian study. Hum Mutat. Feb 2005; 25(2): 125-34. PMID 15643602
- 47. Truta B, Allen BA, Conrad PG, et al. A comparison of the phenotype and genotype in adenomatous polyposis patients with and without a family history. Fam Cancer. 2005; 4(2): 127-33. PMID 15951963
- 48. Bonis PA, Trikalinos TA, Chung M, et al. Hereditary Nonpolyposis Colorectal Cancer: Diagnostic Strategies and Their Implications (Evidence Report/Technology Assessment No. 150). Rockville, MD: Agency for Healthcare Research and Quality; 2007.
- 49. Berg AO, Armstrong K, Botkin J, et al. Recommendations from the EGAPP Working Group: genetic testing strategies in newly diagnosed individuals with colorectal cancer aimed at

- reducing morbidity and mortality from Lynch syndrome in relatives. Genet Med. Jan 2009; 11(1): 35-41. PMID 19125126
- 50. Vos JR, Fakkert IE, Spruijt L, et al. Evaluation of yield and experiences of age-related molecular investigation for heritable and nonheritable causes of mismatch repair deficient colorectal cancer to identify Lynch syndrome. Int J Cancer. Oct 15 2020; 147(8): 2150-2158. PMID 32510614
- 51. Tsuruta T, Todo Y, Yamada R, et al. Initial screening by immunohistochemistry is effective in universal screening for Lynch syndrome in endometrial cancer patients: a prospective observational study. Jpn J Clin Oncol. Jul 08 2022; 52(7): 752-758. PMID 35438162
- 52. Kloor M, Voigt AY, Schackert HK, et al. Analysis of EPCAM protein expression in diagnostics of Lynch syndrome. J Clin Oncol. Jan 10 2011; 29(2): 223-7. PMID 21115857
- 53. Kuiper RP, Vissers LE, Venkatachalam R, et al. Recurrence and variability of germline EPCAM deletions in Lynch syndrome. Hum Mutat. Apr 2011; 32(4): 407-14. PMID 21309036
- 54. Kovacs ME, Papp J, Szentirmay Z, et al. Deletions removing the last exon of TACSTD1 constitute a distinct class of mutations predisposing to Lynch syndrome. Hum Mutat. Feb 2009; 30(2): 197-203. PMID 19177550
- 55. Ligtenberg MJ, Kuiper RP, Chan TL, et al. Heritable somatic methylation and inactivation of MSH2 in families with Lynch syndrome due to deletion of the 3' exons of TACSTD1. Nat Genet. Jan 2009; 41(1): 112-7. PMID 19098912
- 56. Rumilla K, Schowalter KV, Lindor NM, et al. Frequency of deletions of EPCAM (TACSTD1) in MSH2-associated Lynch syndrome cases. J Mol Diagn. Jan 2011; 13(1): 93-9. PMID 21227399
- 57. Kempers MJ, Kuiper RP, Ockeloen CW, et al. Risk of colorectal and endometrial cancers in EPCAM deletion-positive Lynch syndrome: a cohort study. Lancet Oncol. Jan 2011; 12(1): 49-55. PMID 21145788
- 58. Jin M, Hampel H, Zhou X, et al. BRAF V600E mutation analysis simplifies the testing algorithm for Lynch syndrome. Am J Clin Pathol. Aug 2013; 140(2): 177-83. PMID 23897252
- 59. Capper D, Voigt A, Bozukova G, et al. BRAF V600E-specific immunohistochemistry for the exclusion of Lynch syndrome in MSI-H colorectal cancer. Int J Cancer. Oct 01 2013; 133(7): 1624-30. PMID 23553055
- 60. Kastrinos F, Syngal S. Screening patients with colorectal cancer for Lynch syndrome: what are we waiting for?. J Clin Oncol. Apr 01 2012; 30(10): 1024-7. PMID 22355054
- 61. Hampel H, Frankel WL, Martin E, et al. Screening for the Lynch syndrome (hereditary nonpolyposis colorectal cancer). N Engl J Med. May 05 2005; 352(18): 1851-60. PMID 15872200
- 62. Aktan-Collan K, Mecklin JP, Järvinen H, et al. Predictive genetic testing for hereditary non-polyposis colorectal cancer: uptake and long-term satisfaction. Int J Cancer. Jan 20 2000; 89(1): 44-50. PMID 10719730
- 63. Aktan-Collan K, Haukkala A, Pylvänäinen K, et al. Direct contact in inviting high-risk members of hereditary colon cancer families to genetic counselling and DNA testing. J Med Genet. Nov 2007; 44(11): 732-8. PMID 17630403
- 64. Stanley AJ, Gaff CL, Aittomäki AK, et al. Value of predictive genetic testing in management of hereditary non-polyposis colorectal cancer (HNPCC). Med J Aust. Apr 03 2000; 172(7): 313-6. PMID 10844916

- 65. Hadley DW, Jenkins J, Dimond E, et al. Genetic counseling and testing in families with hereditary nonpolyposis colorectal cancer. Arch Intern Med. Mar 10 2003; 163(5): 573-82. PMID 12622604
- 66. Lerman C, Hughes C, Trock BJ, et al. Genetic testing in families with hereditary nonpolyposis colon cancer. JAMA. May 05 1999; 281(17): 1618-22. PMID 10235155
- 67. Codori AM, Petersen GM, Miglioretti DL, et al. Attitudes toward colon cancer gene testing: factors predicting test uptake. Cancer Epidemiol Biomarkers Prev. Apr 1999; 8(4 Pt 2): 345-51. PMID 10207639
- 68. Schmeler KM, Lynch HT, Chen LM, et al. Prophylactic surgery to reduce the risk of gynecologic cancers in the Lynch syndrome. N Engl J Med. Jan 19 2006; 354(3): 261-9. PMID 16421367
- 69. Burke W, Petersen G, Lynch P, et al. Recommendations for follow-up care of individuals with an inherited predisposition to cancer. I. Hereditary nonpolyposis colon cancer. Cancer Genetics Studies Consortium. JAMA. Mar 19 1997; 277(11): 915-9. PMID 9062331
- 70. Guillem JG, Wood WC, Moley JF, et al. ASCO/SSO review of current role of risk-reducing surgery in common hereditary cancer syndromes. J Clin Oncol. Oct 01 2006; 24(28): 4642-60. PMID 17008706
- 71. Bonadona V, Bonaïti B, Olschwang S, et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. JAMA. Jun 08 2011; 305(22): 2304-10. PMID 21642682
- 72. Obermair A, Youlden DR, Young JP, et al. Risk of endometrial cancer for women diagnosed with HNPCC-related colorectal carcinoma. Int J Cancer. Dec 01 2010; 127(11): 2678-84. PMID 20533284
- 73. Auranen A, Joutsiniemi T. A systematic review of gynecological cancer surveillance in women belonging to hereditary nonpolyposis colorectal cancer (Lynch syndrome) families. Acta Obstet Gynecol Scand. May 2011; 90(5): 437-44. PMID 21306348
- 74. Calva-Cerqueira D, Chinnathambi S, Pechman B, et al. The rate of germline mutations and large deletions of SMAD4 and BMPR1A in juvenile polyposis. Clin Genet. Jan 2009; 75(1): 79-85. PMID 18823382
- 75. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic deletions and a genotype phenotype update in 80 unrelated families with juvenile polyposis syndrome. J Med Genet. Nov 2007; 44(11): 702-9. PMID 17873119
- 76. Volikos E, Robinson J, Aittomäki K, et al. LKB1 exonic and whole gene deletions are a common cause of Peutz-Jeghers syndrome. J Med Genet. May 2006; 43(5): e18. PMID 16648371
- 77. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic STK11 deletions in Peutz-Jeghers syndrome. Hum Mutat. Dec 2005; 26(6): 513-9. PMID 16287113
- 78. Kastrinos F, Steyerberg EW, Mercado R, et al. The PREMM(1,2,6) model predicts risk of MLH1, MSH2, and MSH6 germline mutations based on cancer history. Gastroenterology. Jan 2011; 140(1): 73-81. PMID 20727894
- 79. Stoffel EM, Mangu PB, Gruber SB, et al. Hereditary colorectal cancer syndromes: American Society of Clinical Oncology Clinical Practice Guideline endorsement of the familial risk-colorectal cancer: European Society for Medical Oncology Clinical Practice Guidelines. J Clin Oncol. Jan 10 2015; 33(2): 209-17. PMID 25452455
- 80. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High-Risk Assessment: Colorectal. Version 1.2025.

- http://www.nccn.org/professionals/physician_gls/pdf/genetics_colon.pdf. Accessed August 07, 2025.
- 81. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colorectal Cancer Screening. Version 2. 2025. http://www.nccn.org/professionals/physician_gls/pdf/colorectal_screening.pdf. Accessed August 06, 2025.

OTHER REFERENCES

1. Blue Cross and Blue Shield of Kansas Surgery Liaison, August 2010; August 2011; August 2018; January 2019; February 2022.